

Manchester Microscopical & Natural History Society



Established 1880

www.manchestermicroscopical.org.uk

Measurement in Microscopy

Micrometry ... a part of the Microscopist's Armamentarium

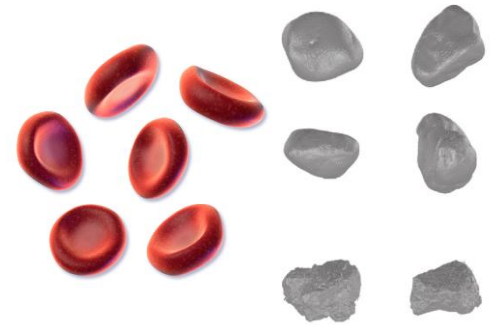


Guess !

Use experience



Use a ruler/transfer scale



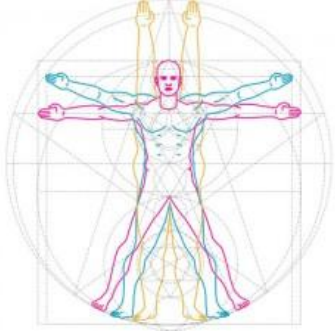
Use a known object

25 years ago! ... MMS: November 2000

Measuring Microscopical Objects

How to measure sizes, surfaces, volumes, lengths, numbers, shapes and patterns of microscopical objects. + Practical

Mike Mahon, February 22nd 2025



See MMS Website
(Downloads pdf) ...

Quantitation



- Sampling
- Morphometry
- Stereology
- Pattern Analysis
- Image Analysis

Questions



- Why measure ?
- What do you want to measure ?
- How do we measure ?
- Are the results unbiased, precise, accurate, valid, meaningful ?

Questions

- Why measure ?
- What do you want to measure ?
- How do we measure ?
- Are the results unbiased, precise, accurate, valid, meaningful ?

Why measure ?

“ When you can **measure** what you are speaking about and express it in numbers, you know something about it; but when you cannot express it in numbers, your knowledge is of a meagre and unsatisfactory kind; it may be the beginning of knowledge, but you have scarcely in your thoughts advanced to the state of **Science**, whatever the matter may be.”

Lord Kelvin (1883)

MMS How big / small is it?

WHAT'S THE POINT OF
ATTACHING A NUMBER
TO EVERYTHING
YOU DO?



IF YOUR NUMBERS
GO UP, IT MEANS
YOU'RE HAVING
MORE FUN.

SCIENCE TO
THE SPIRIT'S
RESCUE
ONCE AGAIN.



Why measure ?

- **Obtain Absolute Size Data**
- **Variability / Constancy**
- **Relative / Comparative Data**
- Experimental v Control Data
- Disease v Healthy Data
- Treatment v Control Data
- Data on Changes / Growth / Ageing / Differences
- Data on Structure / Function Relationships
- Data to Predict / Mathematical Models

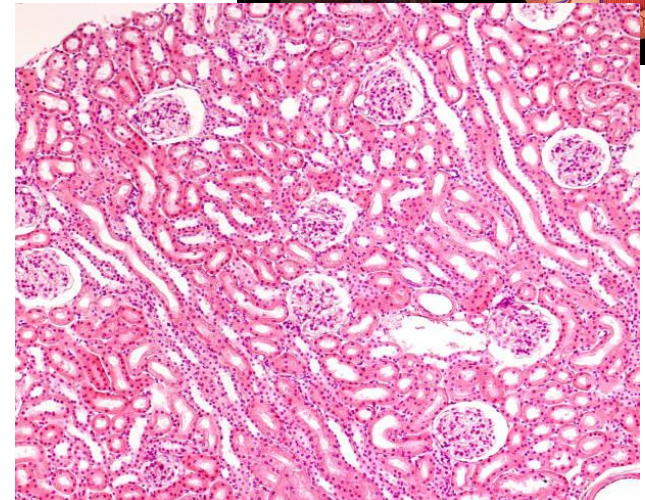
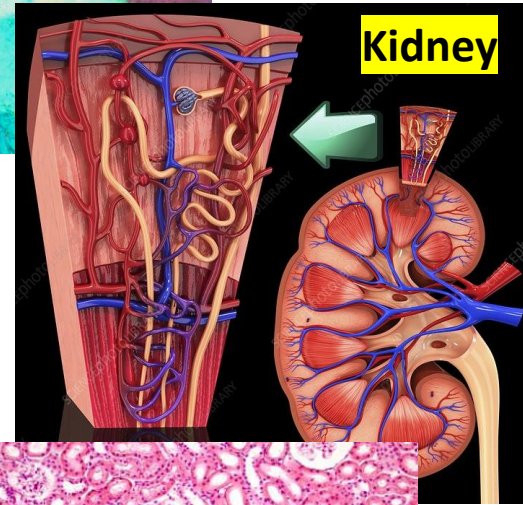
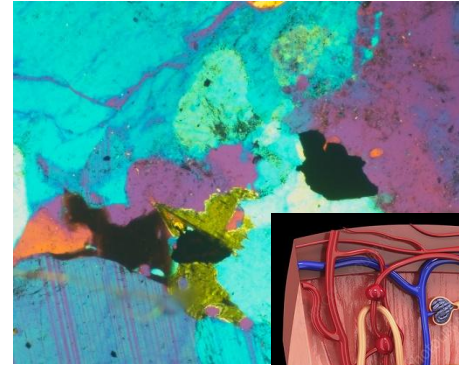
“The plural of anecdote is not data !” ... Measure it yourself !

- Why measure ?
- What do you want to measure ?
- How do we measure ?
- Are the results unbiased, precise, accurate, valid, meaningful ?

Individual Objects



Structures in Rock/Tissue Sections



What?

Things which may be measured

- **Structures (3D)**
 - Number, size, surface area, length, volume, shape, ...
- Mass
 - Density/Weights of organelles, cells, tissues, organs, people, ...
- Shapes & Arrangement
 - Macromolecules, organelles, cells, tissues, organs, people, ...
- Chemical Constituents
 - Storage products, DNA
- Activity – Time (4D)
 - Enzyme activities, intracellular events, cell turnover, movement

What?

Quantitation



Geometrical Properties

- Sampling
- **Morphometry** (... directly)
- Stereology (... indirectly 2D/3D)
- Pattern Analysis
- Image Analysis

(Units: 1mm = 1000 μ m)

Minimum size resolved by ...



0.25 μ m

0.1mm (100 μ m)

Microscopists measure sizes about 1 μ m to 5mm

Optical Properties

- Analytical Microscopy
 - Reflectometry, Phase Contrast/Refractometry, Polarising, Interference, 'Weigh cells', Microdensitometry
- Semi-Quantitative
 - Rating scales, ++++

What?

What to Measure - Structures

- **Size**
 - Lengths, Widths, Heights
- Amount
 - Lengths, L_v ; Surface areas, S_v ; Volumes, V_v
- Numbers
 - N_A , N_v , N
- Shapes
 - Roundedness, Indentedness, S:V ratios, Form Factors, Tortuosity
- Orientations
 - Angles, Isotropic, Anisotropic, Branching
- Locations & Patterns
 - Random, Clumped, Dispersed, Related/Connectivity

What?

Examples

- **Size of diatoms, forams, tardigrades, pollen, bugs, hairs, sand grains, blood cells, muscle cells, ...**
- Number of neurons in brain samples
- Percentage / Number of dividing cells in sample
- Proportion of cells, nuclei, vessels in liver section
- Length of capillary network in tissue
- Surface area of villi in gut, alveoli in lung
- Orientation/branching of Purkinje fibres in cerebellum
- Relationships of organism type 1 to organism type 2

What?

Applications

- **Microscopy**, Histology, Pathology
- Botany, Zoology, Anatomy, Embryology
- Radiology
- Food Science
- Metallurgy
- Materials & Computer Sciences
- Geology
- Ecology
- Geography
- Social Sciences
- Astronomy

- Why measure ?
- What do you want to measure ?
- How do we measure ?
- Are the results unbiased, precise, accurate, valid, meaningful ?

How?

“ One ounce of thought is worth one ton of equipment.”

Lord Rutherford



How?

Pre-Quantitation

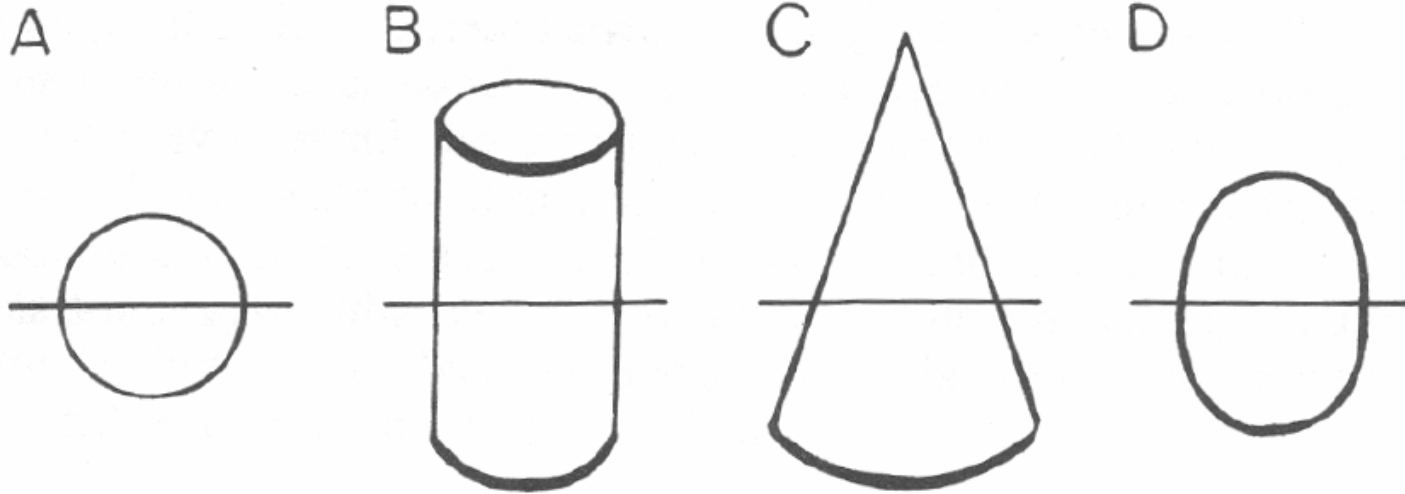


- Qualitative Analysis
 - Observation / Visualisation / Form / Organisation
 - Recording
 - 2D/3D Interpretation / Serial:Thick Sections / Reconstruction
 - Functional Interpretation
- Subjective Quantification
 - **Guesstimate ??**
Variability
 - Amount, many, more, larger, 0 to ++++
 - Activity
- Relate to other levels of organisation, up, down
- Relate to other methodologies – Physiol, Biochem, Living
- Artefacts / Misunderstandings

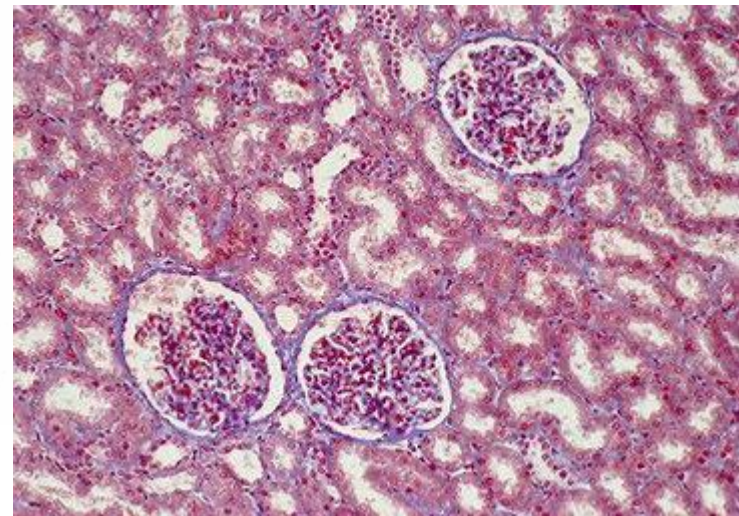
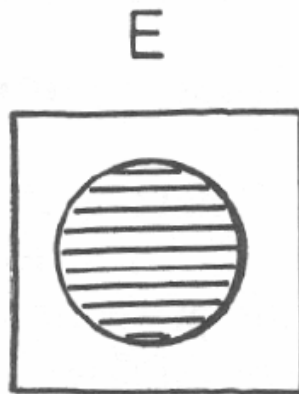
2D < > 3D

Problems using sectioned tissues !

Objects



Sectioned Profiles of objects



2D < > 3D

Problems using sectioned tissues !

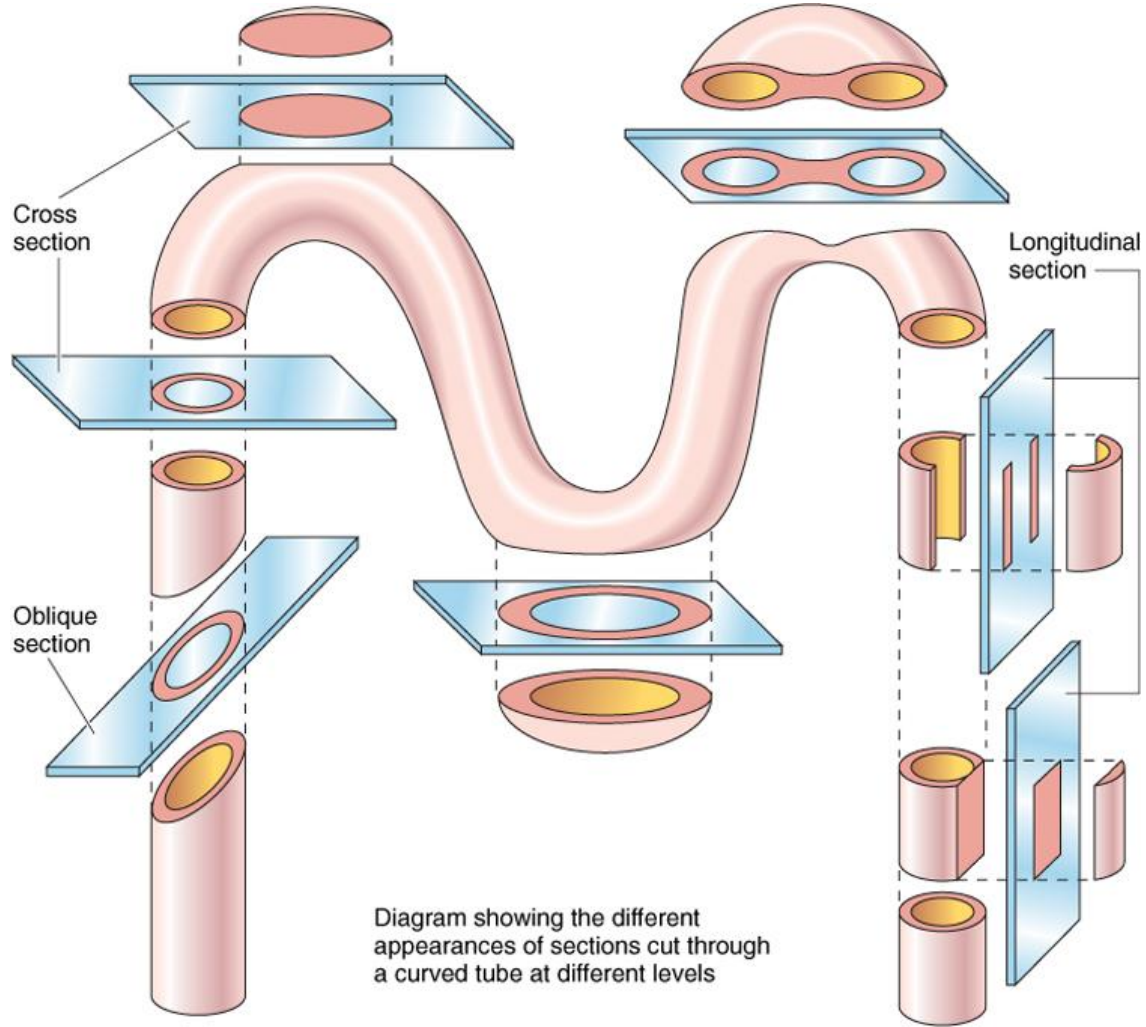
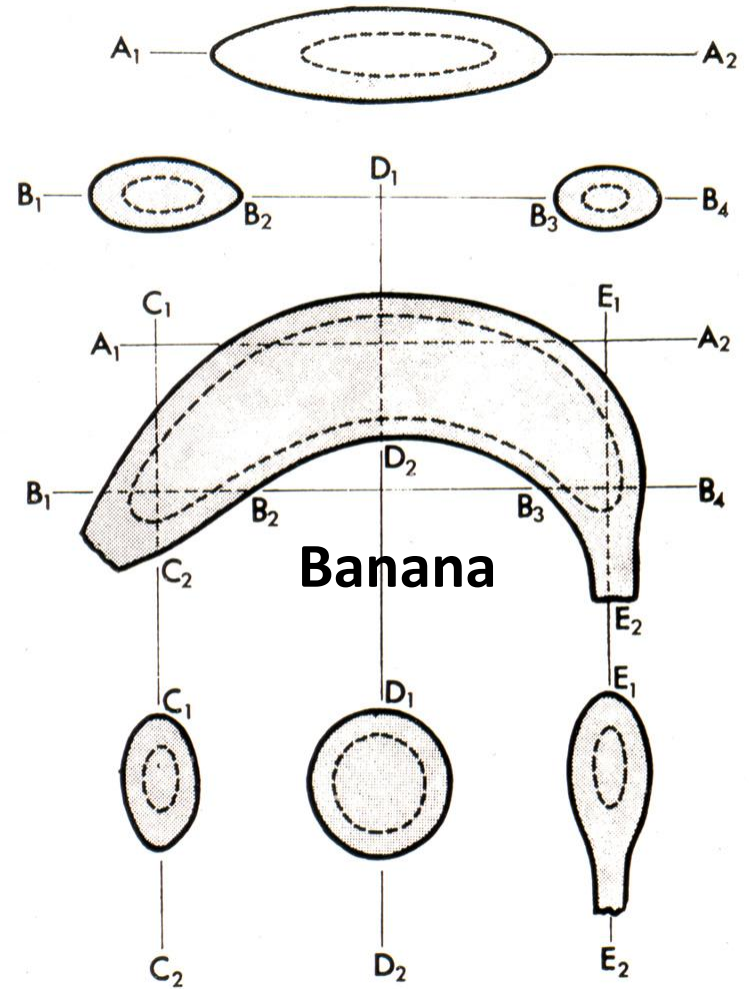
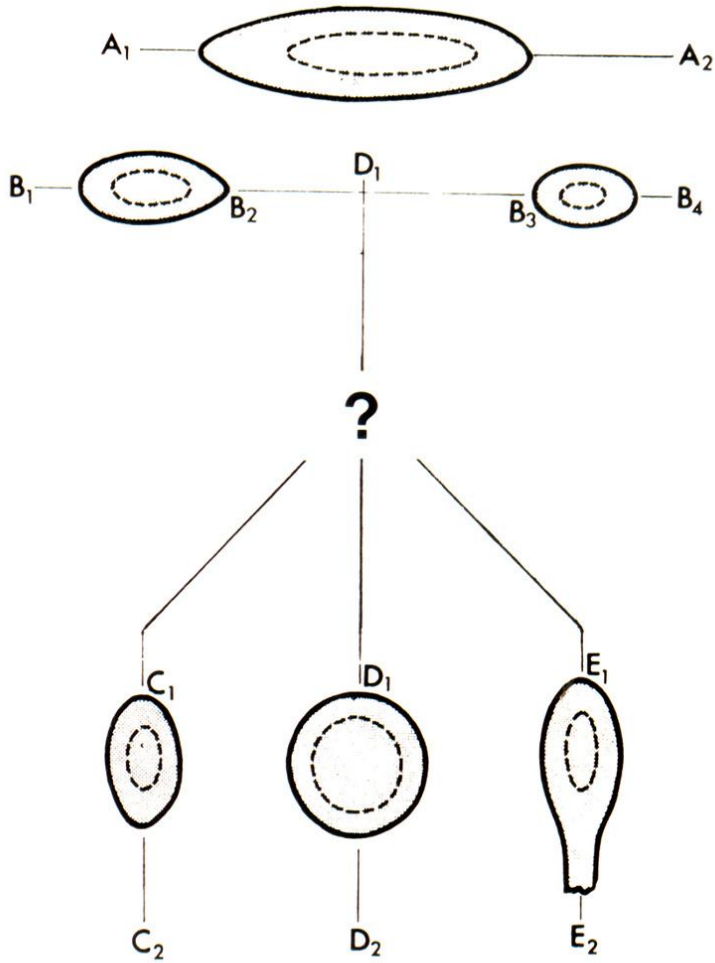


Diagram showing the different appearances of sections cut through a curved tube at different levels

2D < > 3D

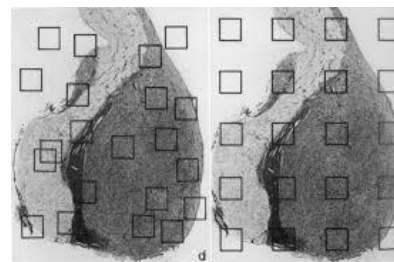


How?

Sampling



- Identify Object
 - Staining
 - Magnification
- Representative
 - Is tissue Homogeneous (Isotropic) / Irregular (Anisotropic)
Heterogeneous (Anisotropic)
Gradiential (Anisotropic)
- Random
 - Completely random
 - Systematic stratified random sampling
 - Zonal oblique sector analysis
- Manual / Automated
- How many samples? (Experimental Design)
 - Individuals / Organs / Blocks / Sections / Micrographs / Items / Measures
 - Hally Formula $RSE = \frac{SQRT(1-Vv)}{SQRT n}$
 - Progressive mean, Log Plots
 - **Do More, Less Well !**



Progressive Mean (Running Average)

Data: 80, 10, 30, 40, 50, 45, 30, 35, 40, 40,

Average
Size
 μm

100

80

60

40

20

0

10

20

30

40

50

60

70

80

90

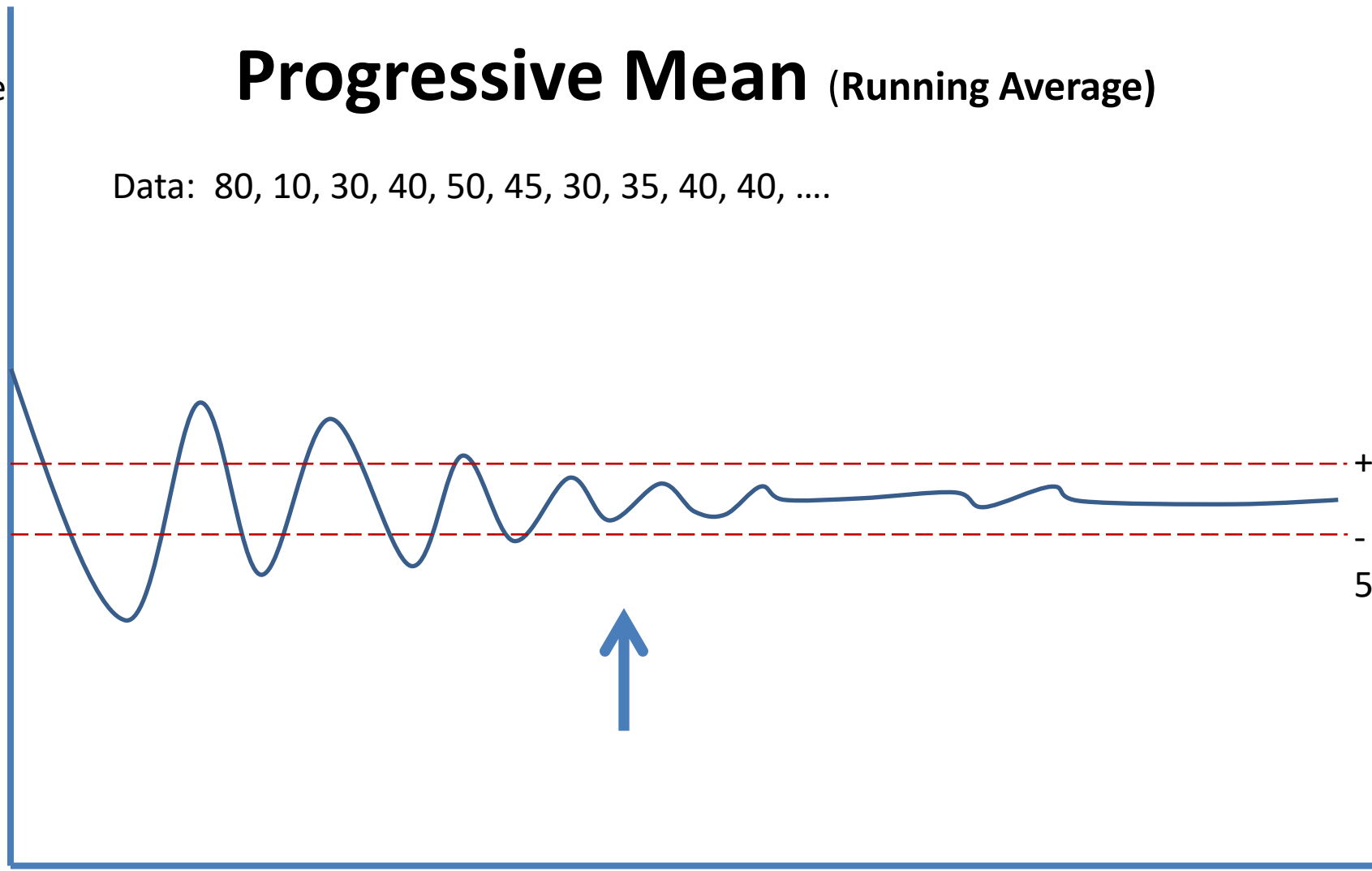
100

Number

+

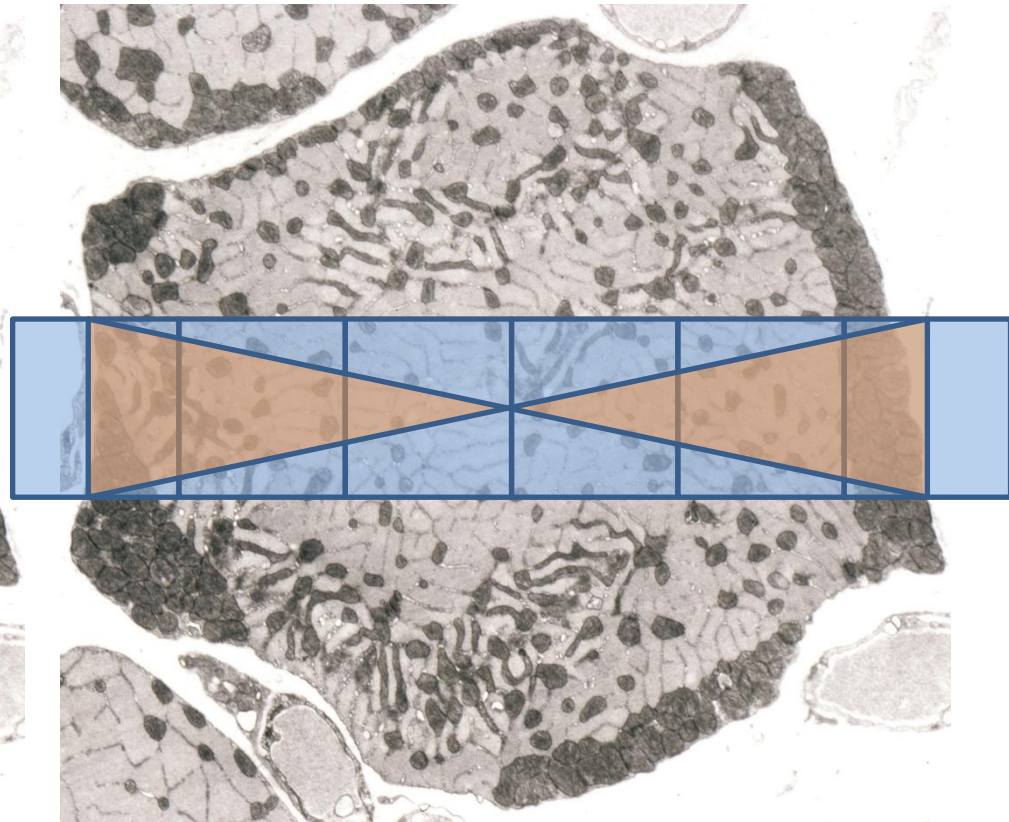
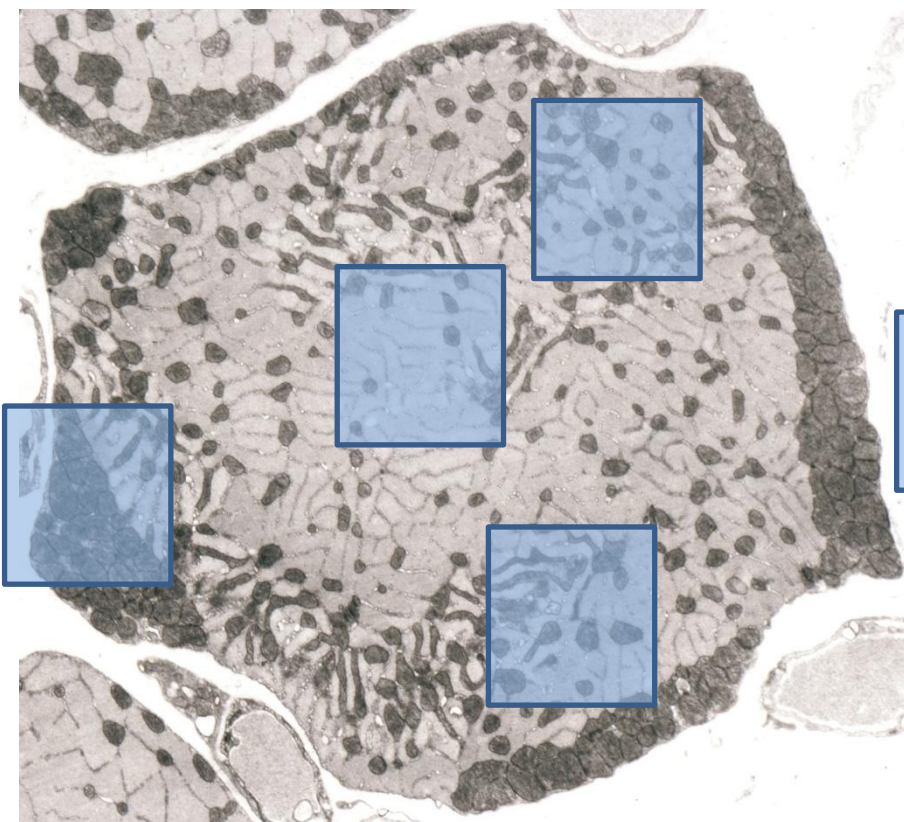
-

5%



Skeletal Muscle – Random Sampling
Micrographs at x18,000

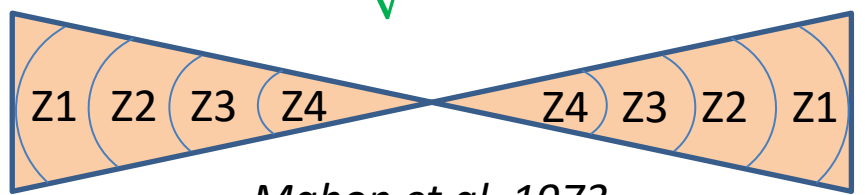
Skeletal Muscle – Zonal Oblique Sector
Analysis Method (ZOSAM)



X

Sampling: $\leq 1, RSE$ ($\sqrt{(1-V_v)/n}$) or *Prog. Mean*

✓

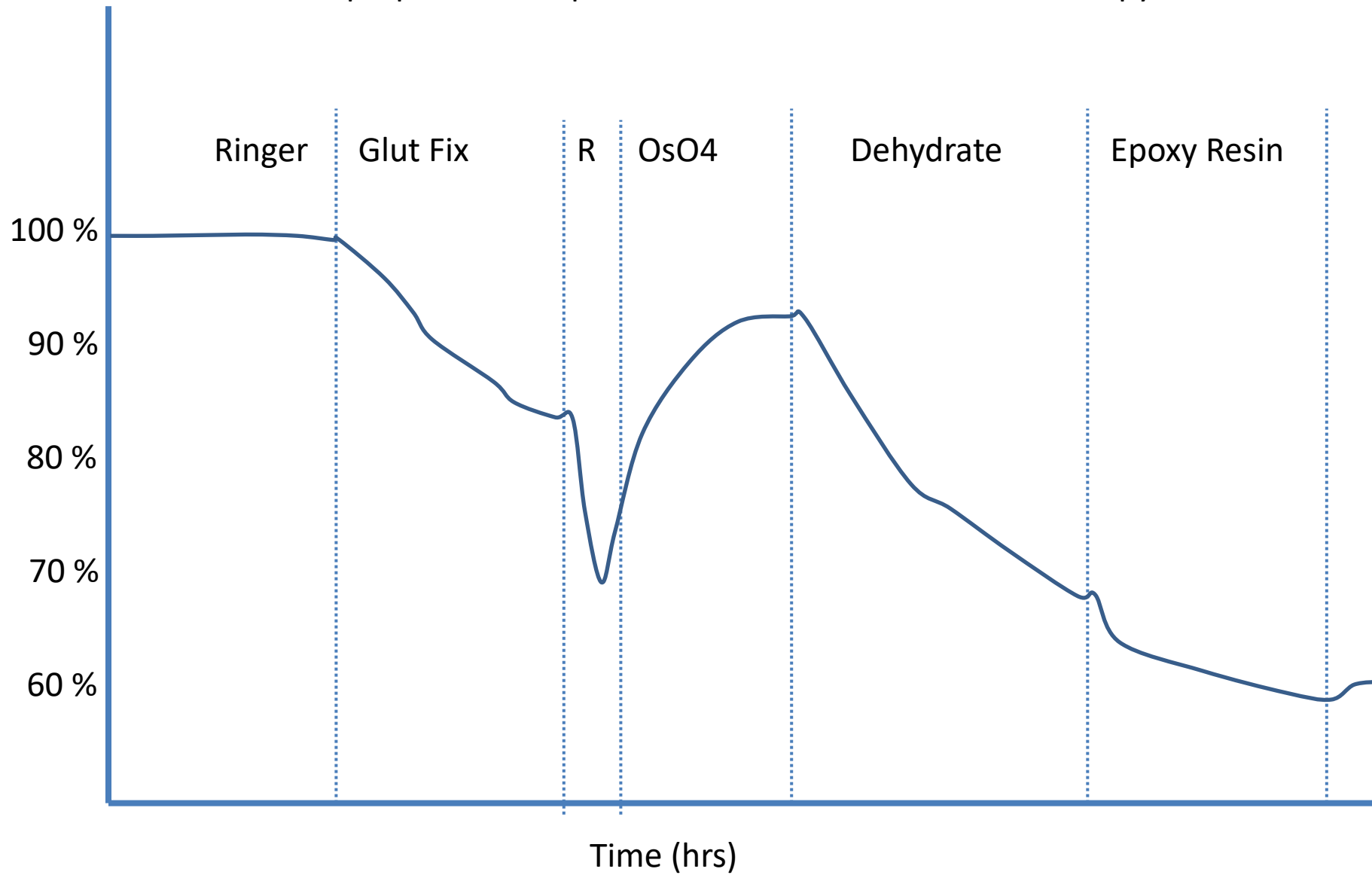


Mahon et al, 1973

Tissue Preparation Effects

- **Fixation Effects** (Sissons, Goldspink, Strickland 1960s, 1970s)
 - Flemmings - 3-15%
 - Carnoys -19-36%
 - Bouins -23%
 - Formalin -23-30%
 - Zenker -30-40%
- Length change 10%
- Areal change 21%
- Volume change 33%
- Need a Standard Ringers, Frozen, ...

Tissue preparation steps for transmission electron microscopy



How?

Which Method ?

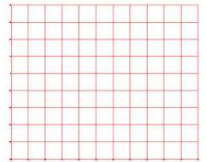
- **Morphometry**

- direct measurement of structures



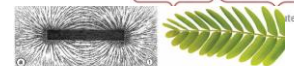
- **Stereology**

- extrapolation from 2D to 3D using simple counting methods



- **Image Analysis**

- combination of above using digital imaging and computers in manual or automatic modes
- plus data presentation and analysis



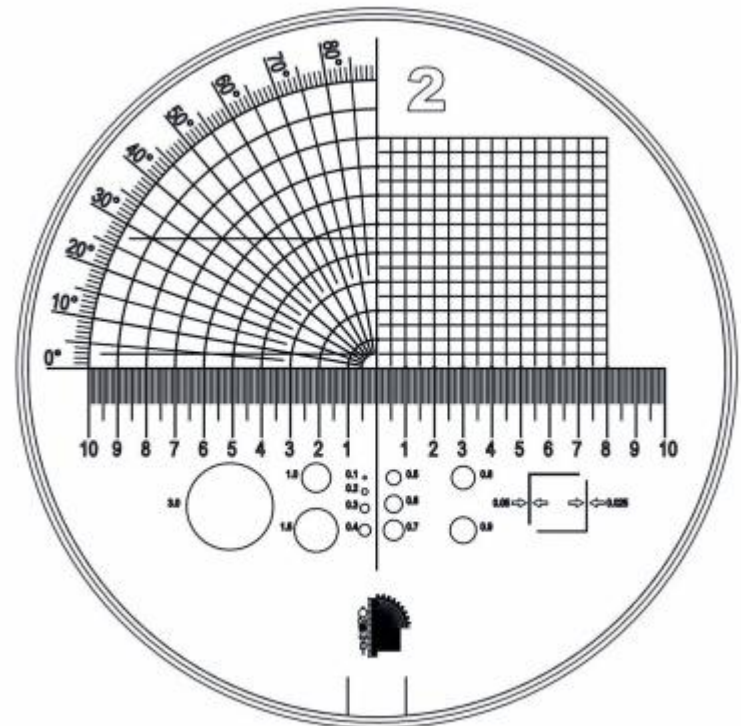
[See Russ, 2019 Robust Measurements](#)

If object is large enough ..



Measuring Magnifier (Loupe)

+ Reticle / Graticule



Microscopy Practical

- 1. Magnification Calibration**
- 2. Morphometry**
3. Stereology
4. (Pattern/Shape analysis)

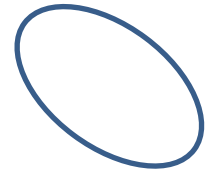
How?

Morphometry

Measurement of Form

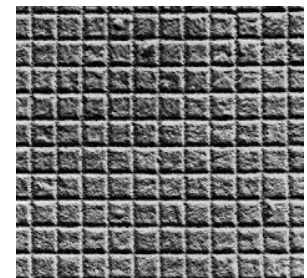
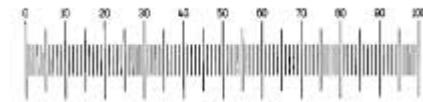


- Diameters & Areas
 - Longest, shortest/narrow, Orthogonal, Feret, Average, Random chord
 - From area (πr^2), Circle of best fit
- Shape
 - Axial ratio, S:V ratios or Form factors (A/P^2 , $1=4 \pi A/P^2$), Angularity (Gulfs & Peaks)
 - Shape of best fit (Identikit), Reconstruction
 - Fourier Analysis, Fractals (Mandelbrot)
- Volumes, Lengths ⚠, Surfaces ⚠, Numbers ⚠ ⚠ ⚠, Thickness

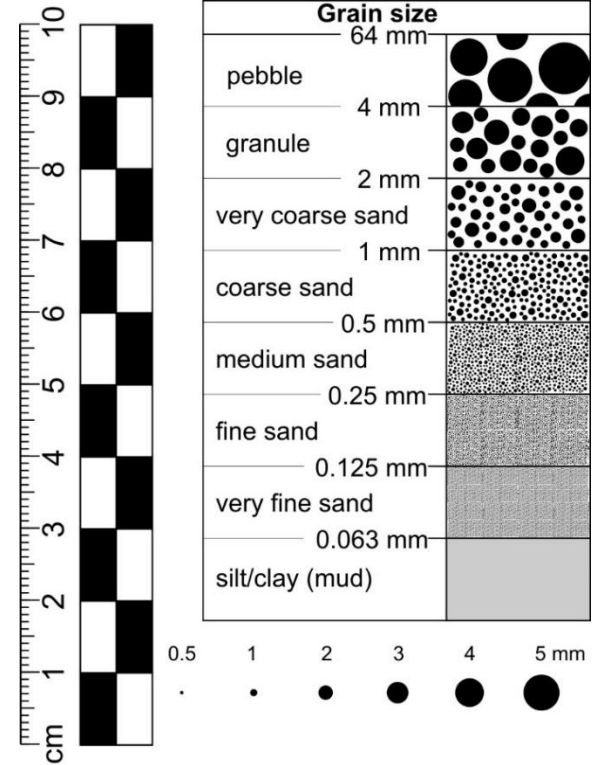
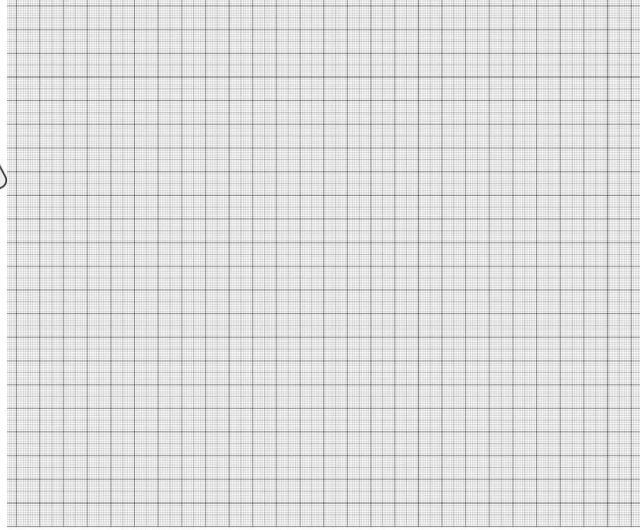
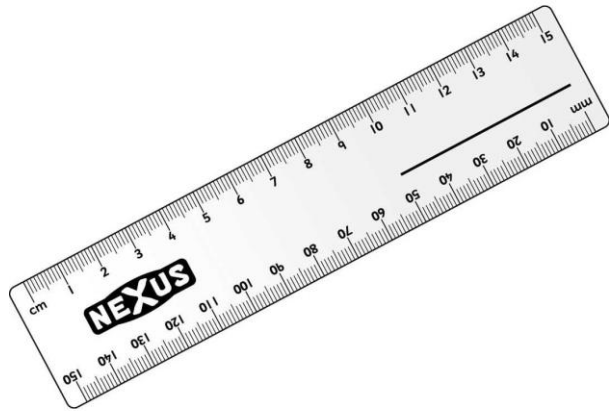


Equipment & Magnification Calibration

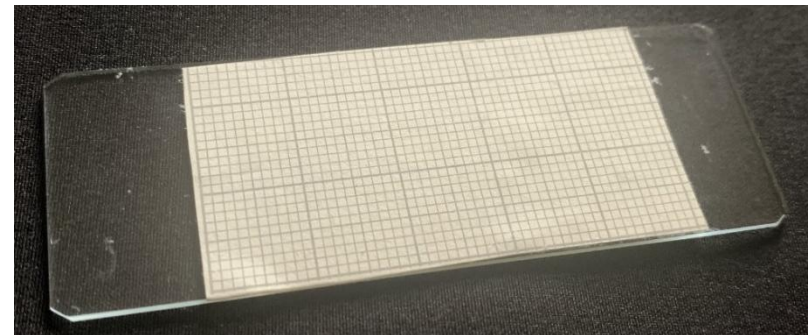
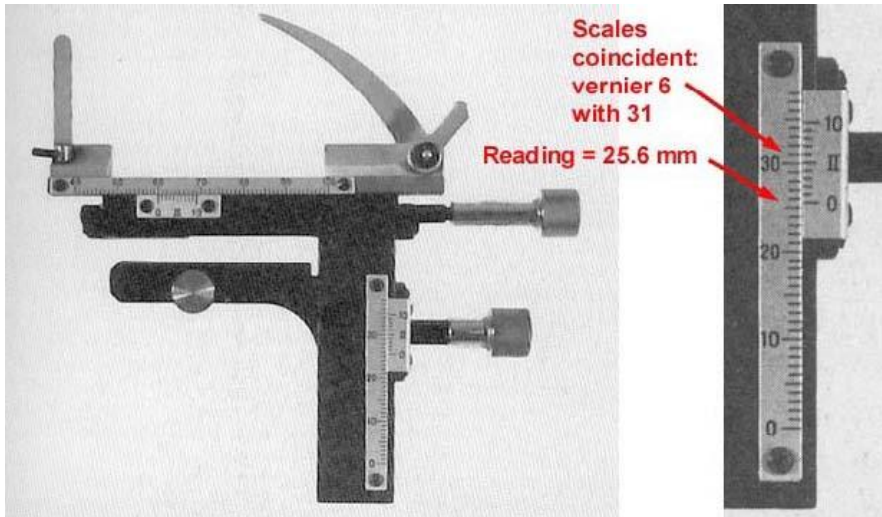
- Rulers, Calipers, Graticules, Stage Micrometer, Cut & Weigh
- EM: Diffraction Grating, Latex Spheres, Crystals
- Filar micrometer, Image Shearing micrometer
- Photographs, Drawings, Projection (Camera Lucida/Drawing Tube)
- Thread, Map Measurer (Opisometer), Planimeter
- Stereological lattices
- Image Analyser



Magnifier or Stereomicroscope

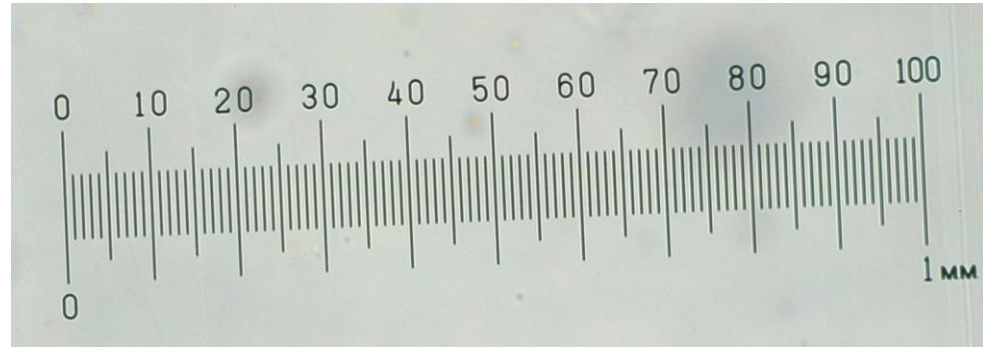


Compound Microscope



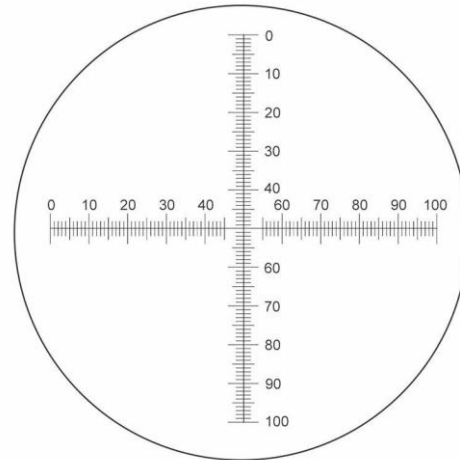
MM scale slide – 1mm squares

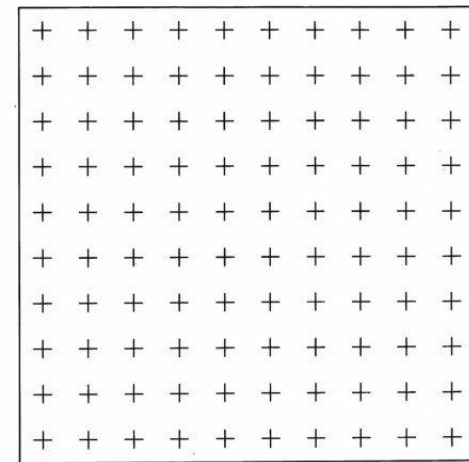
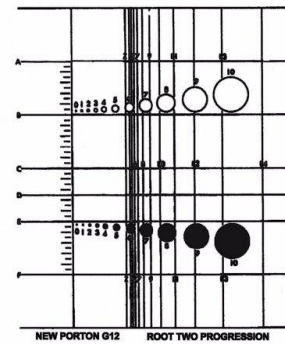
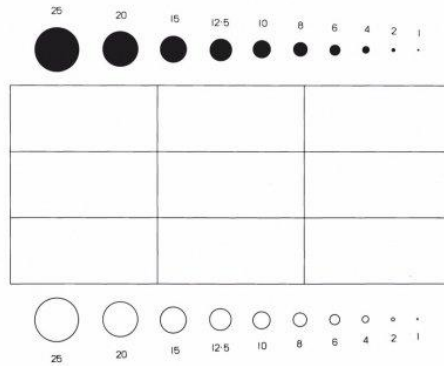
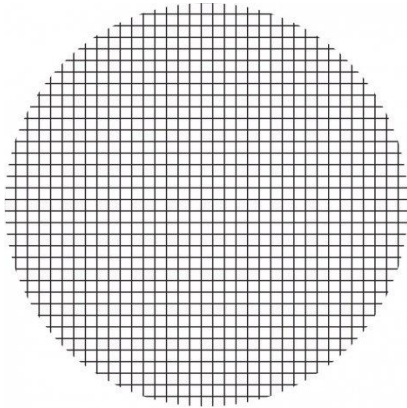
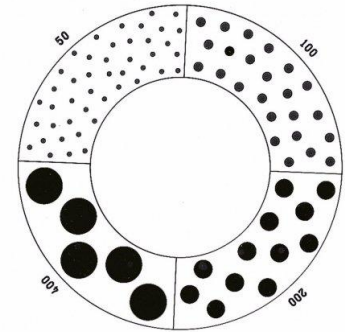
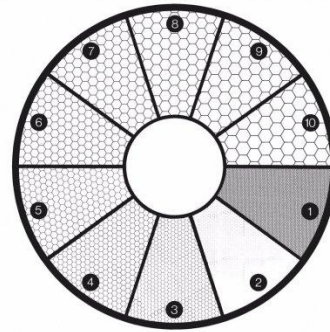
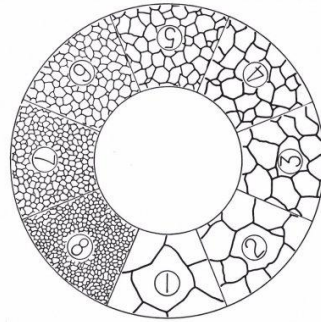
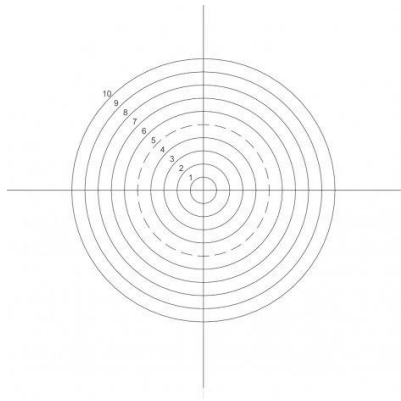
Stage Micrometer Slide



- As a guide / **guestimate**
- Measure width of **field of view** for each objective
- **Magnification / Scale bar**
- **Transfer scale to Eyepiece Graticule**

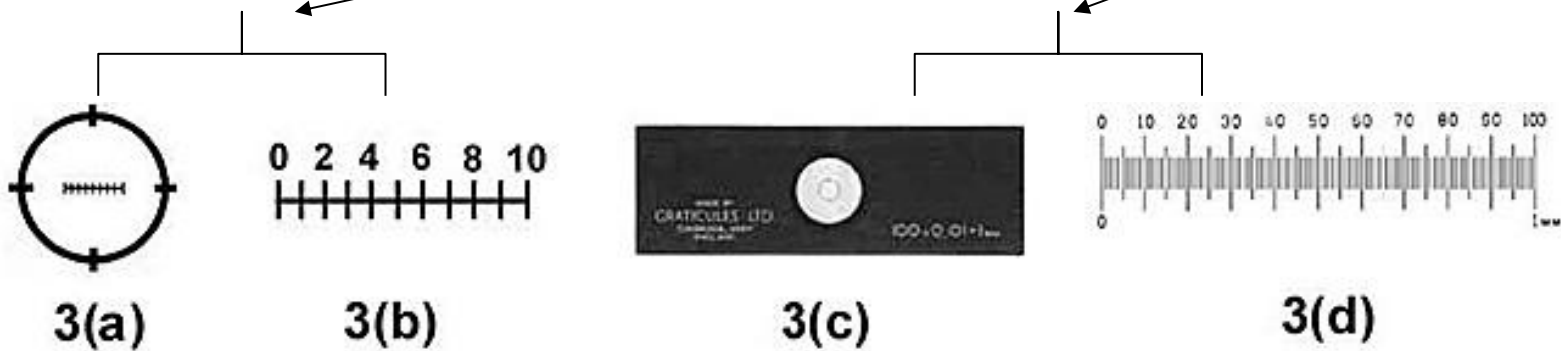
Eyepiece Graticule Scale



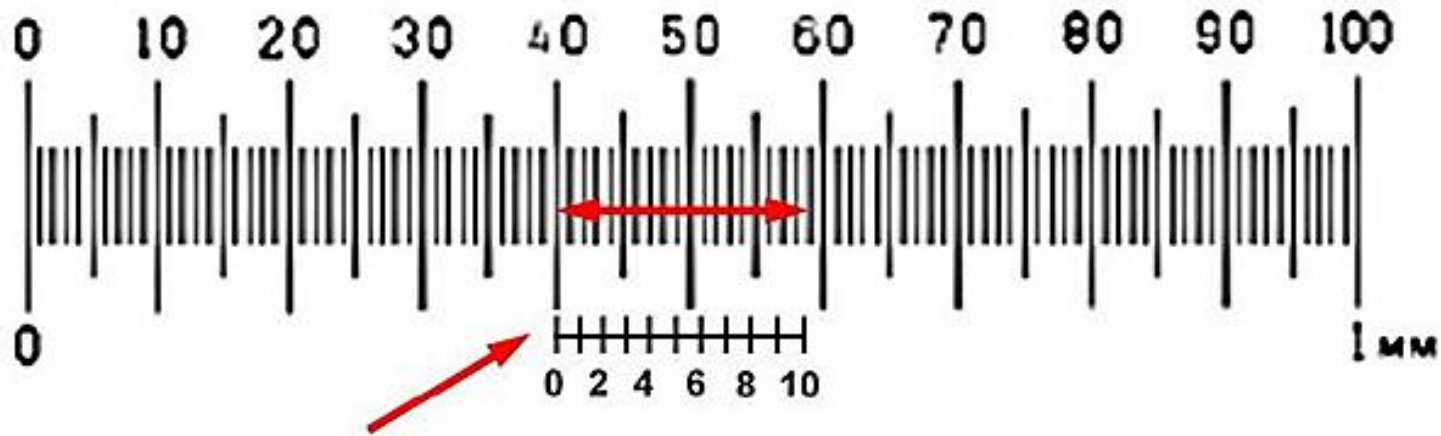


... some of the Eyepiece Graticules made by Graticules Optics Limited

How to calibrate and Eyepiece Graticule against a Stage Micrometer



1



Calibration of arbitrary scale: 10 units = 190 μm , at this magnification

Each arbitrary unit = 19 μm

2

**Then replace Stage Micrometer with your slide and 'measure' objects with Eyepiece scale
e.g. 2 units = 38 μm**

Transfer Scale ...

+ Take a Photo !

Filar Micrometer Eyepiece

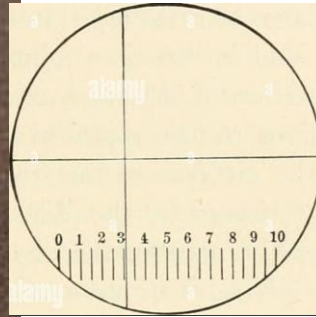


Image Splitting Eyepiece (Watson)



Image Shearing Eyepiece (Vickers)

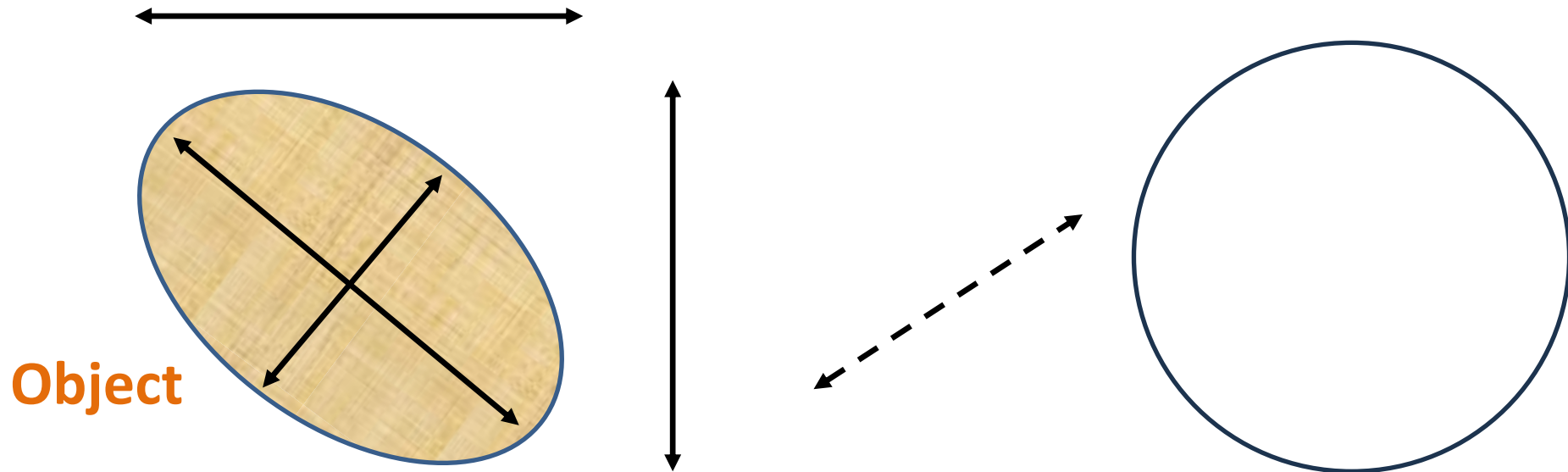
... very precise

Measuring Larger Items precisely ...

Travelling Microscope



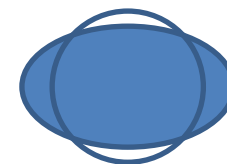
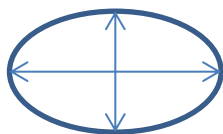
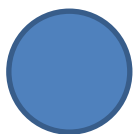
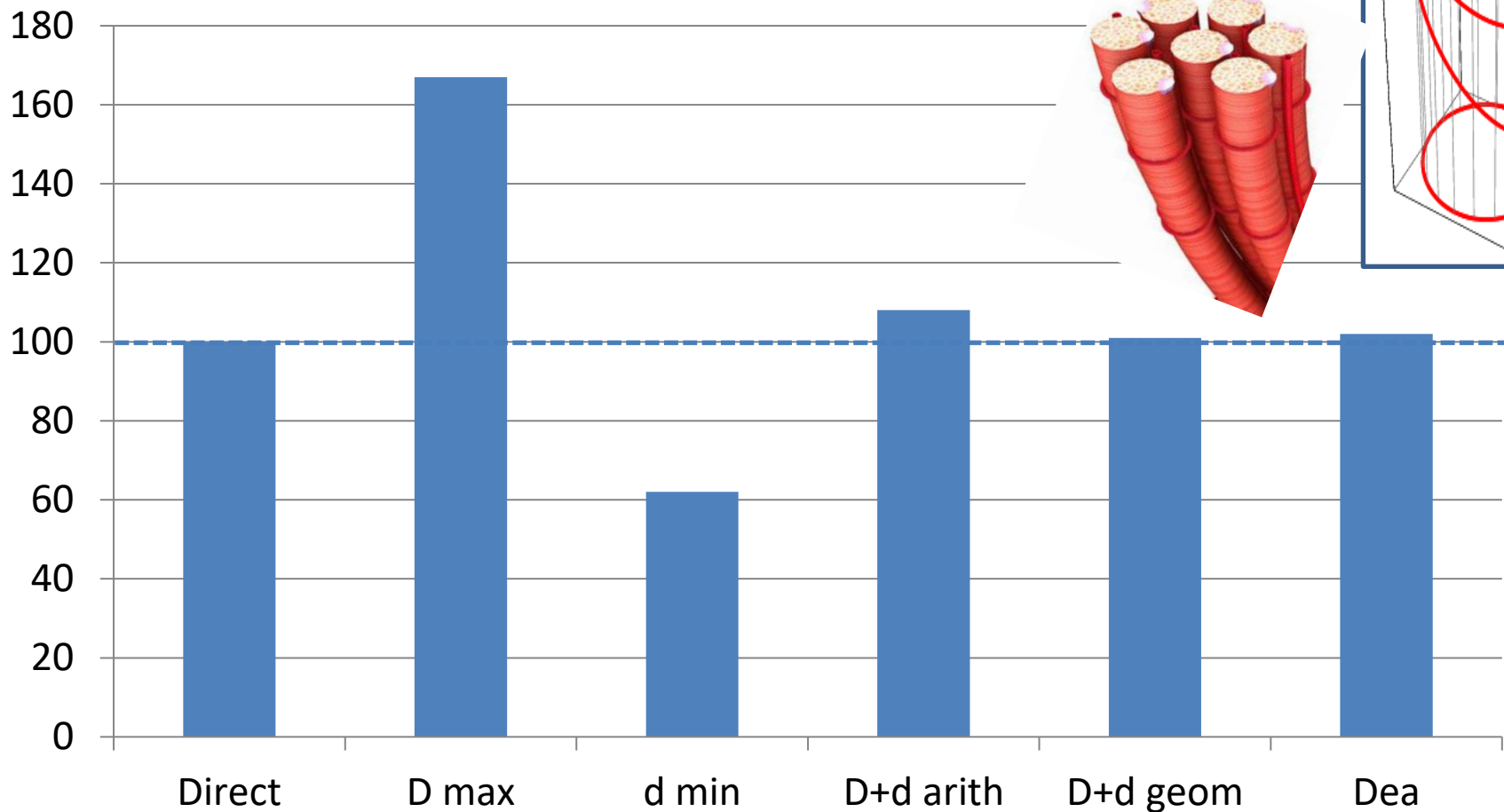
Diameter: Some Problems to consider



- Narrow diameter
- Maximum diameter
- Average diameter
- Orthogonal diameter
- Feret (Caliper) diameter
- Random chord diameter
- Diameter of Circle of best fit
- Diameter calculated from measured area using πr^2

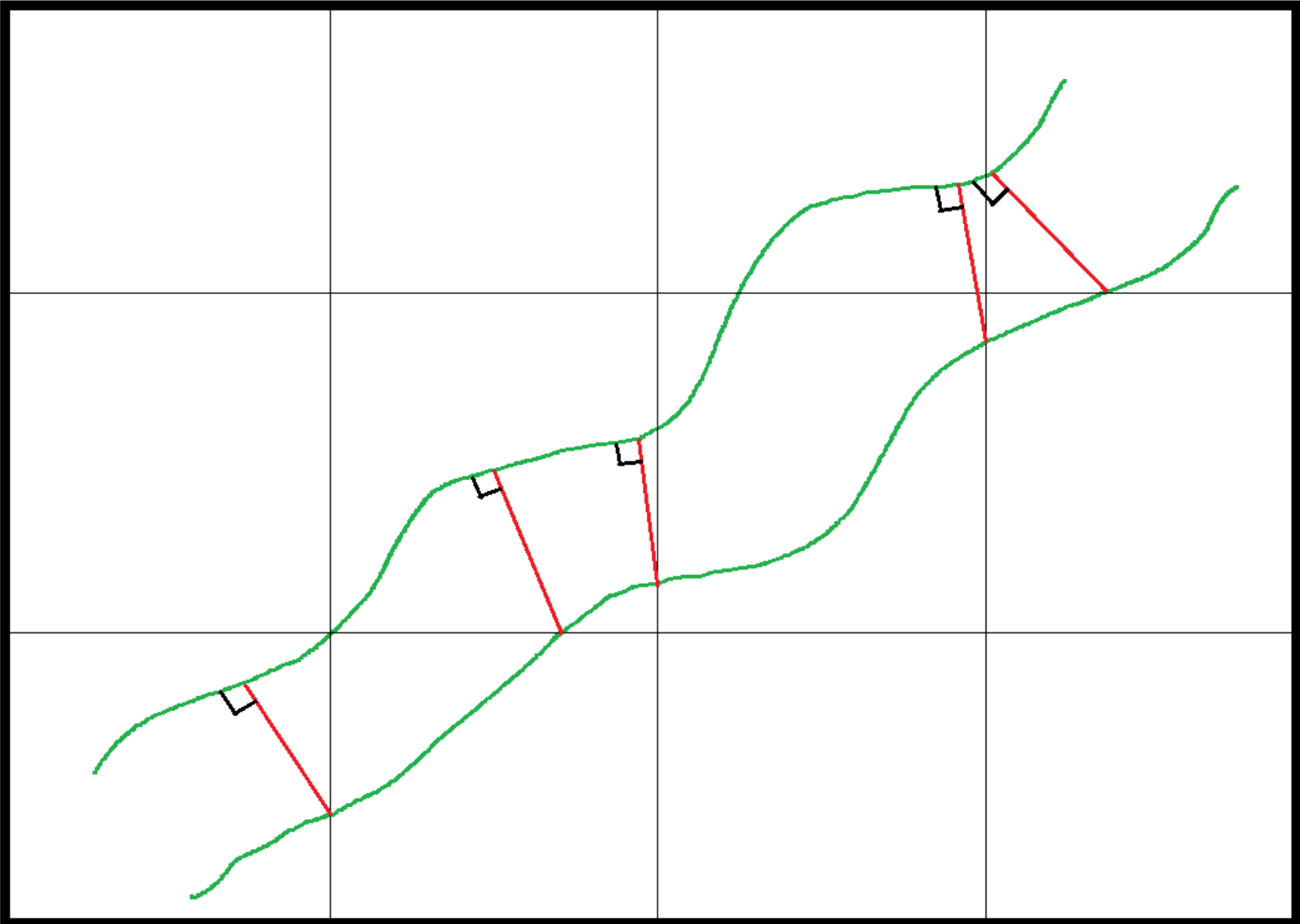
Diameters of Cylinder Profiles

Diameter (Muscle Fibres)



Some problems ...

Thickness Measurements



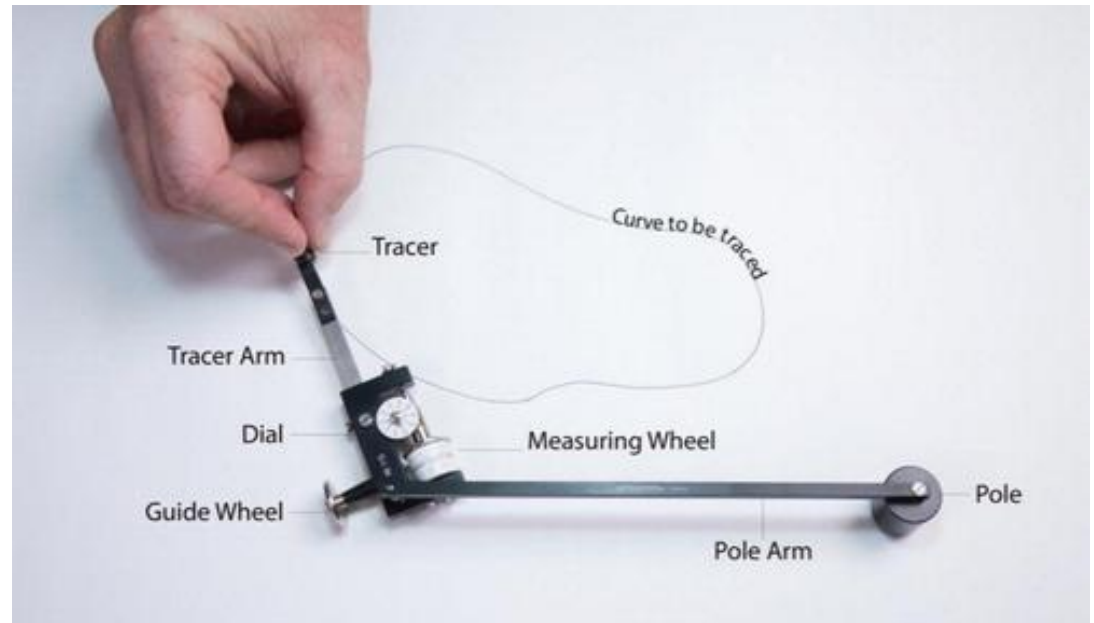
Map Measurer (Opisometer) for measuring Lengths



... or use a cotton thread !

On projected images, drawings, photographs

Planimeter For measuring Areas



Length



- Dependent on Magnification
- How long is the coastline of Britain? (Mandelbrot (1967) Science)



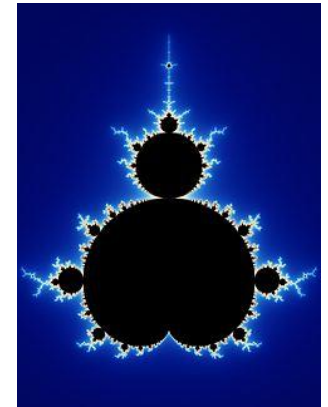
U 200 km
L 2400 km



U 50 km
L 3400 km



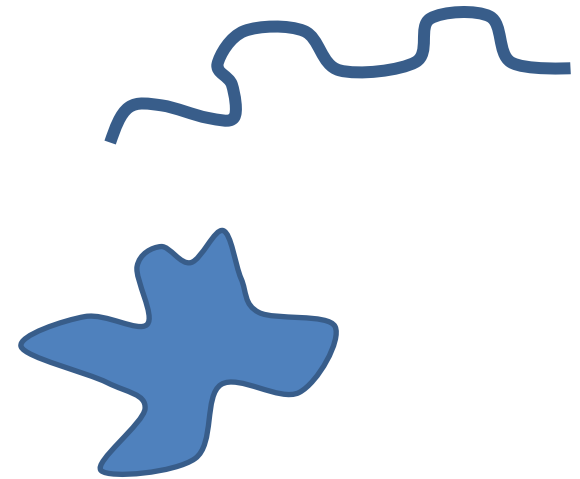
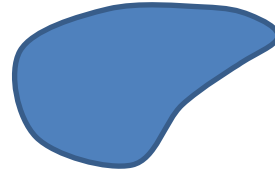
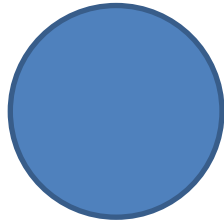
U ?? km
L 12450 km



[See also Russ et al 2018](#)
[The Problem of Perimeter](#)

Fractal Dimension of
Wiggleness 1.25 !

Shapes



Form Factor

1.0

0.8

0.4

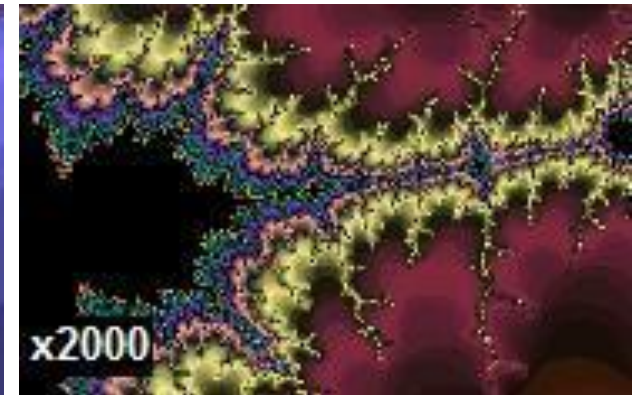
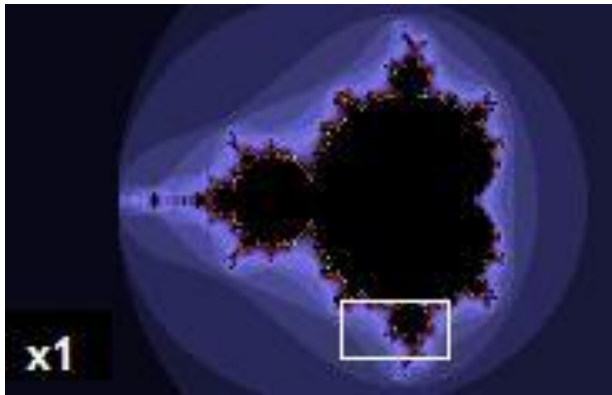
$$F = \frac{4 \times \pi \times \text{Area}}{\text{Perimeter}^2}$$

Gulfs

0

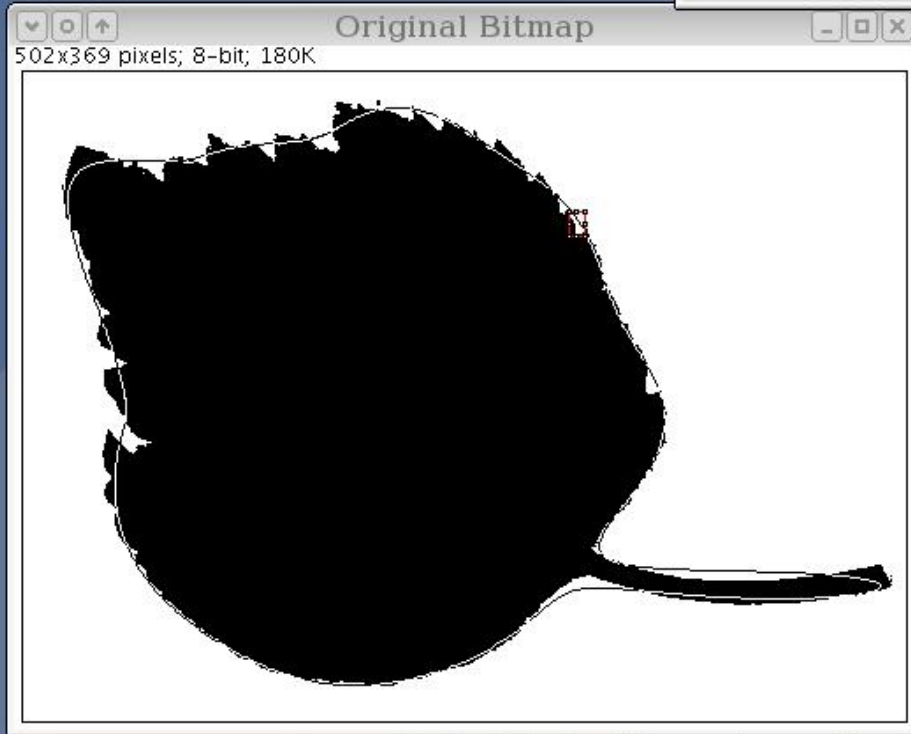
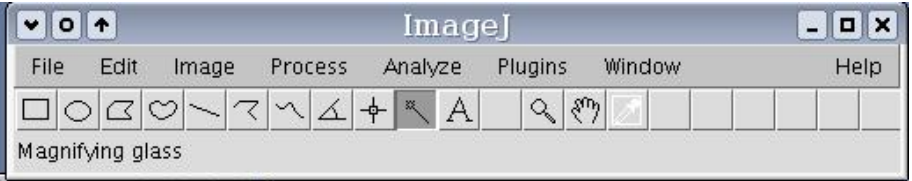
1

5



Fractals – Mandelbrot Set

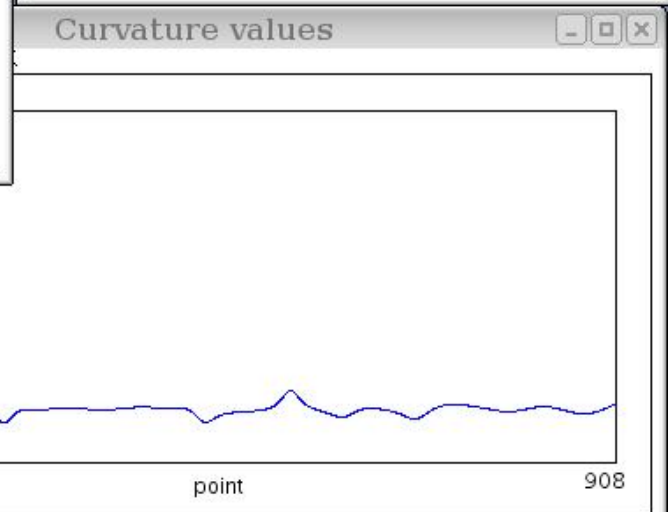
Fourier Analysis of Shape Splines



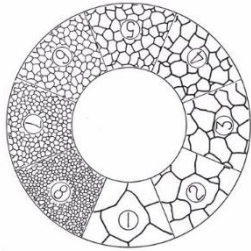
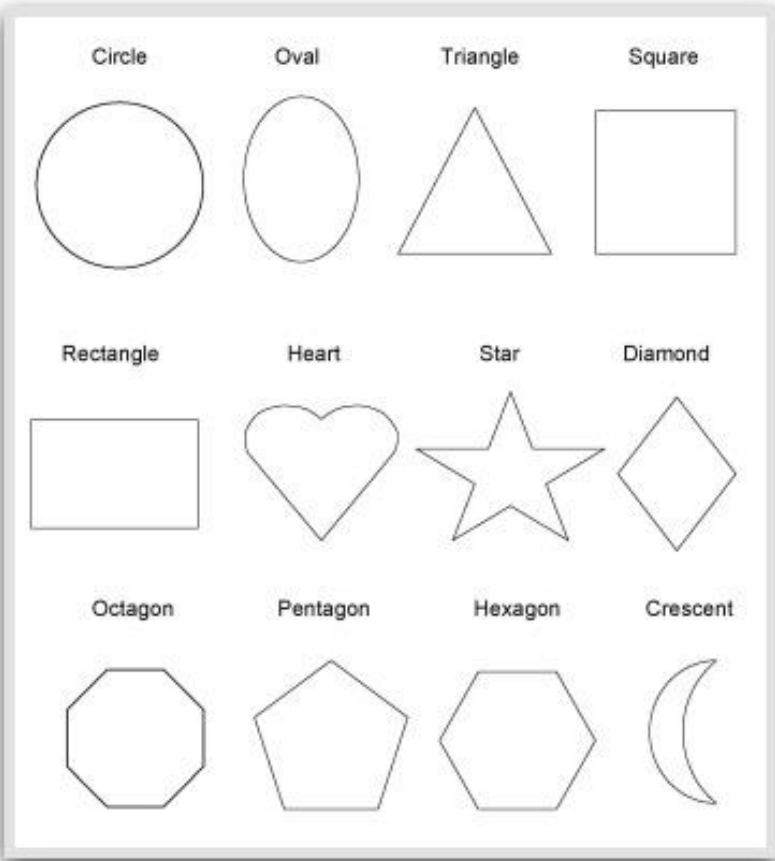
Results

	ax	ay	bx	by
3	-19.217	1.817	28.348	-2.687
4	-24.824	13.000	-4.479	14.474
5	-2.337	3.177	-16.395	6.302
6	-6.342	-10.334	-10.648	-1.942
7	11.848	4.940	-11.013	0.918
8	10.298	-2.063	-0.936	1.363
9	9.235	0.315	6.396	2.284
10	0.049	-2.735	5.418	-2.759
11	-2.334	-0.483	3.379	-0.920

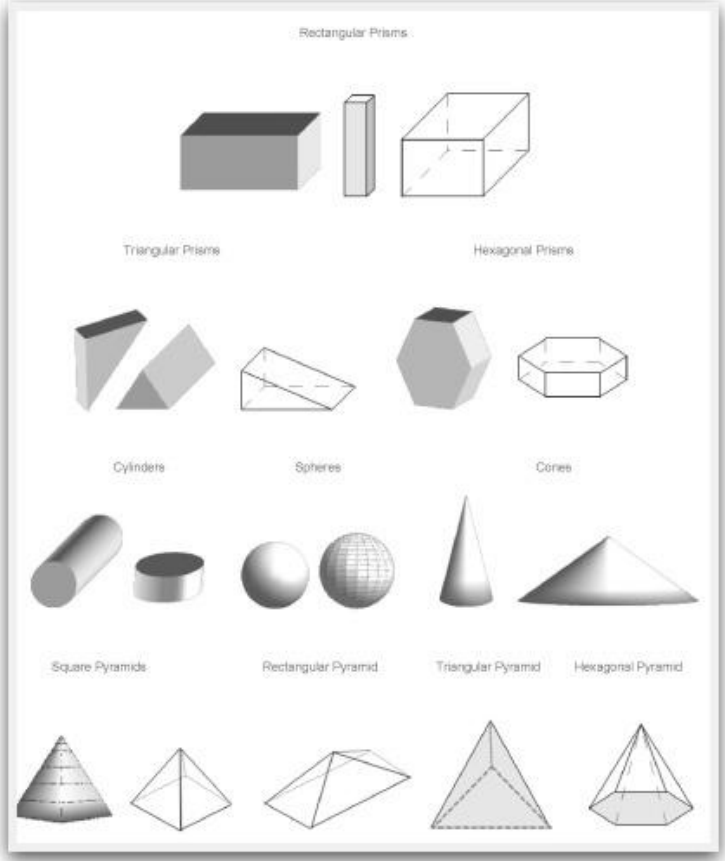
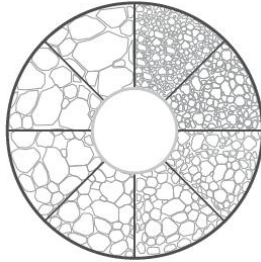
Took 8.09 seconds to calculate autocorrelation.



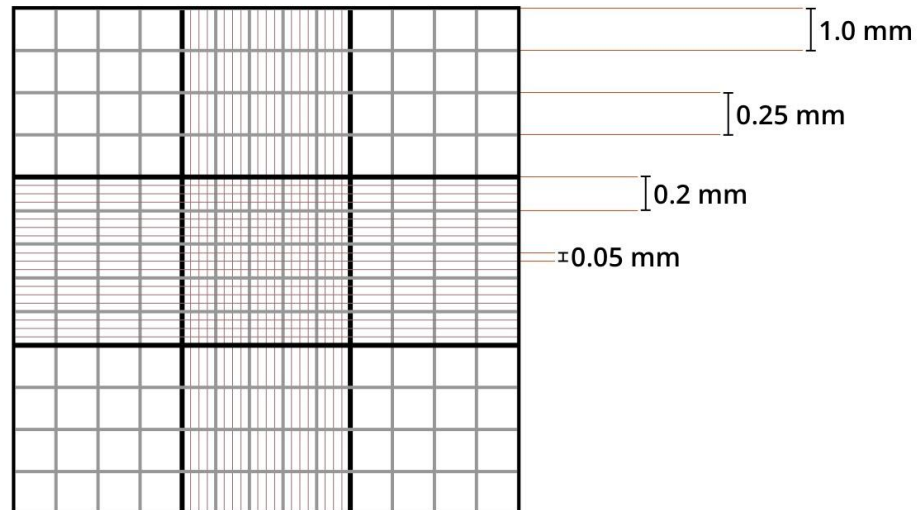
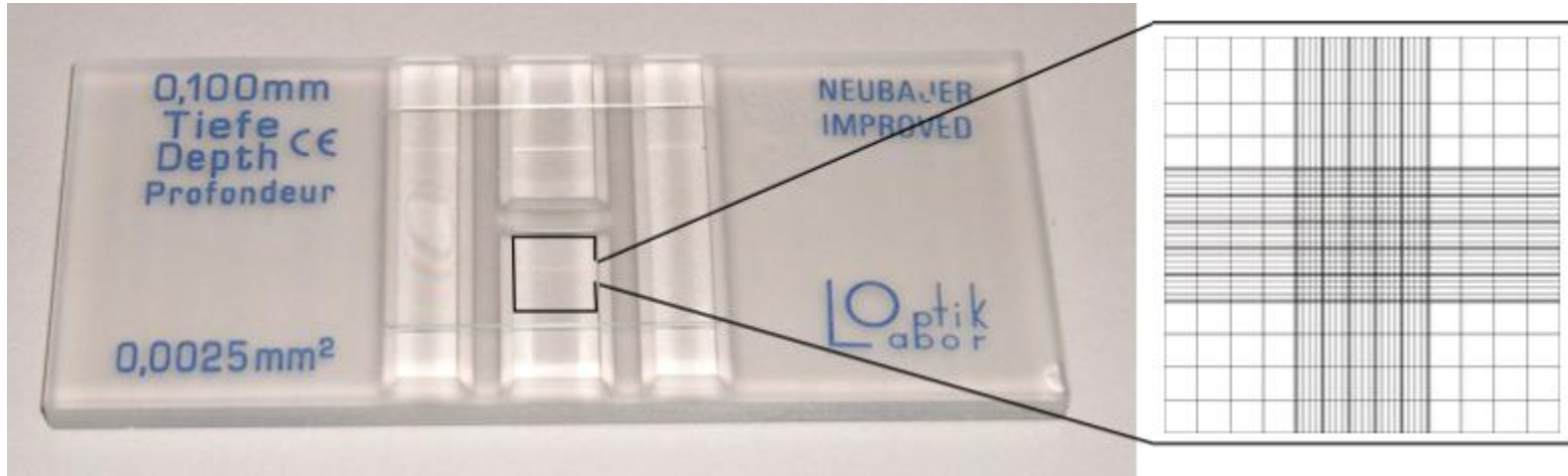
List Save... Copy...



Identikit Match



Numbers: Haematocytometer Slide



Number of cells in a 1mm² square x 10⁴ = No. cells/ml.

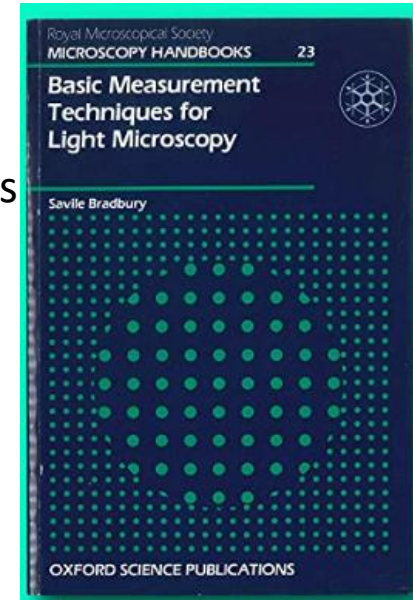
Reading: Bradbury S 1991 Basic Measurement Techniques for Light Microscopy RMS #23

Early History of Measurement in Microscopy

- 1665 Hooke - comparisons, scale bars on drawings, counts
- 1673 Leeuwenhoek - compared 'Little animalcules' v sand grains, rbcs
- 1679 " - made a graticule of 600 hairs per inch
- 1716 Hertel – stage micrometer
- 1718 Jurin – hair micrometer
- 1753 Baker – calibrated lattice wire micrometers
- 1771 Adams – screw micrometer
-
- 1904 Wright - eikenometer
- 1936 Patterson & Cawood – particle size eyepiece graticules

- Travelling Microscope
- Haemocytometer slides

- 1960 Barer /Dyson – image shearing/splitting micrometer
-
- **Counts & Proportions** (*Geology, Metallurgy, >Biology*)
- 1847 Delesse – areas \equiv volumes
- 1851 Sorby – tin foil cutouts
- 1916 Shand – mechanical automation
- 1939- 1990s– various electronic 'image analysers'

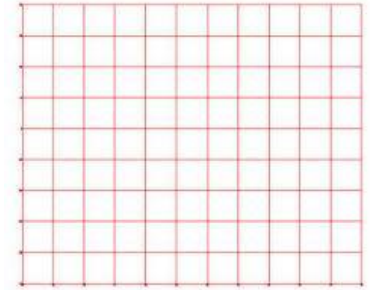


Henry Sorby – FRS, RMS, MMS

How?

Stereology 1

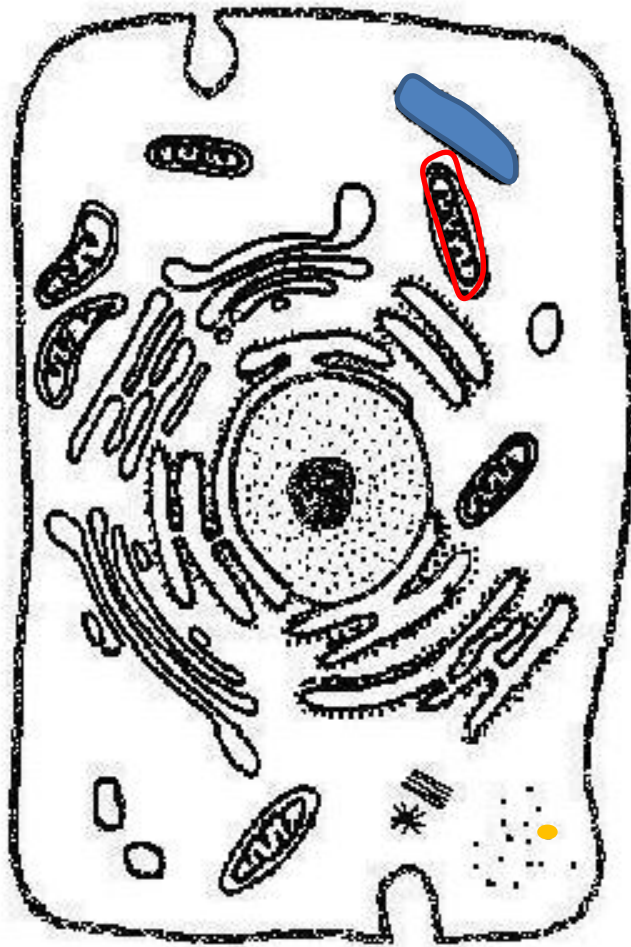
Extrapolation from 2D to 3D



Geology, Metallurgy, Engineering, Astronomy, Biology

- History
 - 1777 Buffon
 - 1850s Delesse, Sorby
 - 1961 ISS, Models; Elias, Weibel, Williams, Mayhew, Cruz-Orive
 - 1983 Unbiased/Designer: Gundersen (Cavalieri, 1635)
- Dimensional Reduction
 - Volumes, Surface Areas, Lengths, Numbers
 - Volume=3D, Area=2D, Length=1D, Point=0D
 - V_v , S_v , L_v , P_p Point Counting, Line Cuts, Counting
- Equipment
 - Probes (Sections, Lattices, Tally Counters, Image Analysers)
 - Isotropic Probes, Merz Lattices, Cycloids

Dimensional Reduction



OBJECT

SECTION

3D VOLUMES



AREAS



2D

2D AREAS



LENGTHS



1D

1D LENGTHS



POINTS



0D

0D POINTS



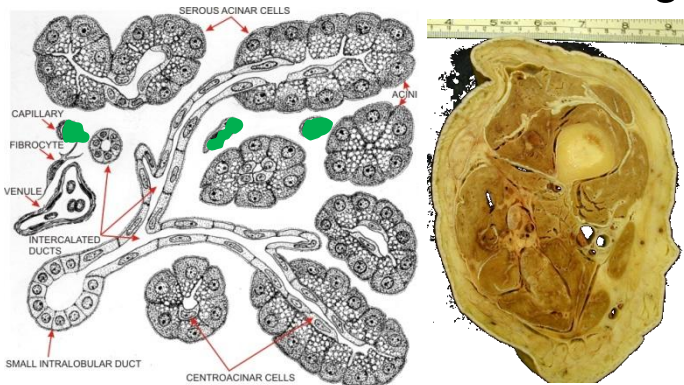
Hit



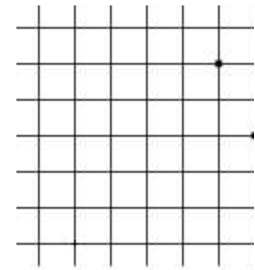
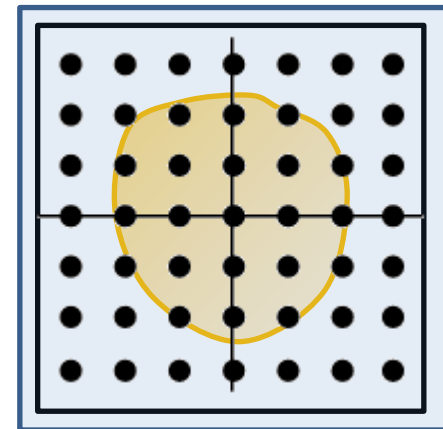
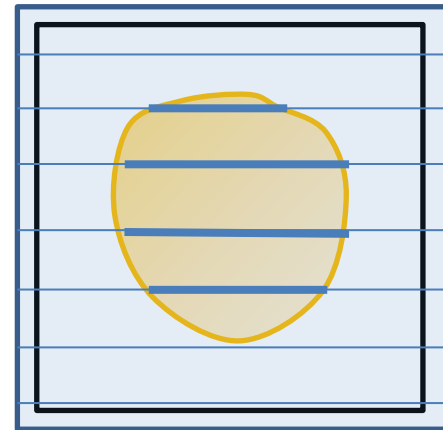
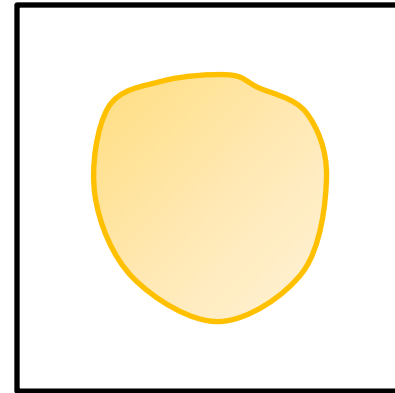
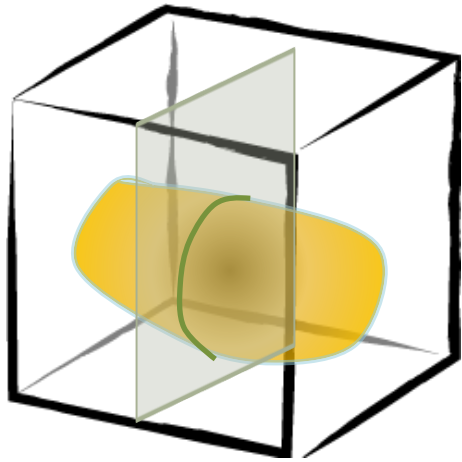
0D

Miss

X



Volume Fraction



$$V_V = A_A = L_L = P_P$$

Reconstruct

Planimetry

Measure

Count

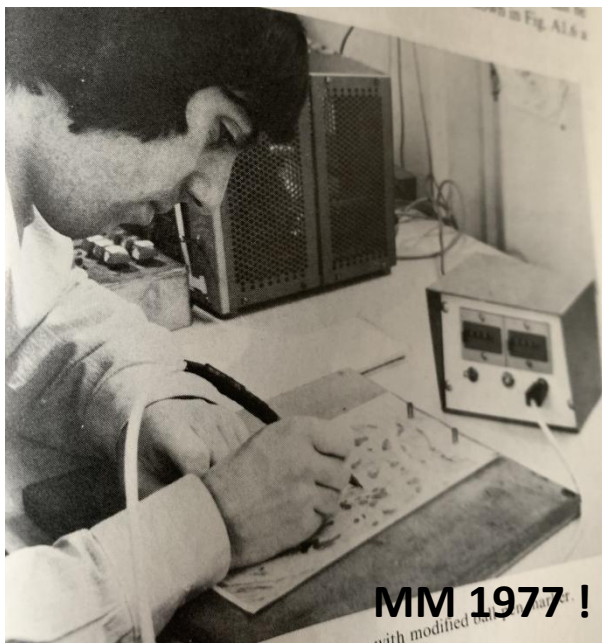
Cut & Weigh

Opisometer

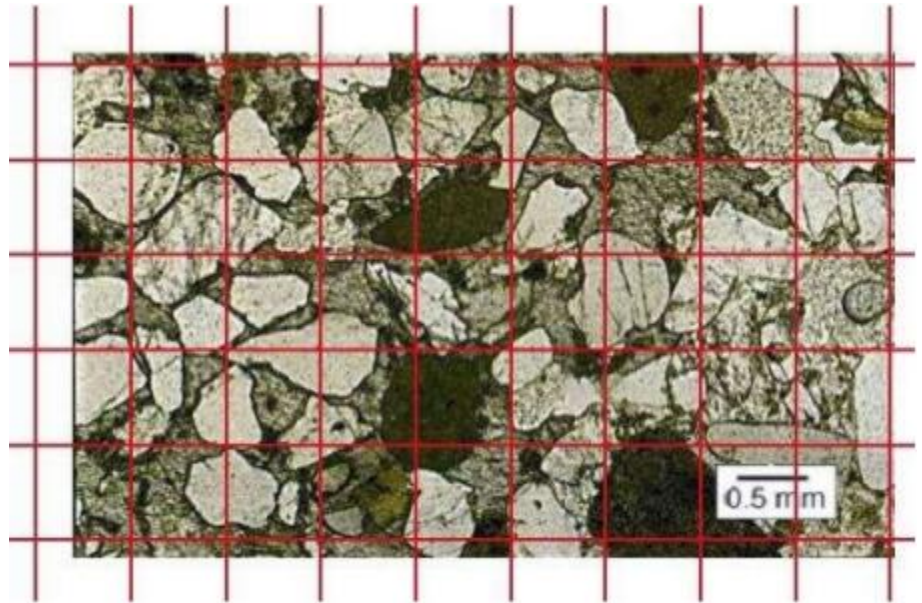
Count Squares Computer

So the relative number of points on object versus background equates to the fractional volume.

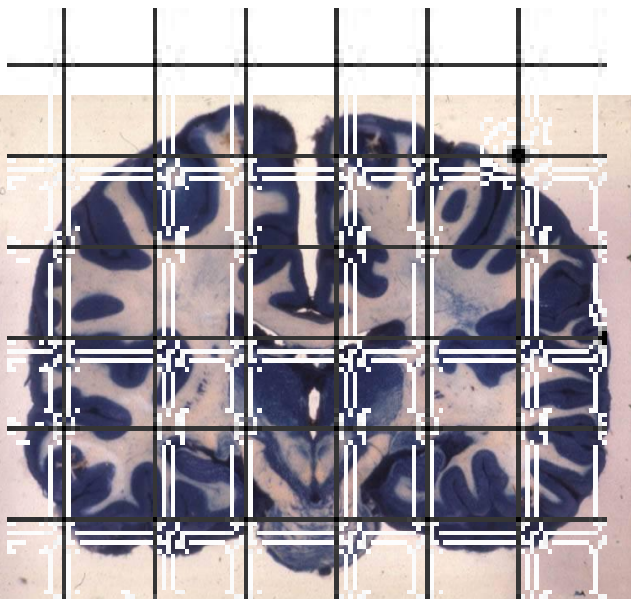
Point Counting Lattice Overlay ...Cheap!



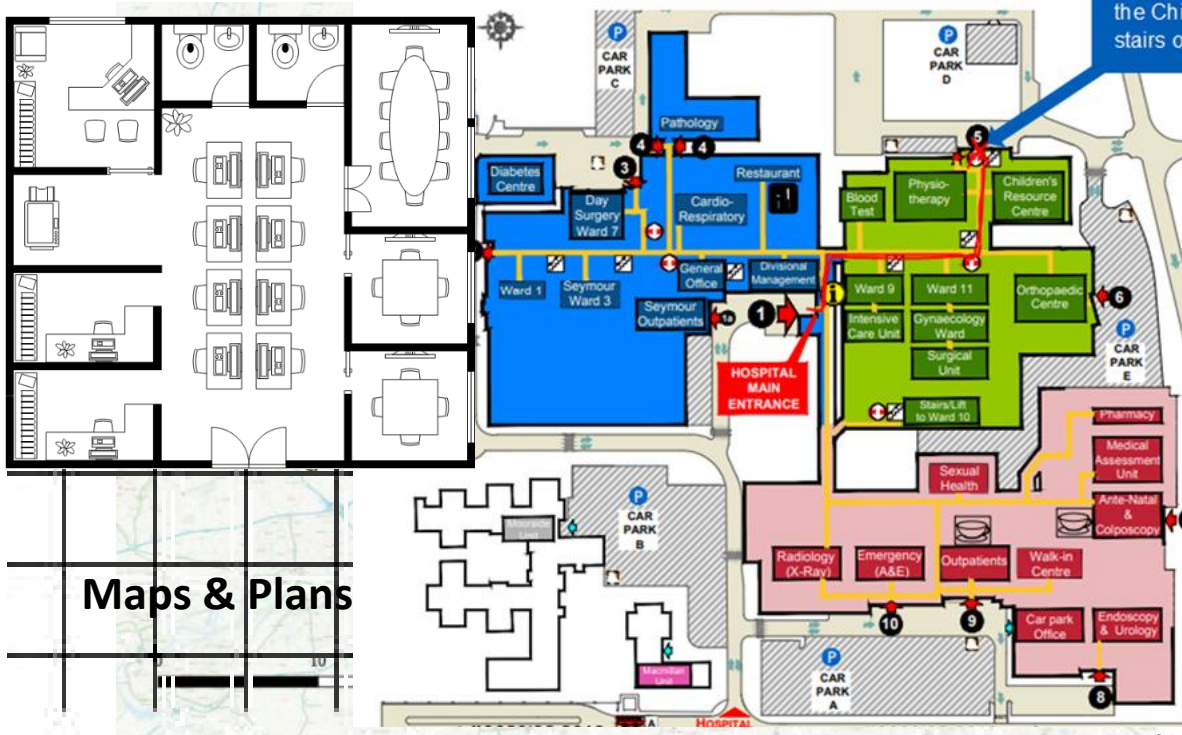
MM 1977!



Metallurgy & Geology

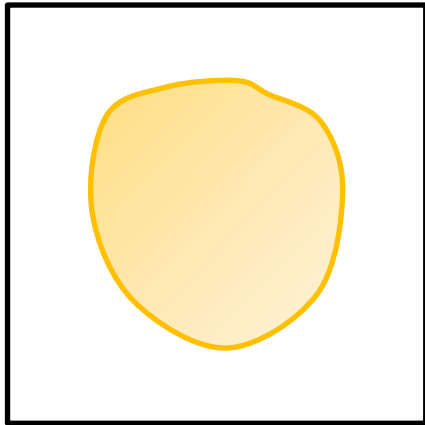


Brain – Grey v White Matter

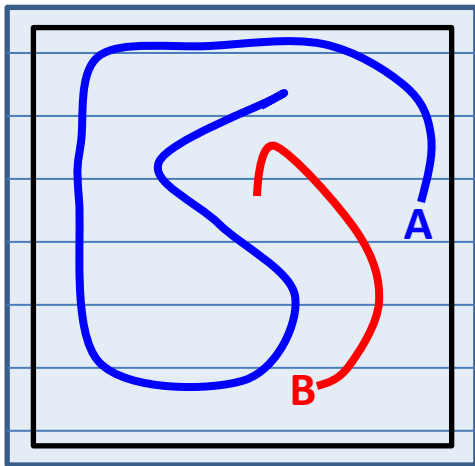


Maps & Plans

Surface Density



$$S_v = L \times 4/\pi$$



A= 14 cuts

B= 5 cuts

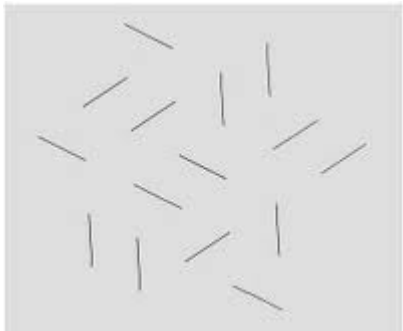
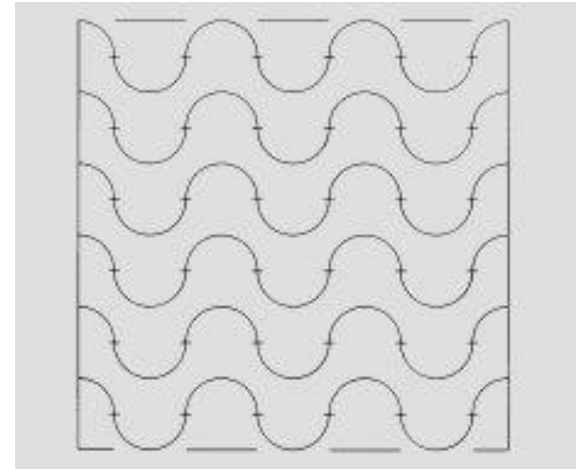
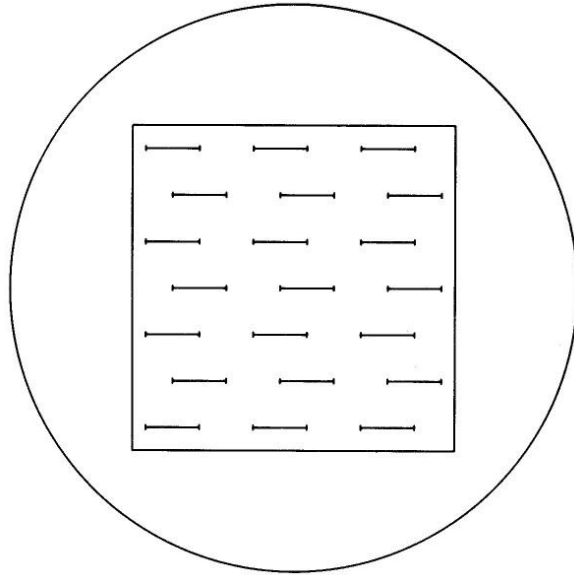
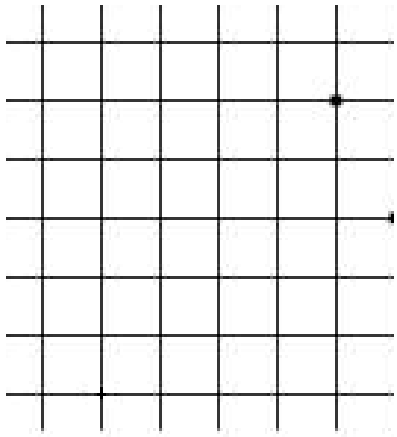
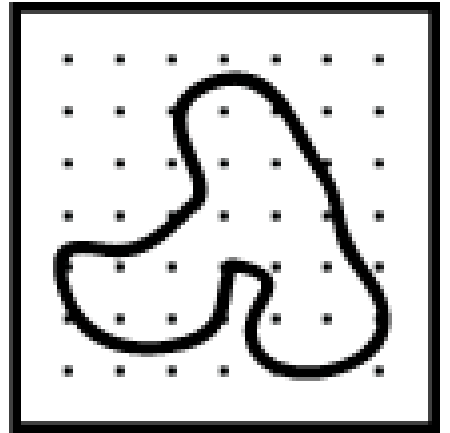
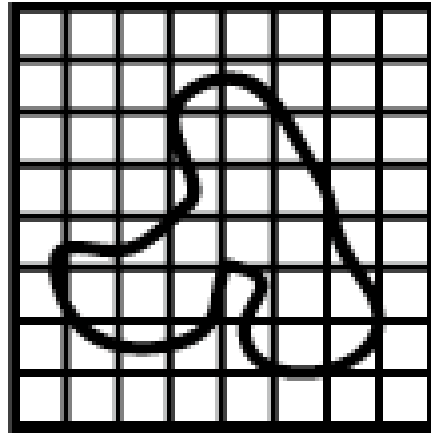
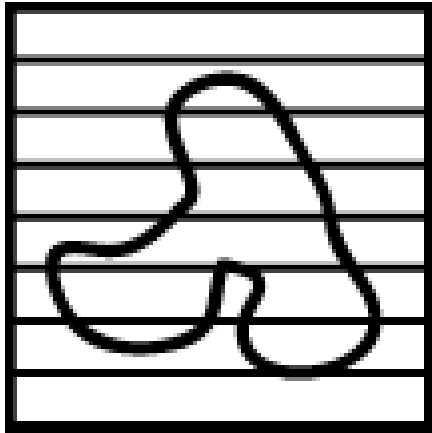
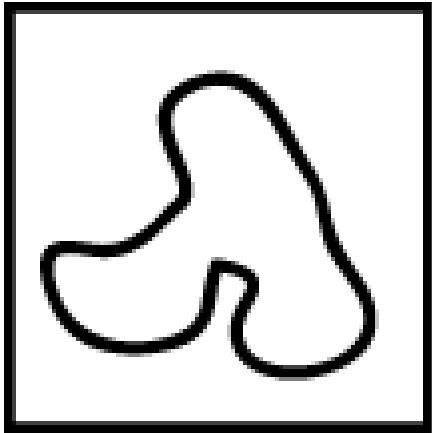
A is about 3 times longer than B !

$$S_v = 2 \times l_L$$

$$L = \pi / 2 \times l_L$$

Line Cut Lattice Overlay ...Cheap!

Buffon's Needle (1777)



Stereology Lattices (Probes)

... also available as Eyepiece Graticules

Use Hally formula for number of hits

How?



Numbers

... in tissue sections

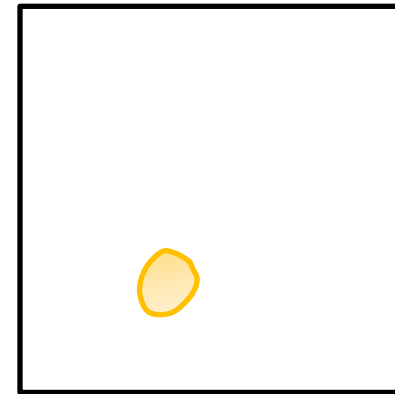
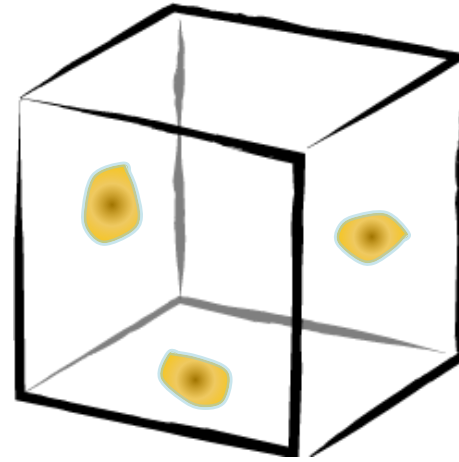
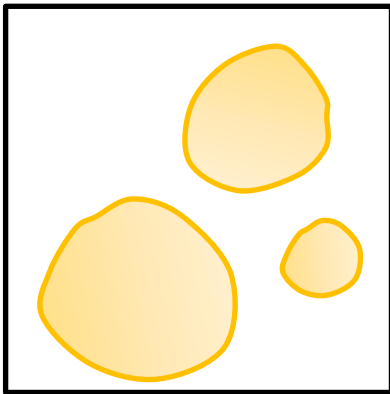
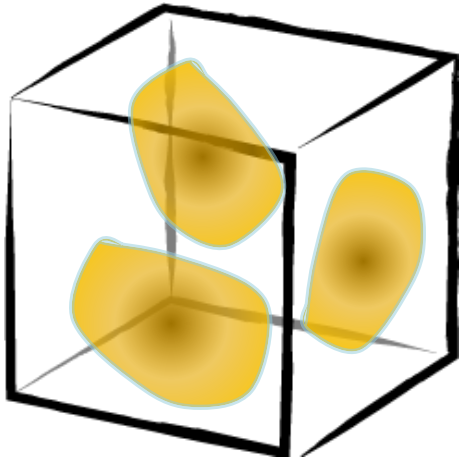


- N , N_v , N_A
- Counting Objects or Profiles ?
 - Loose cells/smears or sections?
- Depends on size and shape, section thickness
- Reconstruct
- Correction procedures (for Size, Shape, Populations)
 - Abercrombie (1946) $D = dx4 / \pi$
 - Schwartz-Saltykov (1958) Unfolding
 - Avoid: use Design Based Stereology



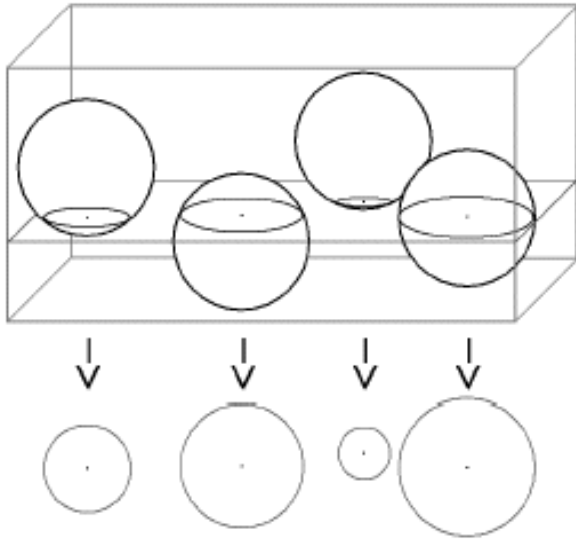
Although the answer is 3 items .. If they are smaller they are cut less often and apparently seem to be fewer!
So you need to measure their size as well.

Numerical Density

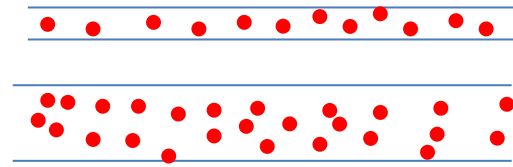


$$N_V = N_A / \bar{D}$$

Size corrections

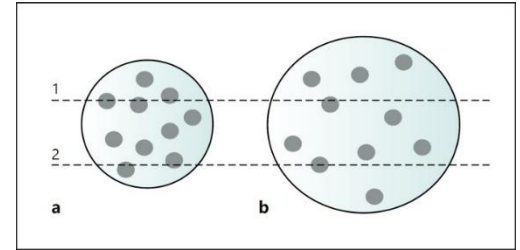


Section Thickness ... (Holmes)

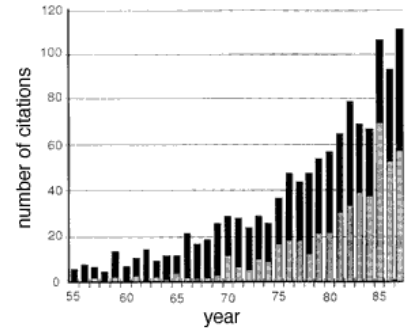


Change in Reference Volume

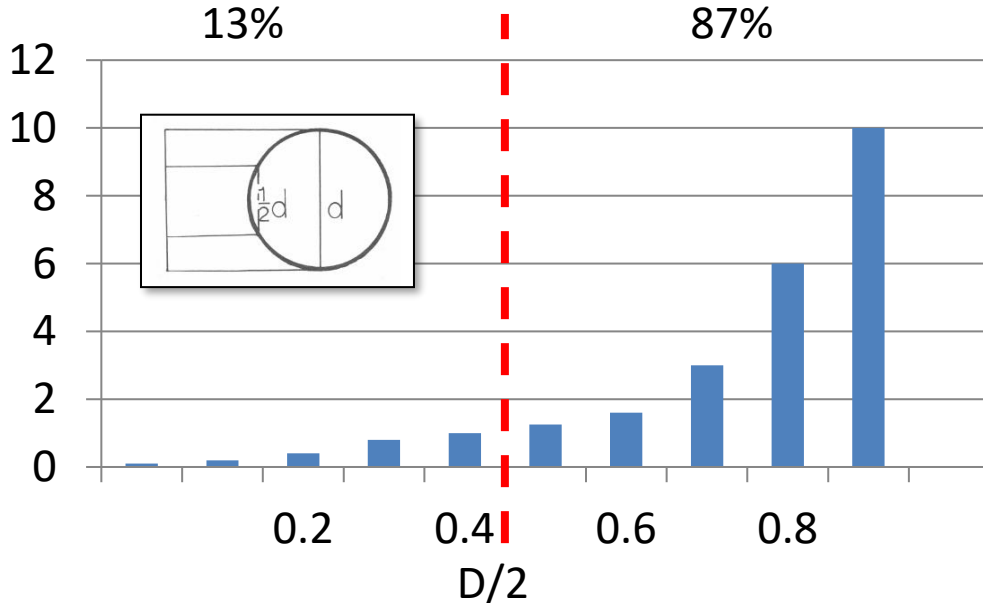
Monodispersed (Same Size)



...aargh!!



Profile Size



Abercrombie (1946) Correction Factor ...

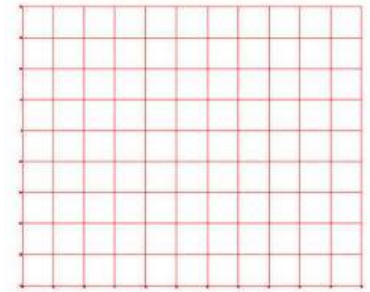
$$D = d \times 4/\pi \quad (\text{ie } \times 1.273)$$

Polydispersed (Different Sizes)

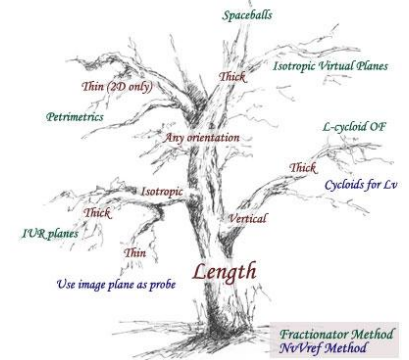
Schwartz / Saltykov Unfolding
 Shape (Ellipsoids, Cylinders, ...) !!

How?

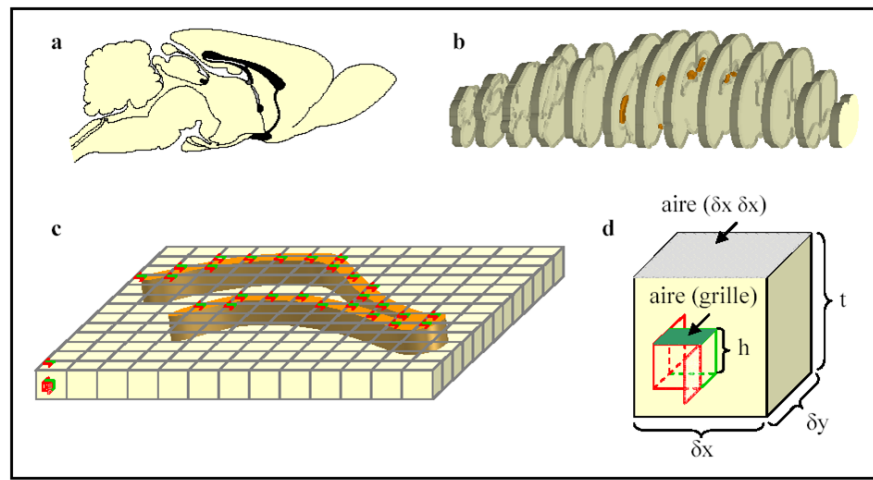
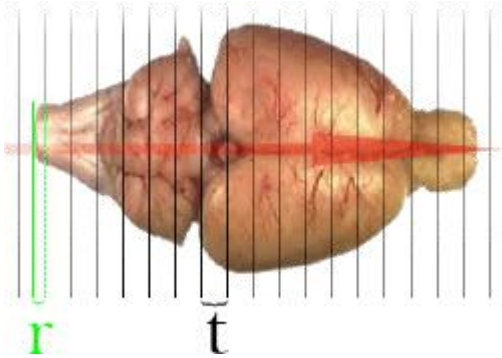
Stereology 2



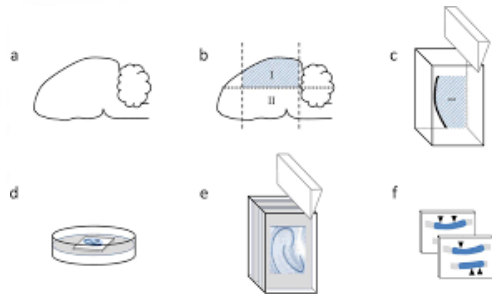
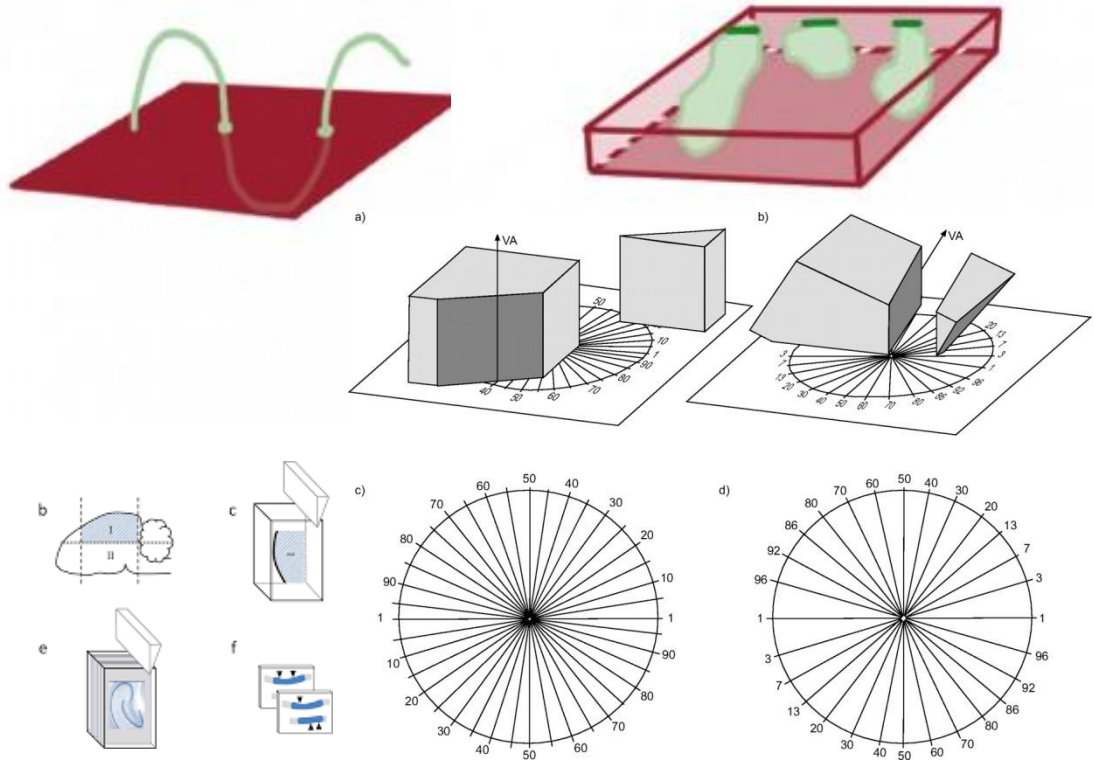
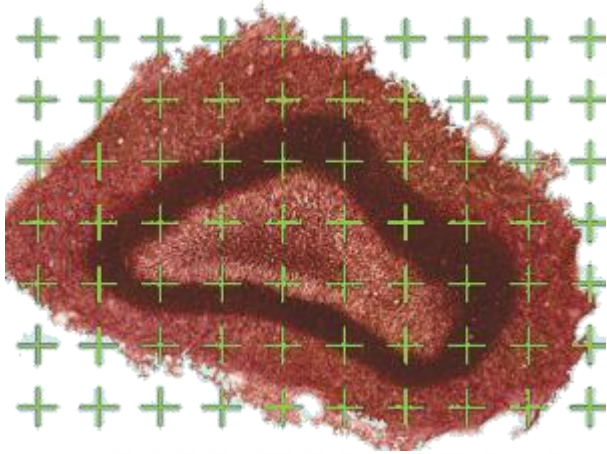
- New Design Based Stereology
 - Fractionator, Disector, Nucleator, Surfator, Proportinator, Selector, Rotator, Cycloids
 - Surface Weighted Star Volume
 - Unbiased Brick, Isotropic Fakir
 - Spaceballs, Petrimetrics
- Equipment / Design
 - Specialised Sampling; IUR, VUR, SRS sections
 - Thick sections, Optical sections
 - Unbiased Counting Frames



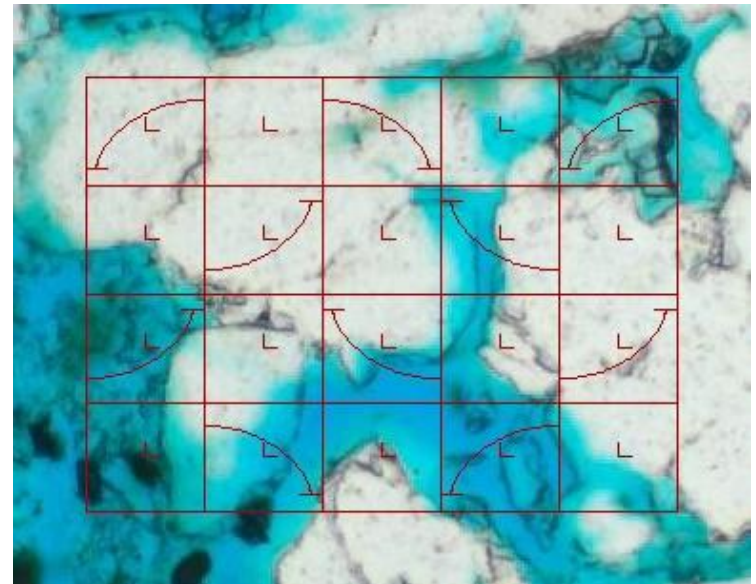
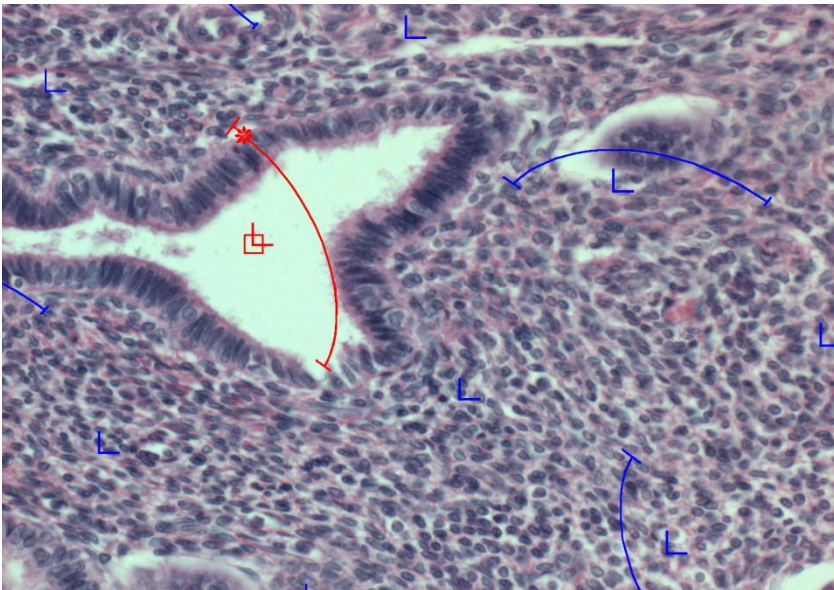
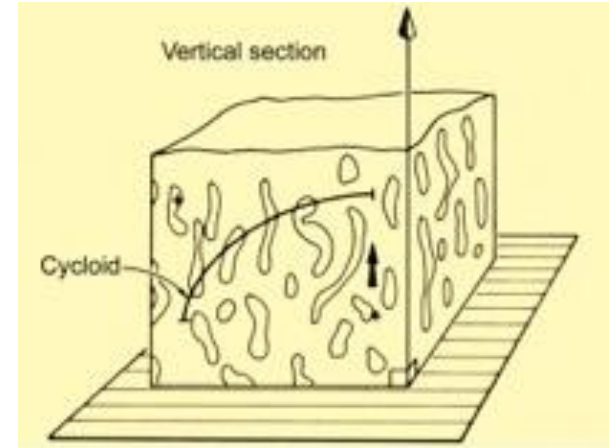
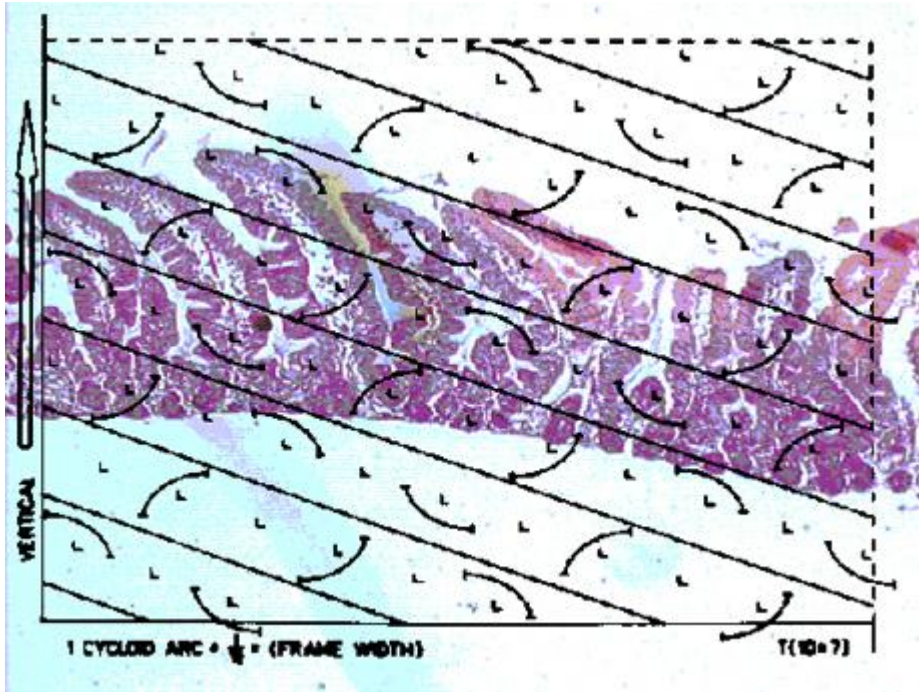
Cavalieri Method (1635)



Random start then serial sections, random slices, randomised probes ...

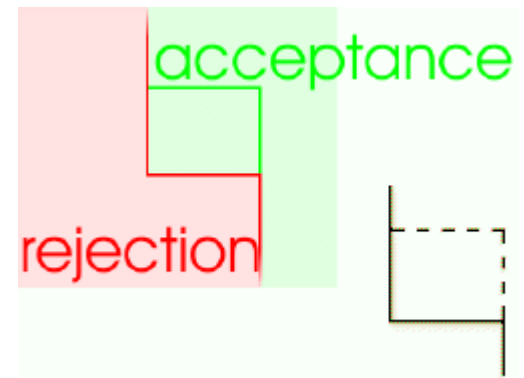


Cycloids for estimating Surface Area in a Volume



Unbiased Stereology for numbers

Gundersen



- **Measuring (see Picture 1)**

Since small objects are more likely to fit as complete objects within a measuring field it is best to remove this bias by measuring ALL objects within the frame and ALL objects hitting the dashed (allowed) lines but NOT those hitting the solid (forbidden) lines.

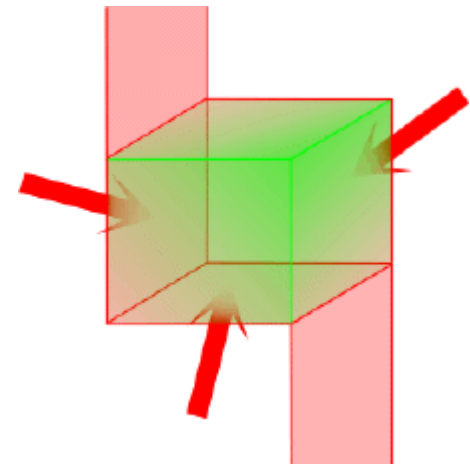
- **Counting (see Pictures 1 & 2)**

Objects allowed in the reference section (Picture 1 - A,B,C,D) are then checked in the next (look-up section (Picture 2)). If they DO NOT APPEAR in the look-up section these are the objects which are counted - ie object A ONLY.

Therefore one object occupies a volume equal to the frame area times the section spacing.

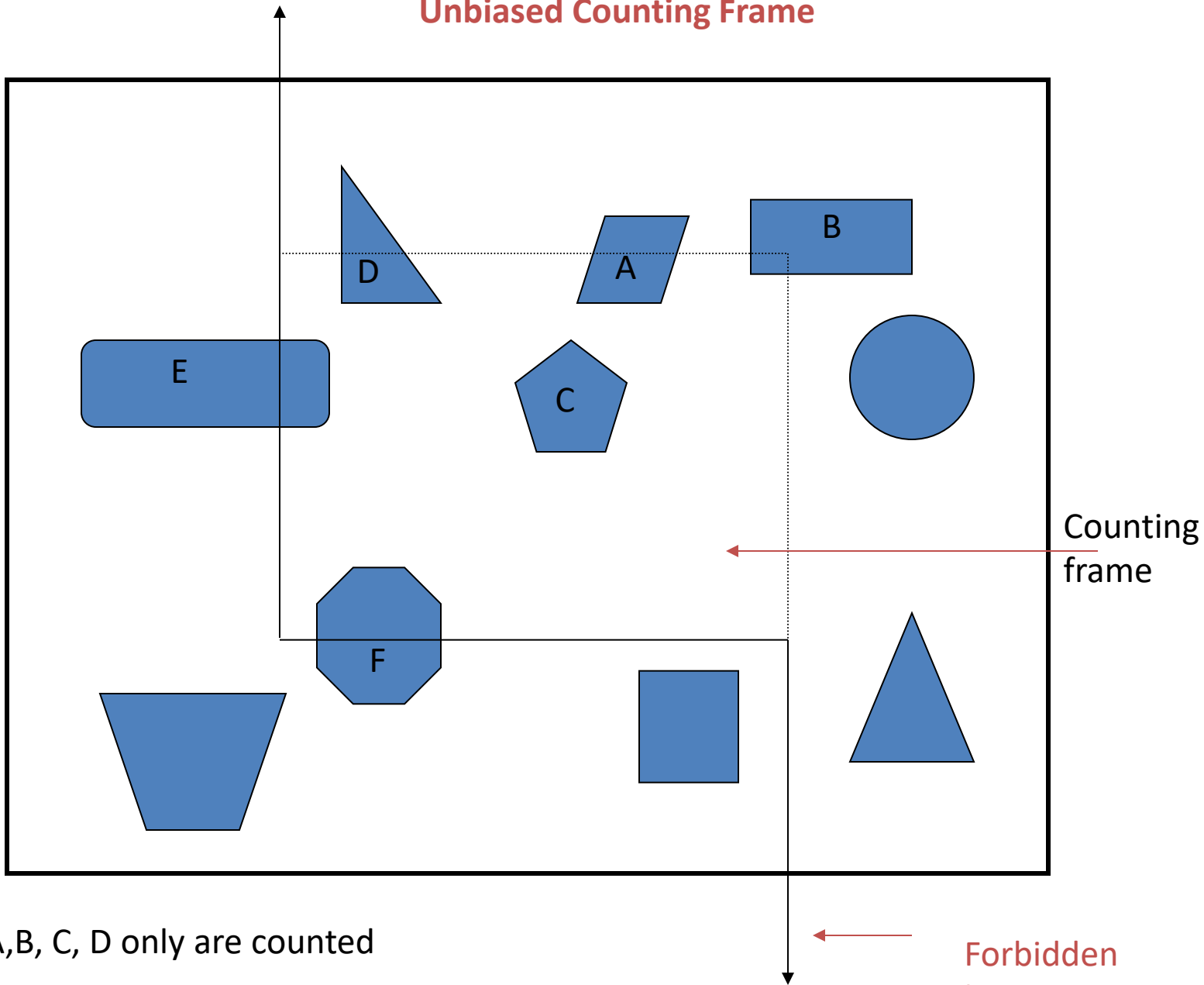
This is called the Disector Method.

- See: Howard CV, Reed MG (1998) Unbiased Stereology. RMs Handbook 41; Bios Scientific.



2D

Unbiased Counting Frame



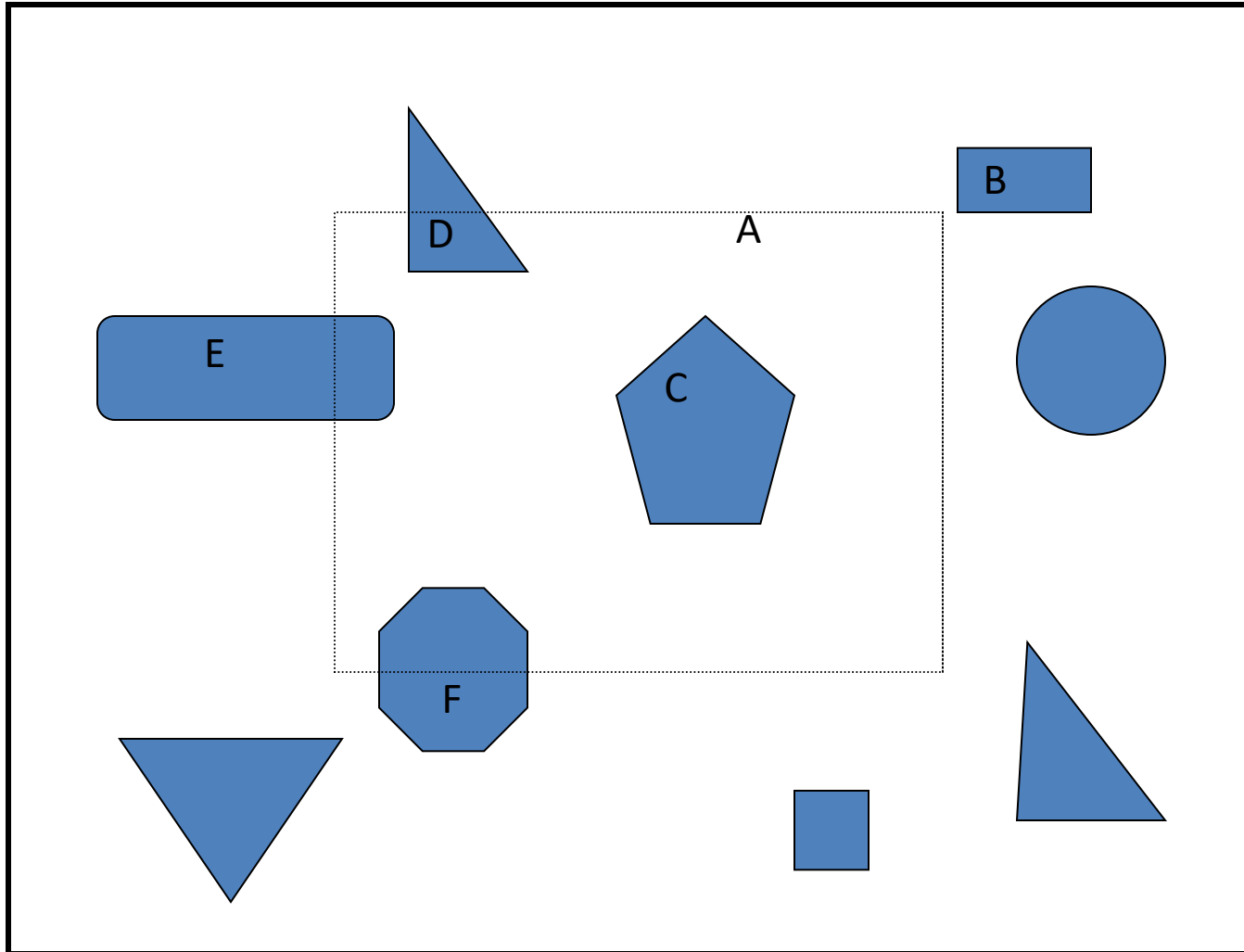
Objects A,B, C, D only are counted

1) Reference Section

Forbidden lines

2D

Disector method



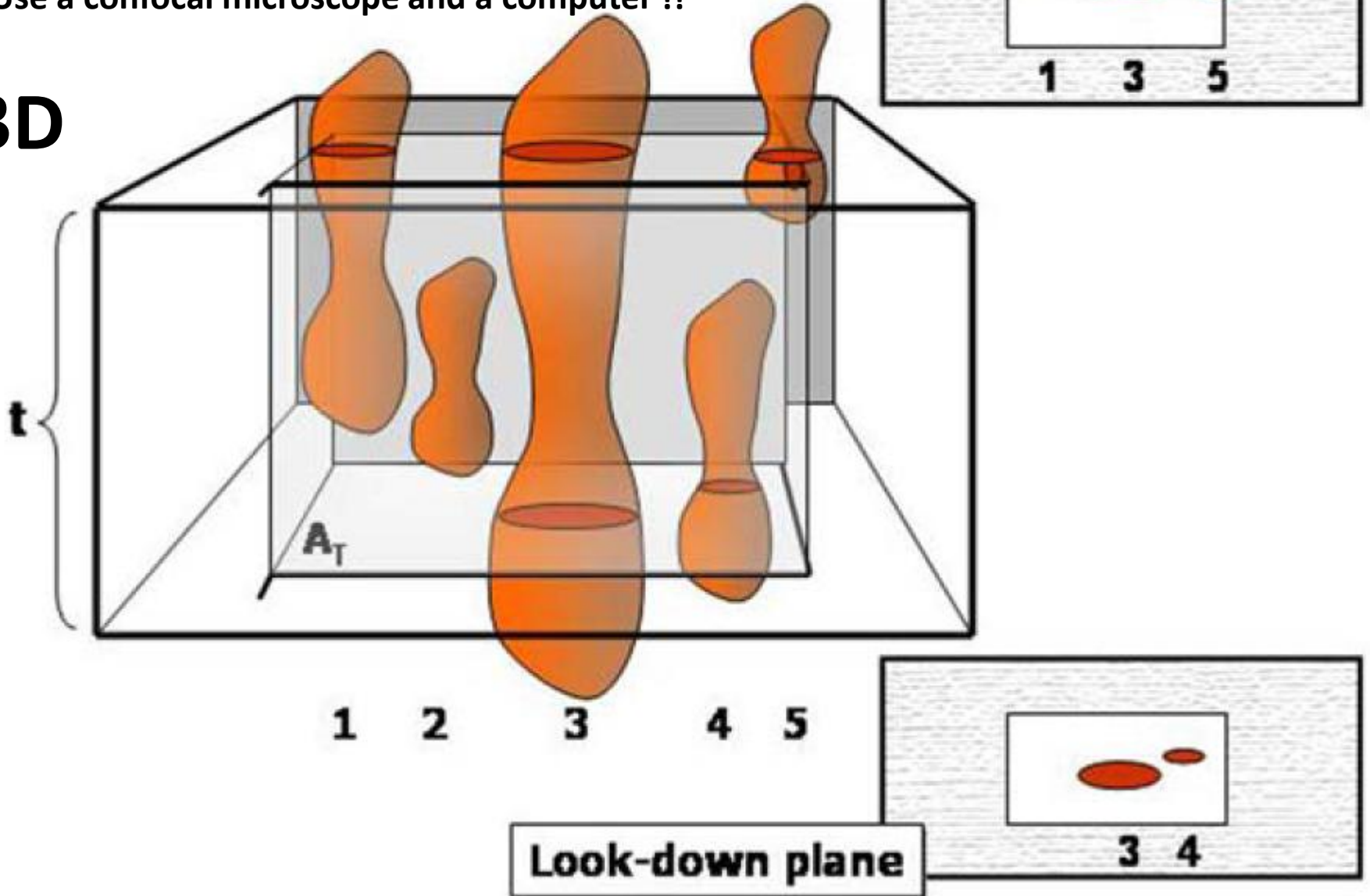
2) Look-Up Section

Of the objects counted in the reference section (A,B,C,D) only the objects NOT present in the look-up section are counted, ie only object A is counted.

Tedious to do with serial sections ...
alternative

Use a confocal microscope and a computer !!

3D



Commercial Stereology Software Packages

Free

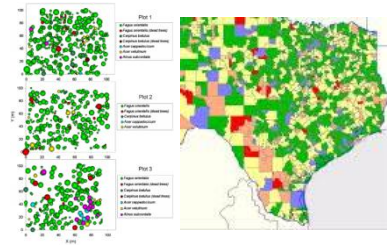
- Stepanizer
- MorhoJ
- 3Dslicer
- Imagepad



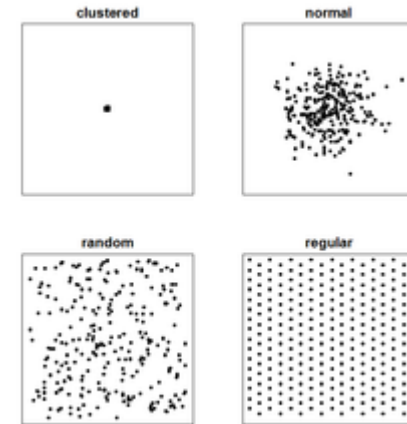
How?

Pattern Analysis

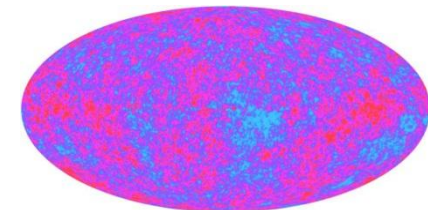
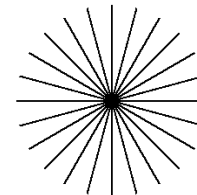
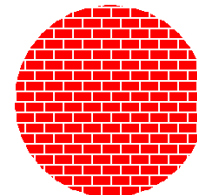
Measurements of 'Organisation'



- Location / Distribution / Spatial Arrangement / Association / Connectivity / Interaction
 - ???: Random, Regular, Clumped, Dispersed, Associated/Related
 - Distance: Nearest Neighbour, Mean free path
 - Grouping: Enclosed, Contiguity, Runs Test, SPAM
 - Autocorrelation
 - Tessellation / Joins / Overlay methods
 - Regional Density, Point Swarms
- Orientation / Branching
 - Dendritic methods (fields, segments, nodes)
 - Isotropic, Anisotropic



See: Uylings, Berry, Aherne, Underwood, Johnson, Sokal & Rohlf, James, Mahon, Cruz-Orive, Diggle, Unwin

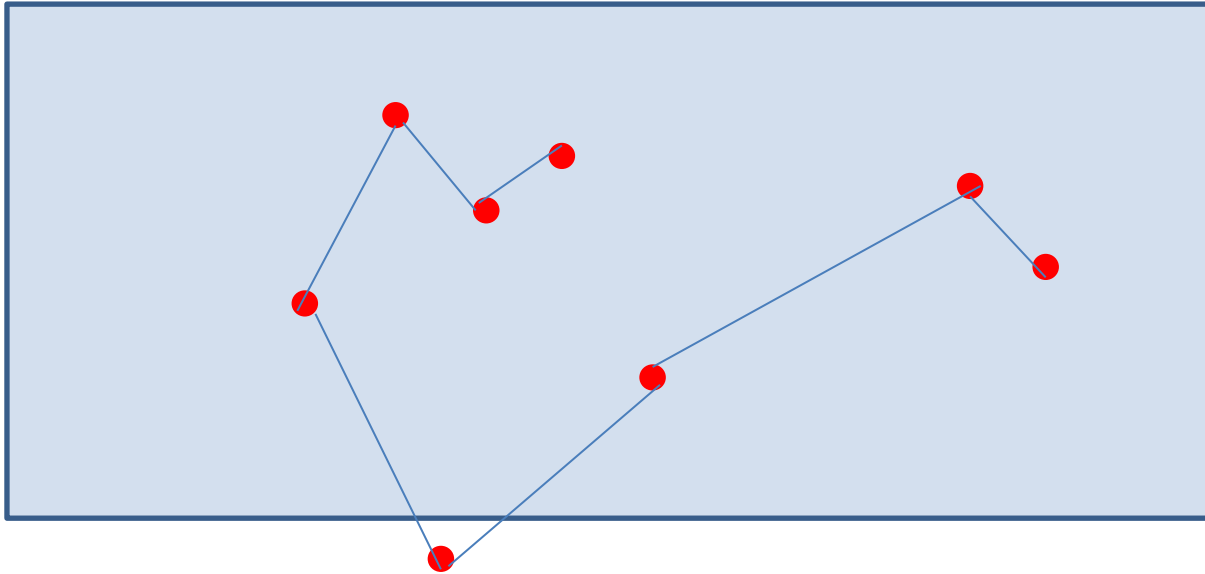


Nearest Neighbour Analysis

- Developed from Ecology studies (Poisson distribution)

Clark & Evans, 1954, Kendal & Moran, 1963

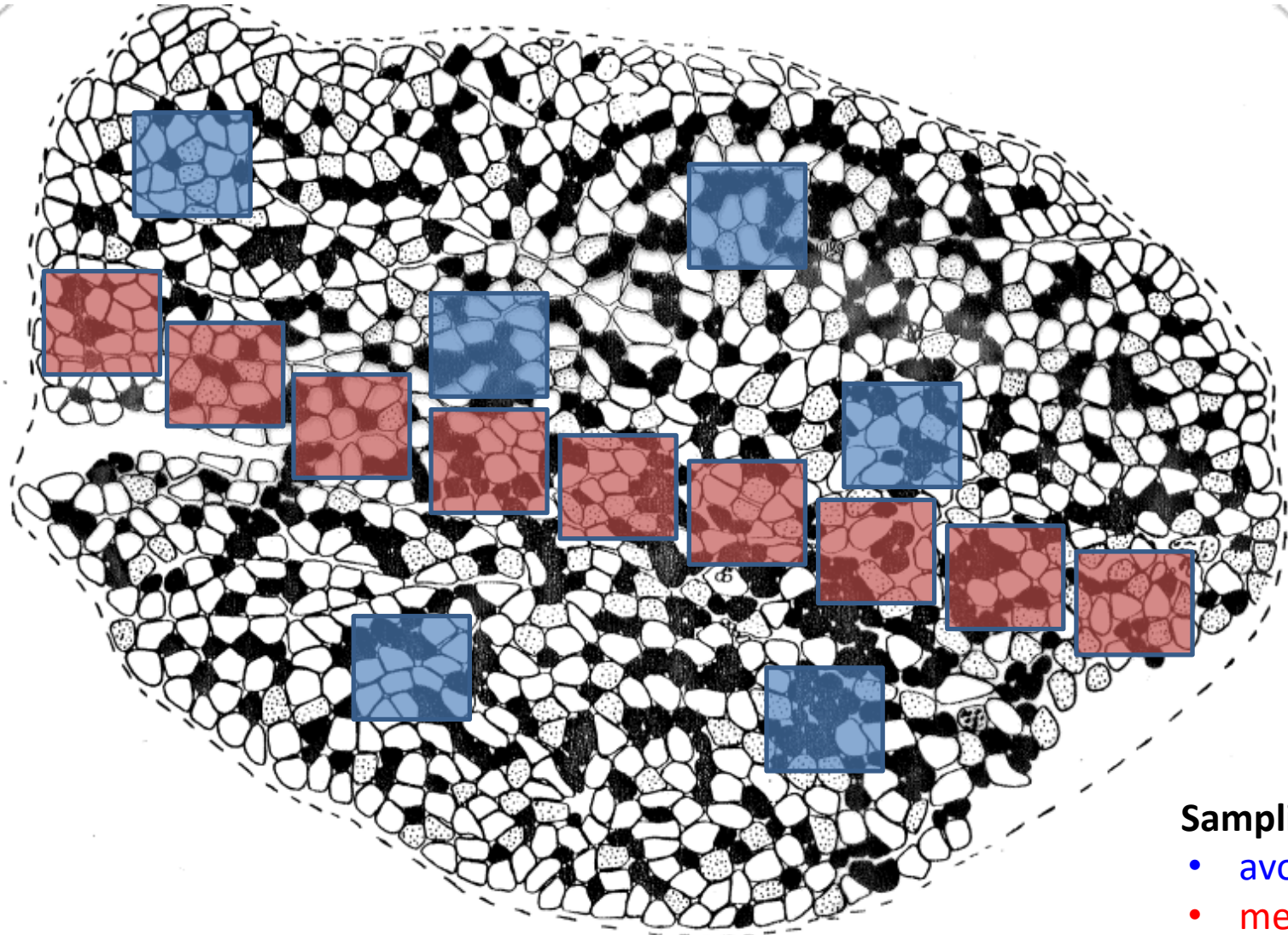
- Average distance between objects or Mean area / cell



- Begin at random cell / point

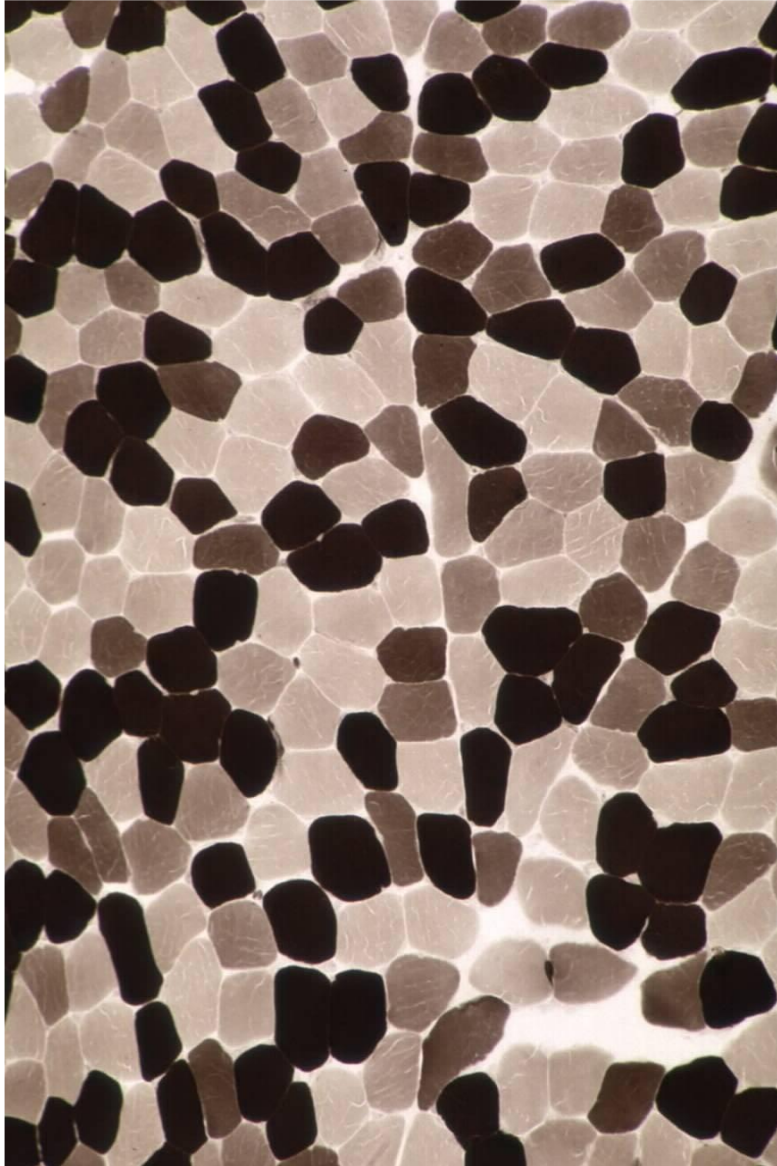
Problems: reflexive pairs / high densities

Homogeneity / Heterogeneity

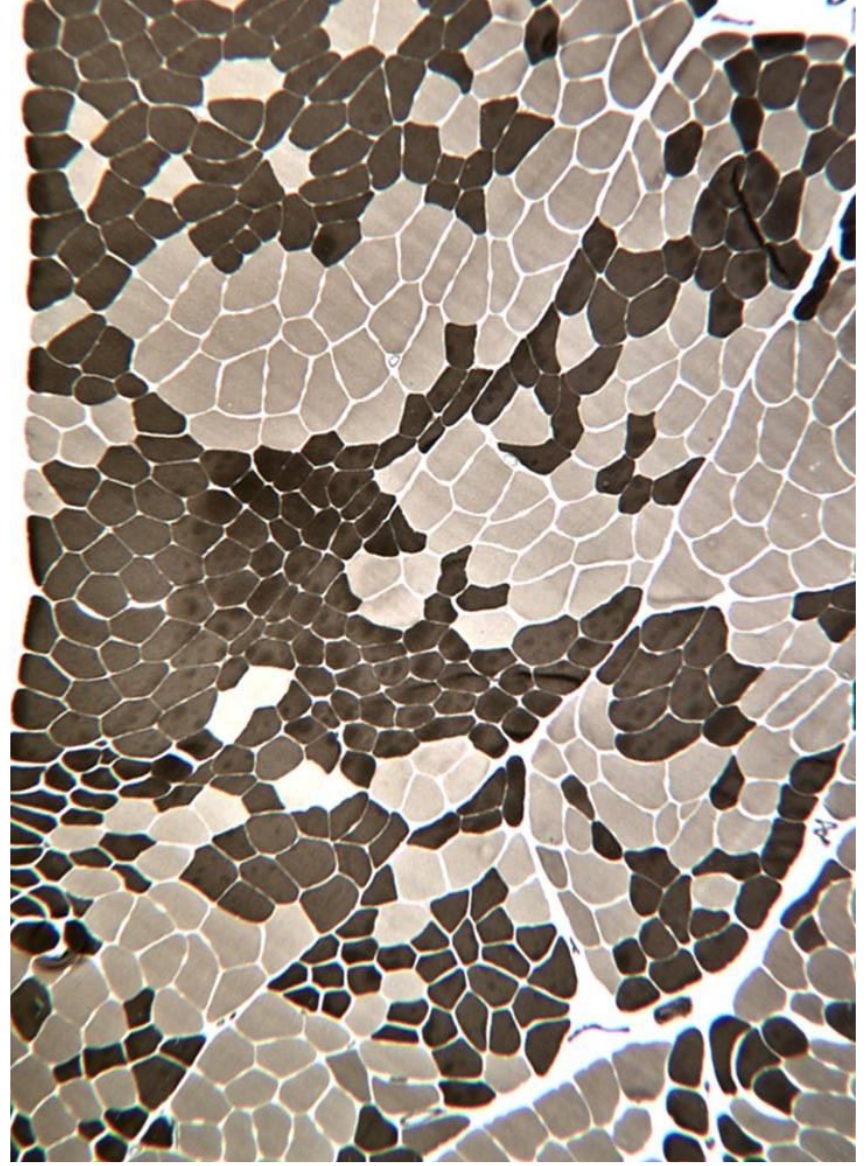


Randomness ?

Healthy Muscle



Diseased Muscle

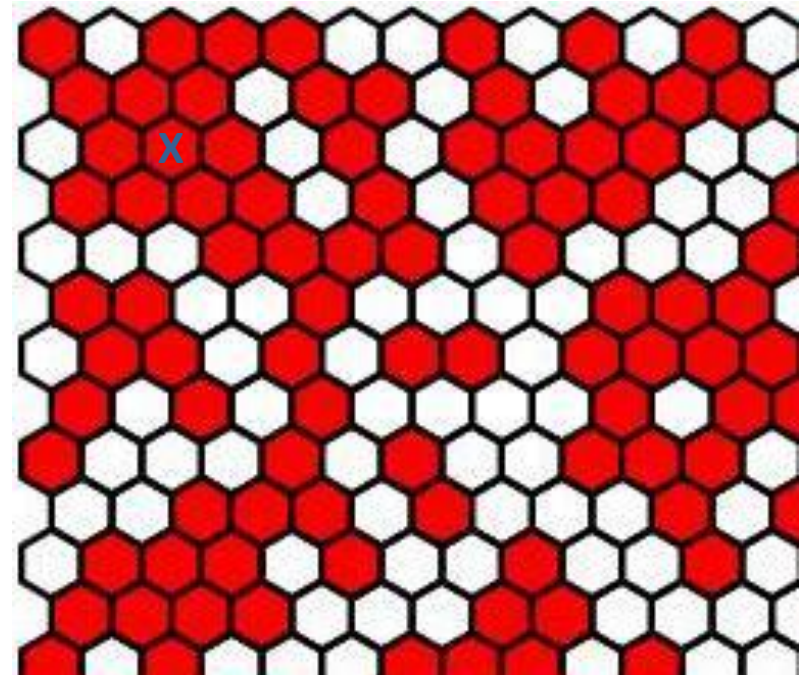


DISTRIBUTION

Enclosed “cell” method

- Observed versus Expected
- Predicted $E = Np^7 \pm SD$
- Depends on percentage occurrence
 - 30% $R = 0$ enclosed
 - 50% $R = 1$
 - 70% $R = 8$
 - 90% $R = 50$

Johnson, 1973



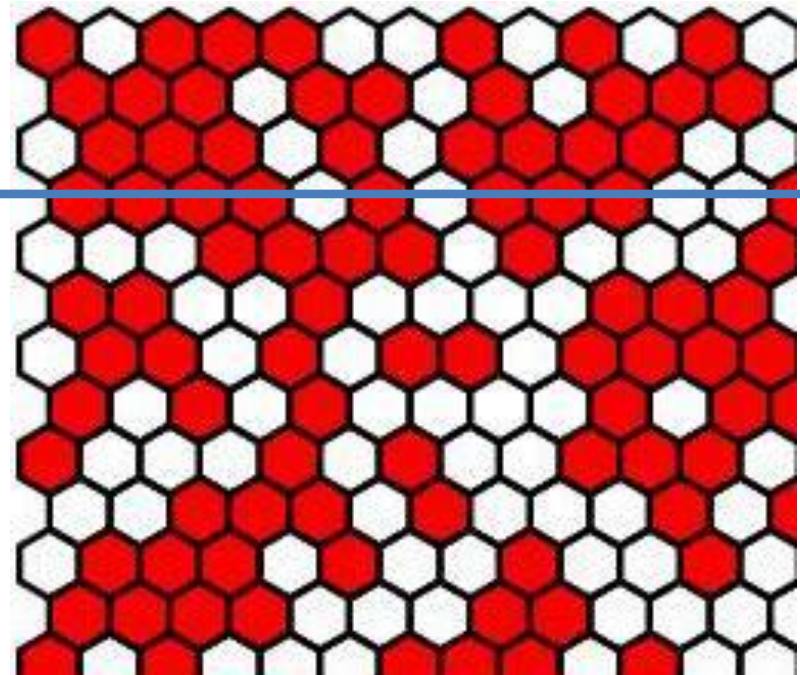
DISTRIBUTION

Runs Test

- RRRR W RRR WWWW R WW
- R W R W R W R W R W R W R W R
- $N = 15, n_1 R = 8, n_2 W = 7$
- $\text{Exp } F = [2 \times (n_1 \times n_2 / n_1 + n_2)] - 1 = 6.5$
- $T = (F - \text{Exp } F) / \text{SD}$
- Distribution IS Random
- Distribution IS NOT Random
- Predict Runs eg 100 cells 60% R = 47 +/-5 runs

$r = 6$

$r = 15$



Sokal & Rohlf, 1973

DISTRIBUTION

Run Lengths (Clumps)

Roach, 1968

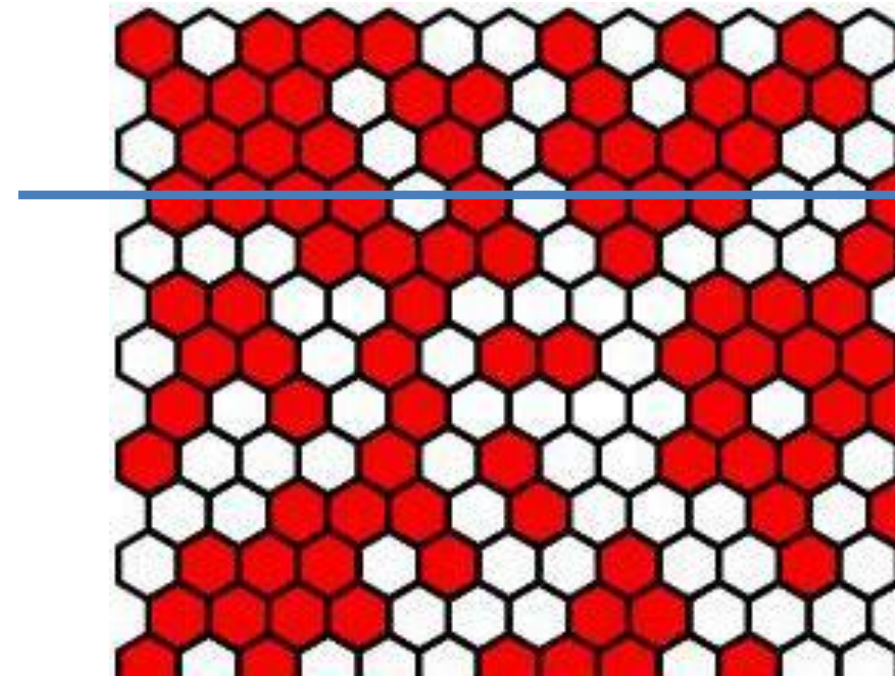
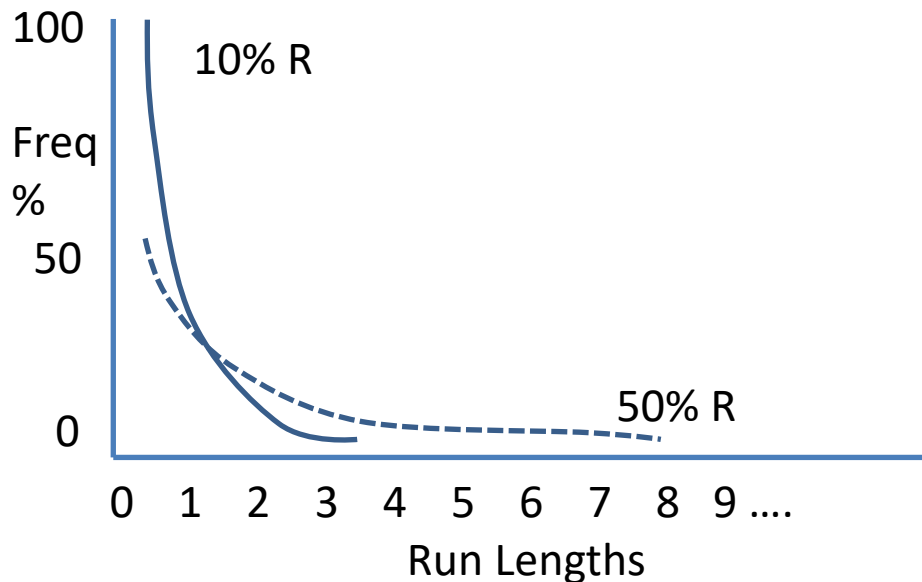
- RRRR W RRR WWWW R WW
- R W R W R W R W R W R W R W R
- Predict expected run lengths for a random distribution

Avlength R = 2.66

Avlength W = 1.00

$$Cl_R = N_R + P_{RL^{R-1}} \times (1 - P_R)^2$$

$$Cl_W = N_R \times P_R \times (1 - P_R)^{LW}$$



DISTRIBUTION

Contiguity

Underwood, 1970 Gurland, 1975 James, 1980

- Apply test line and look at intersections with boundaries
- Need to know length of test line and

$$\begin{aligned} N_{RR} &= R_R \\ N_{WW} &= W_W \\ N_{WR} &= W_R_W \end{aligned}$$

- Use stereological SV formulae modified for 2D

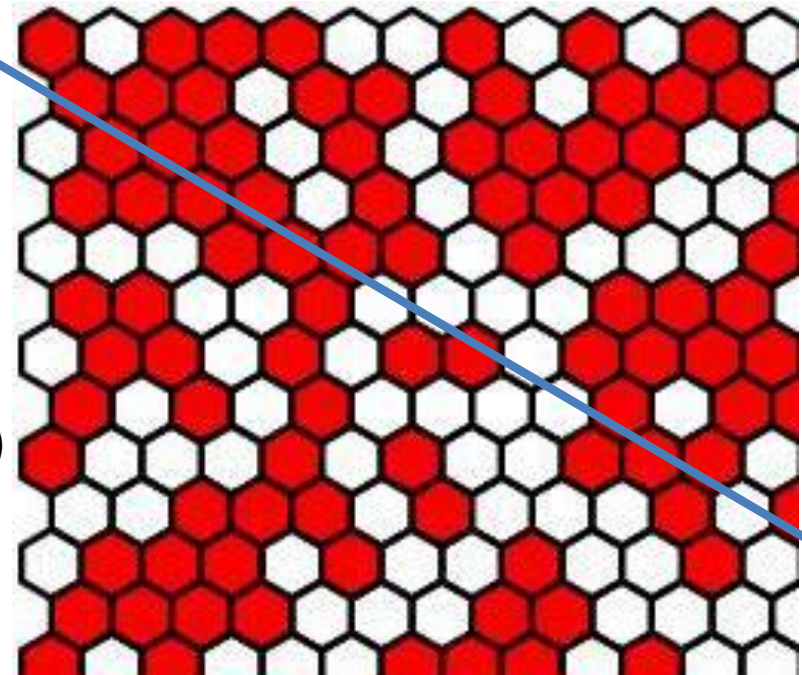
- $L_A = \pi/2 \times P_L$

- Estimate Interface lengths for

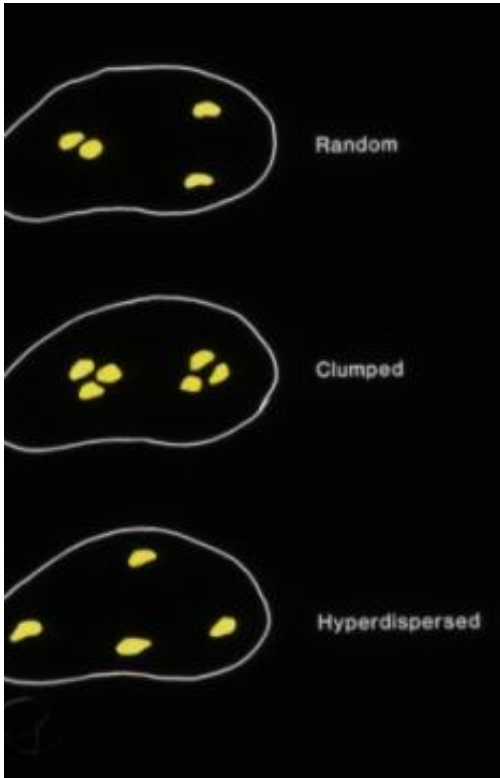
$$\begin{aligned} LA_{RR} \\ LA_{WW} \\ LA_{WR} \end{aligned}$$

- **Index of Contiguity**

$$C_{RR} = LA_{RR} / (LA_{RR} + LA_{WR})$$

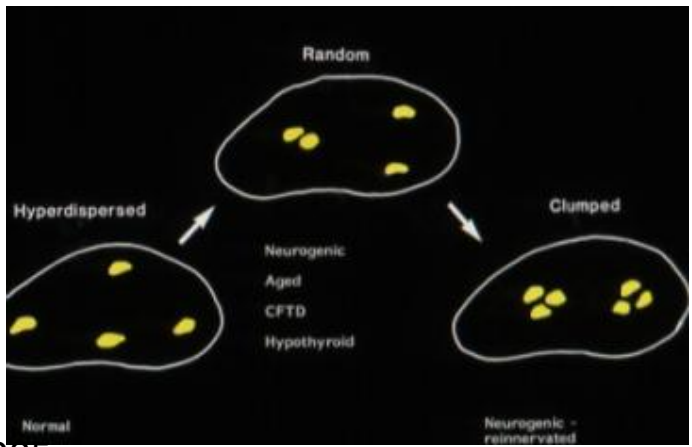
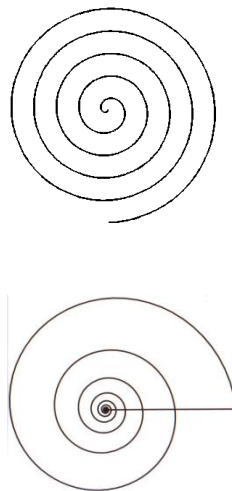
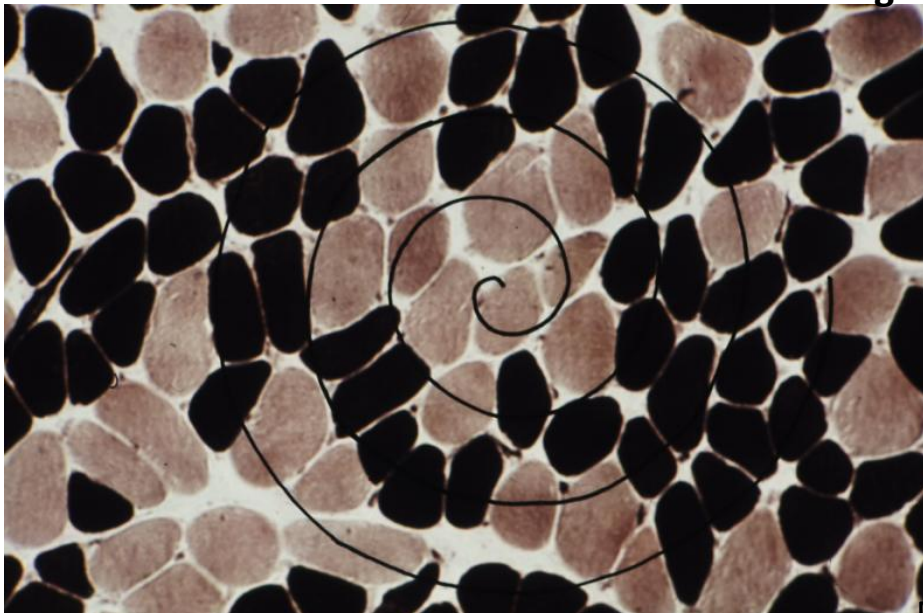
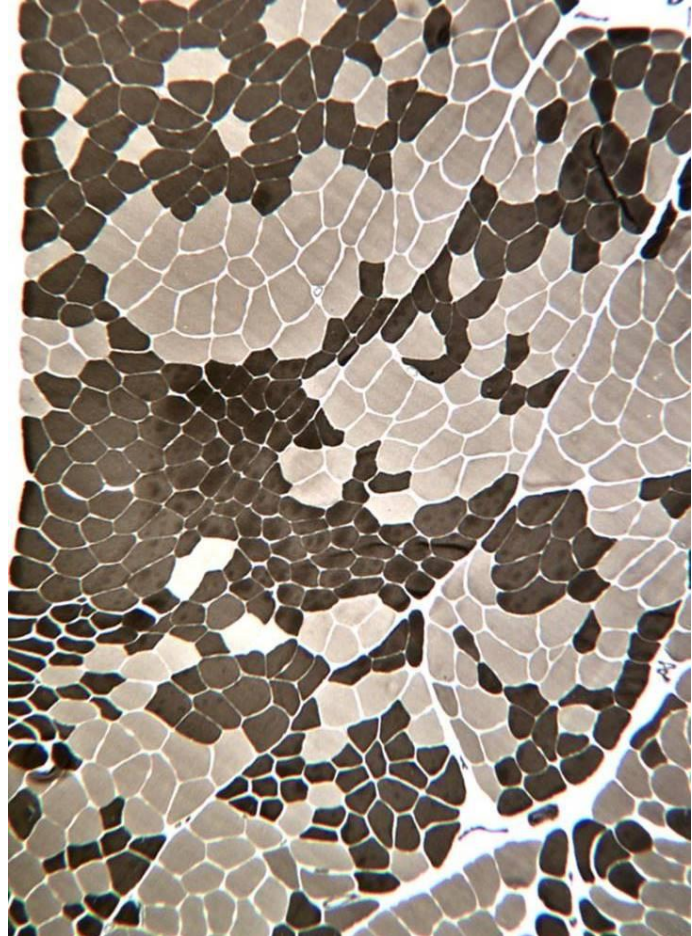


DISTRIBUTION



Spatial/Spiral Pattern Analysis of Muscle

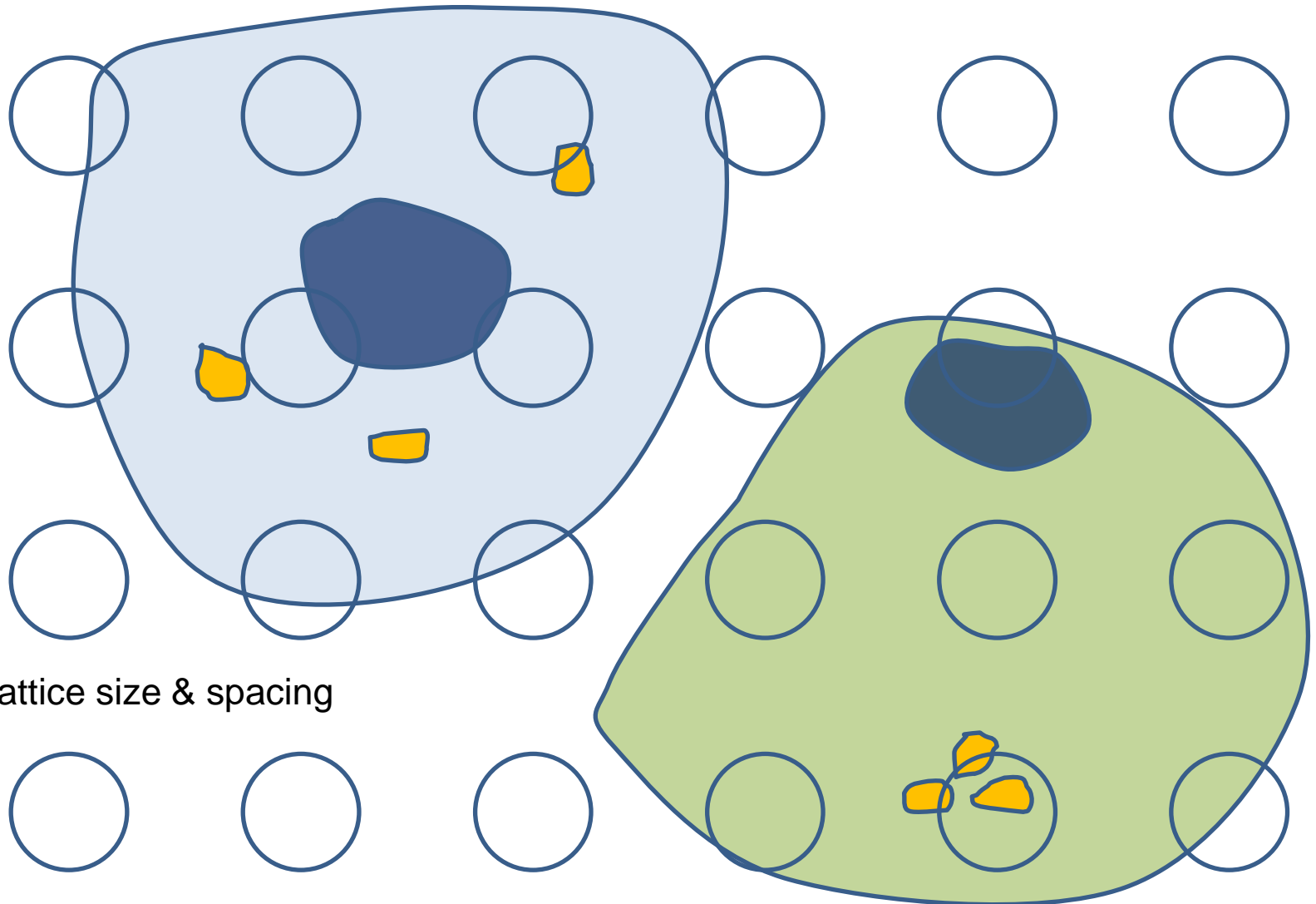
Runs Test-Run Lengths-Contiguity



Measure of **Association** – Circle Overlay Method

Cruz-Orive, 1976

eg: Eccentric nuclei, organelle clumping or autoradiography (*Williams, 1977*)

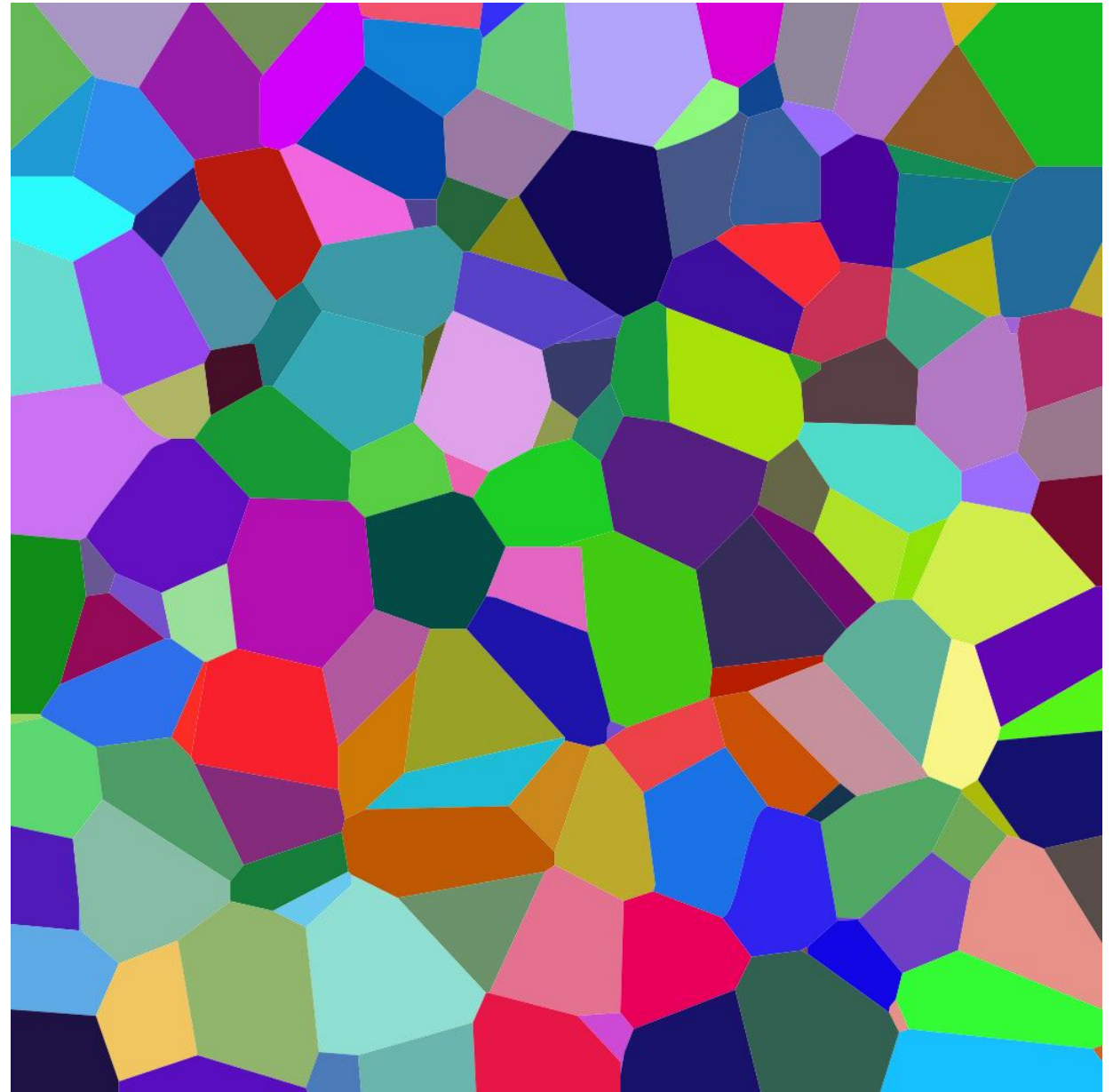
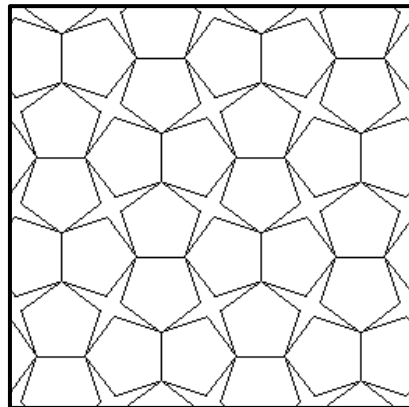
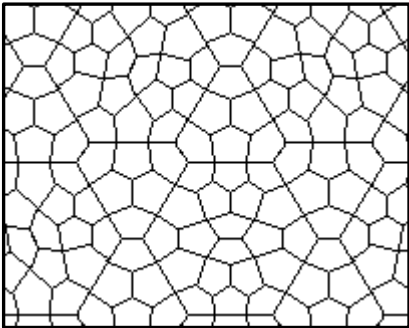
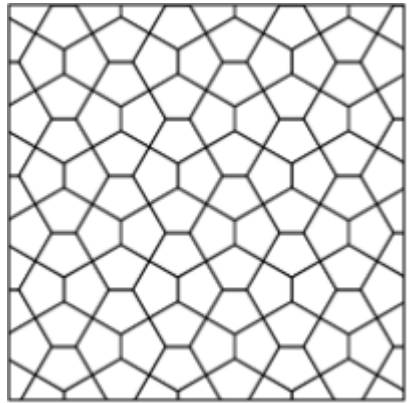


Problems: lattice size & spacing

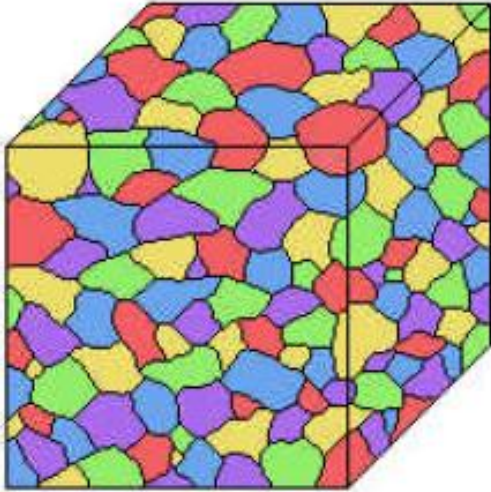
Shows that in the pathology example to the right, more likely to get circles overlapping groups of orange objects, normal (left) orange associated with nucleus.

Tessellation

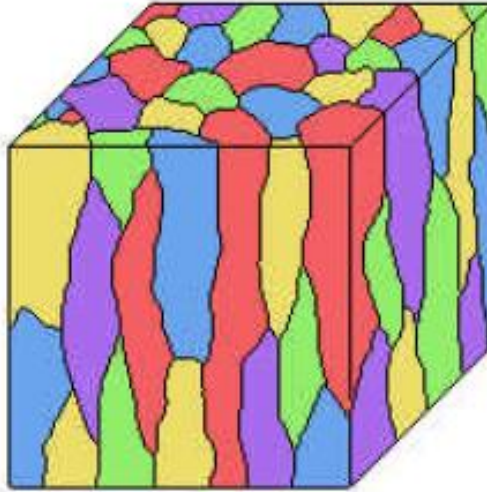
Apply lattices and use mathematical concepts of “Lattice tessellation of congruent domains”



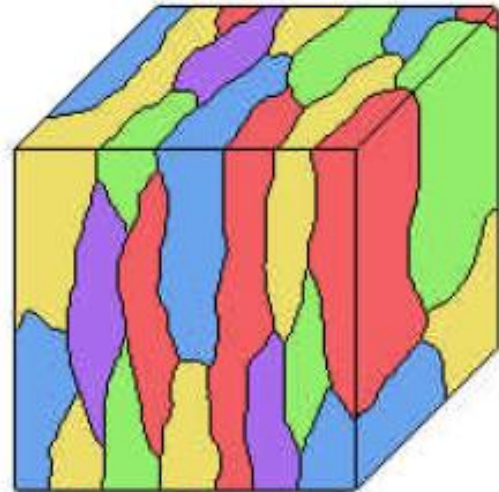
ORIENTATION



a

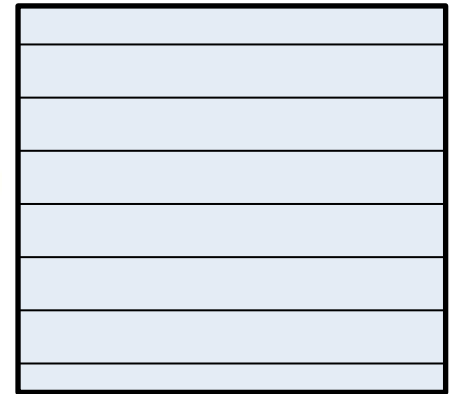


b

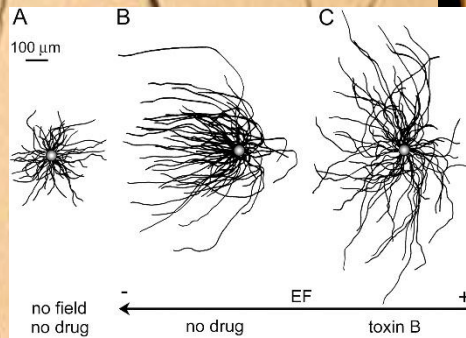
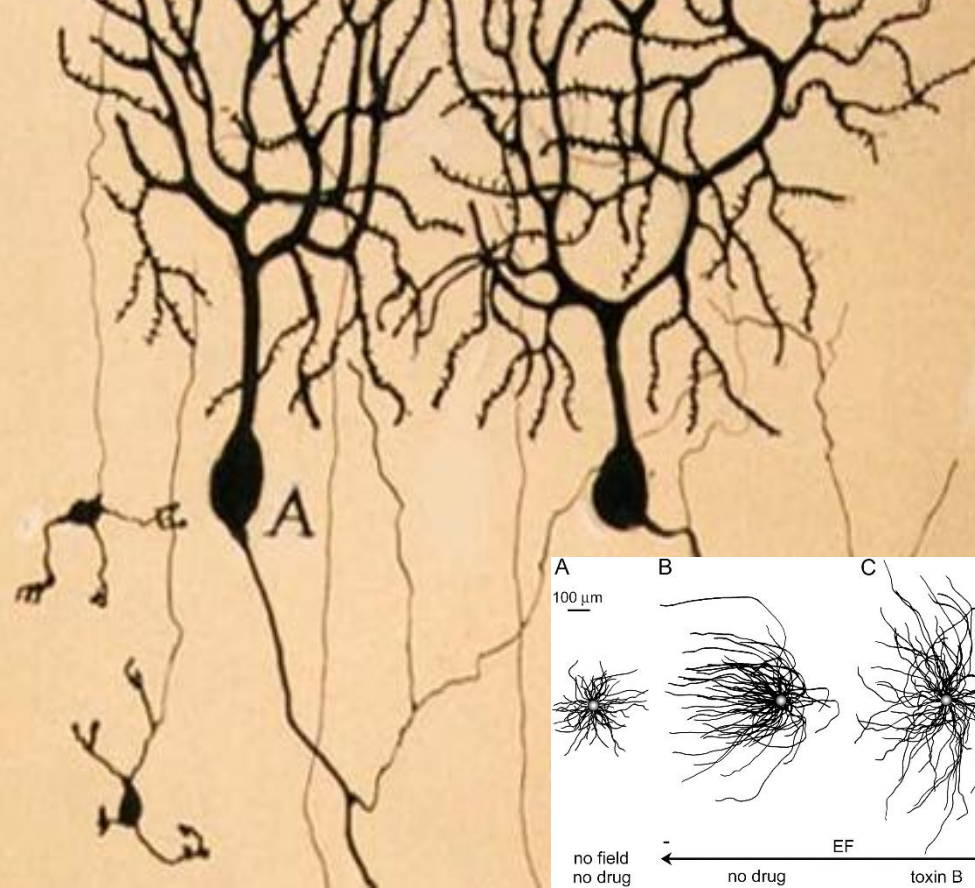
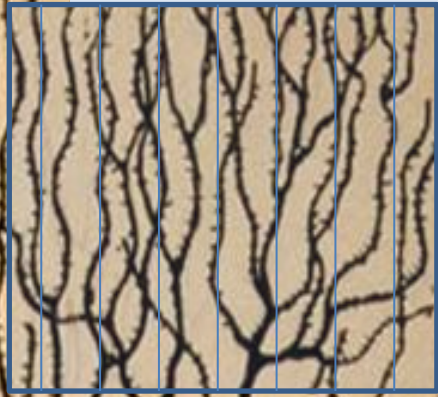


c

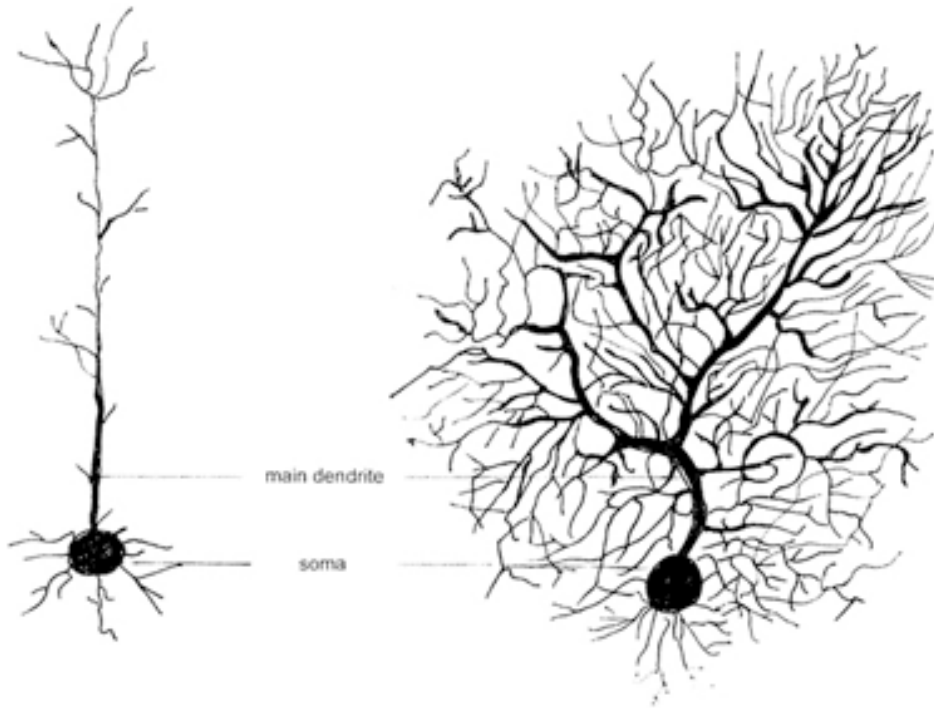
Apply line lattice



ORIENTATION

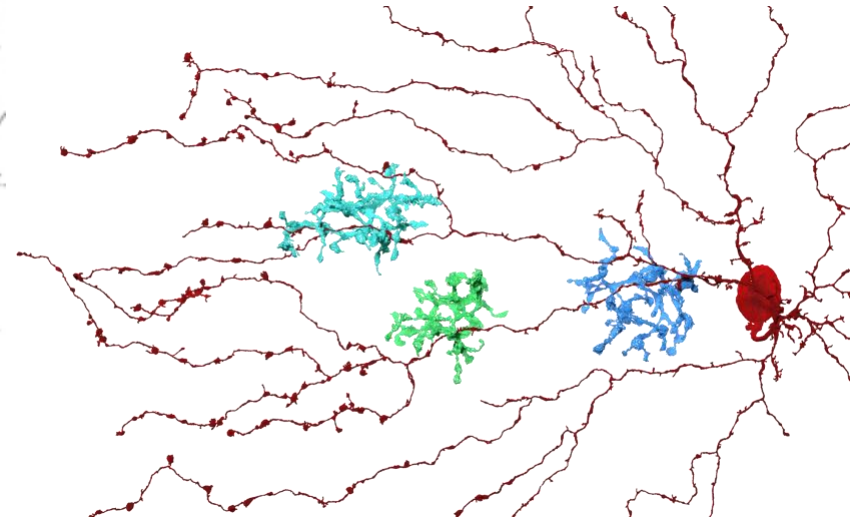


BRANCHING



Sparse growth of dendrites
in an aging, inactive brain

Typical dendritic growth in an active brain



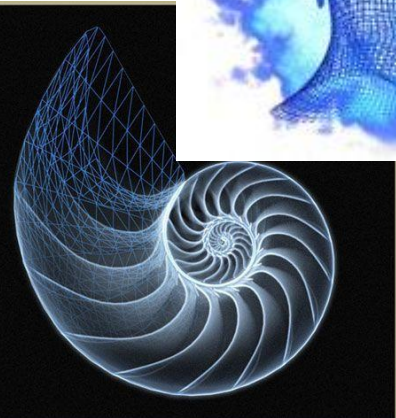
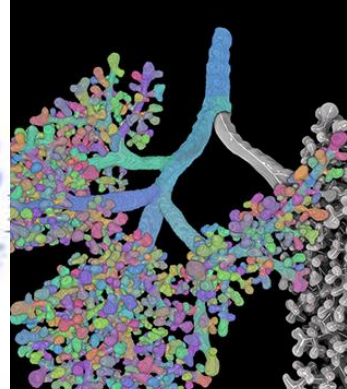
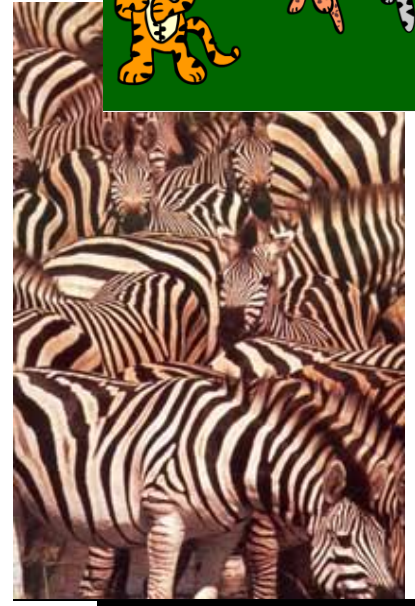
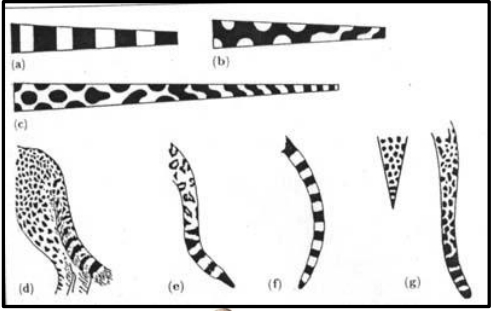
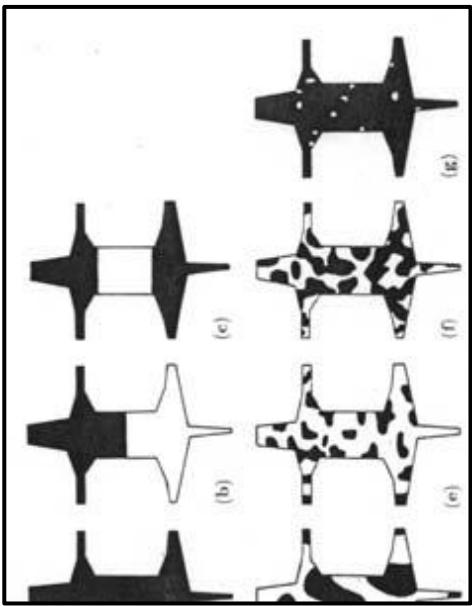
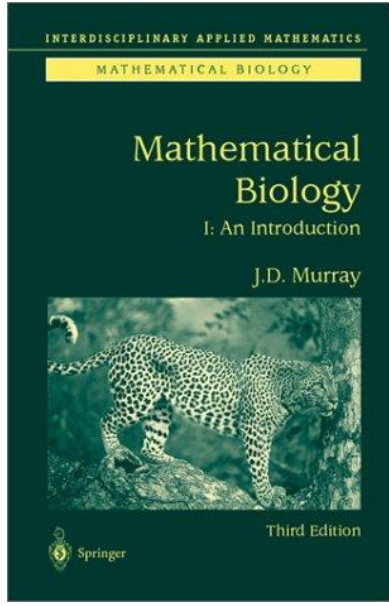
Topology

- Arbor or Tree analysis
- Vertex / Branch / Segment
- Bifurcations / tri .. / multi ..
- Angles

Berry, 1980s

Mathematical Modelling of Topography & Development

1952 Alan Turing , 1980 JD Murray



$$U = \mu_p^0 \theta_p + \mu_p^0 \theta_p + \chi \theta_p \theta_p$$

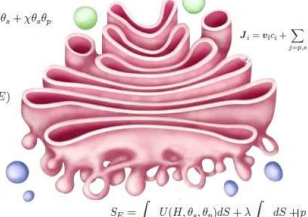
$$J_i = \nu_i \alpha_i + \sum_{j=p,\alpha} \alpha_{ij} \nabla_S \theta_j + D_i \nabla_S \alpha_i$$

$$\nabla_y \cdot (y \cdot \nabla u_n E)$$

$$\Delta_S (bH)$$

$$\sum_{j=p,\alpha} \alpha_{ij} \nabla_S \theta_j$$

$$S_E = \int_{\partial \Omega} U(H, \theta_p, \theta_p) dS + \lambda \int_{\partial \Omega} dS + \int_{\Omega} dV$$

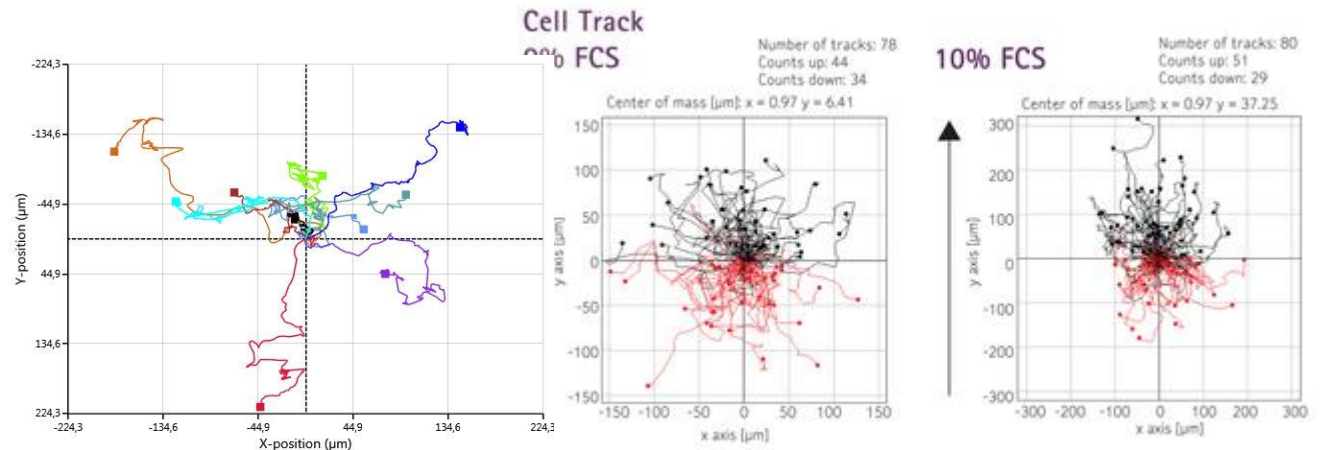
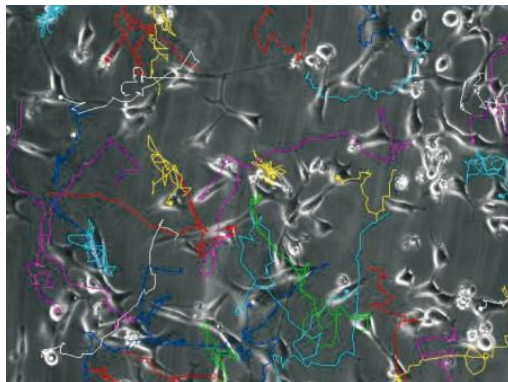
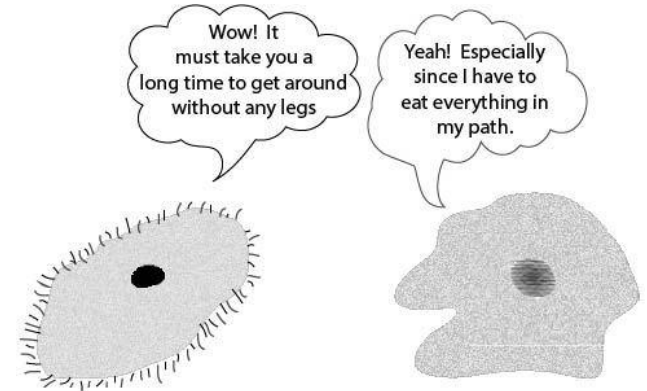


How?

Movement

- Organisms, cells, organelles
- Subjective recording

- Tracing
- Direction
- Velocity
- Diffusivity
- Association

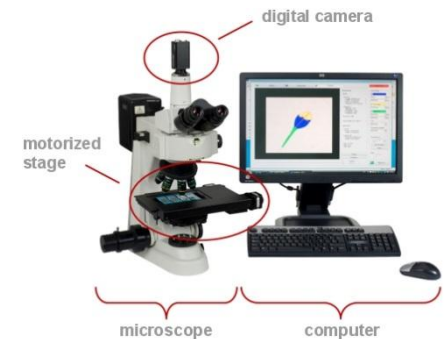


How?

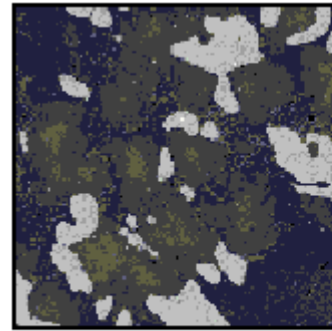
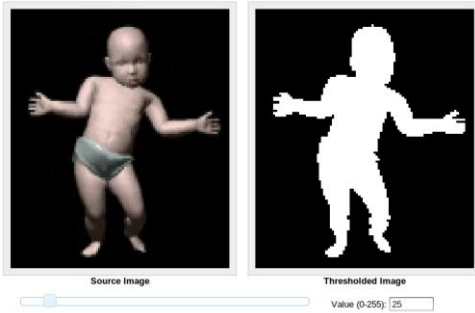
Image Analysis



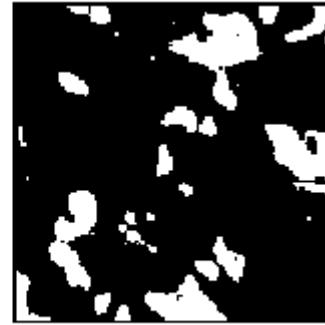
- Automated/Computerised Measurement vs Eye/Brain
- Early: 1969-1990 ... then software based
 - MOP, Quantimet, Magiscan, Imagan, **NIH Image-J**, Matlab, LAS, Image Pro, i-Solution, QuPath
 - **Recent (2010-now):** Machine Learning, AI
- Procedure
 - Sampling
 - Calibration
 - Image capture
 - **Segmentation**
 - Thresholding, Edge Detection, Erosion/Dilation
 - Object detection
 - Measurement
 - Size, shape, number, density (IOD?), arrangement, ...
 - Data analysis and display
- + / -
 - Speed & Measurement / Identification, User, Cost, GIGO



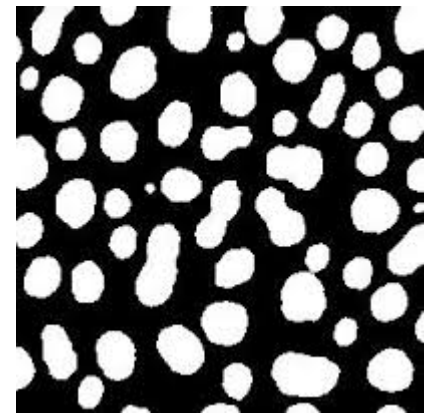
Thresholding



Original image



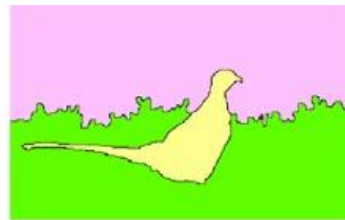
Thresholded binary image



Segmentation



Original Image



Human Seg



Machine Seg



Original Image

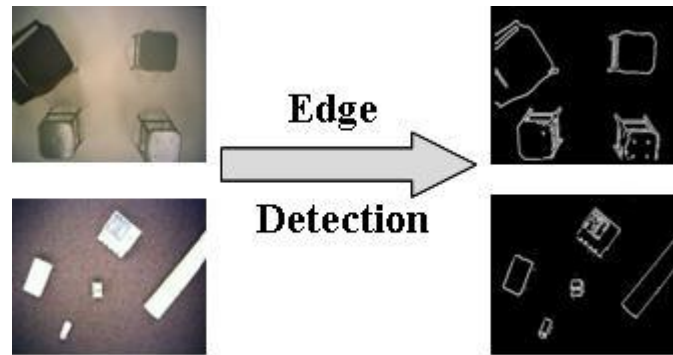


Human Seg

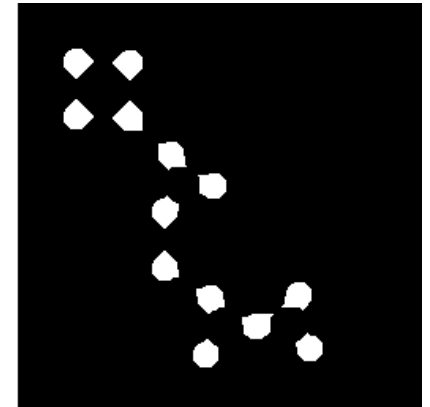
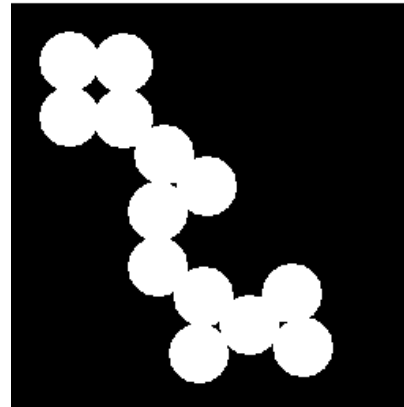


Machine Seg

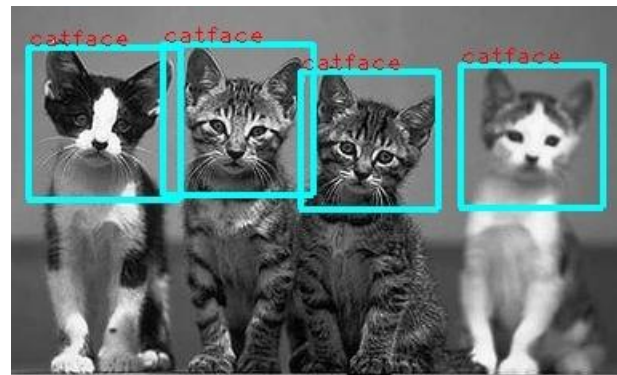
Edge Detection



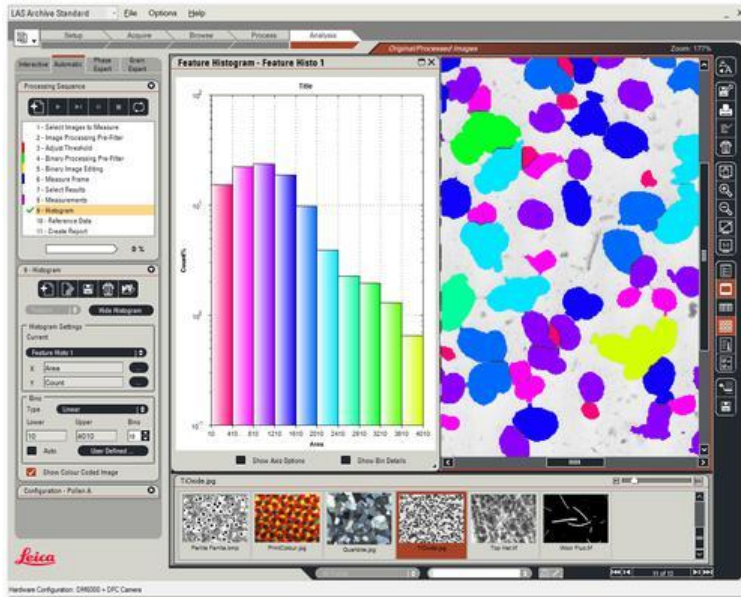
Erosion / Dilation



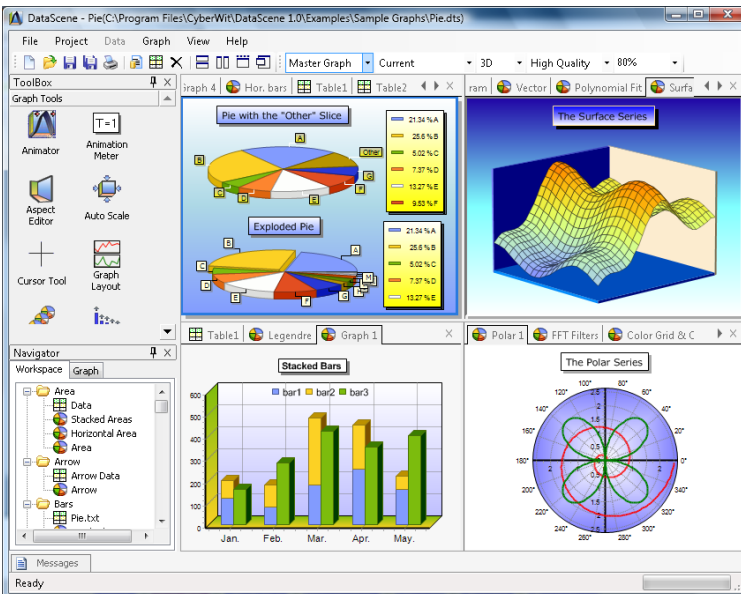
Object Detection



Problems – brilliant results output **But** does the user know what the machine is doing and have they considered bias, caveats etc. ?? !!



1980s advertisement ...



**THANKS TO MAGISCAN – IMAGE ANALYSIS IS NO LONGER
THE PRESERVE OF THE SPECIALIST**

Results ?

GIGO

Garbage In – Garbage Out !

“Garbage to ten decimal places is still garbage !”



Are the results unbiased, precise, accurate, valid, meaningful ?

Statistics !!!

- **Accuracy**
 - Degree of closeness to true value
- **Precision**
 - Related to reproducibility and repeatability
 - Improve by increasing sample size
- **Bias**
 - Random or Systematic error
- **Valid**
 - Measurement system which is **ACCURATE** and **PRECISE** and **UNBIASED**

Introduction - sources of variability in measurement

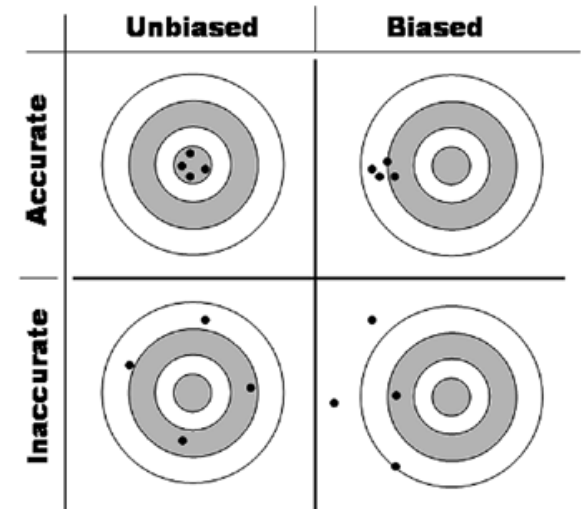
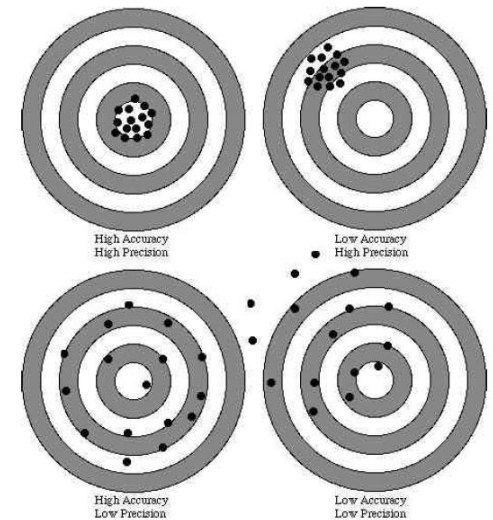


Fig. 2 Schematic illustration of the difference between accuracy and bias. The top row of targets shows accuracy that is the hits are closely clustered together. The bottom row shows inaccuracy and there is a marked scatter of hits. In the left hand column the average of the cluster of hits tends towards the bull s-eye, which means that they are unbiased. The right hand column shows the converse case, these hits are biased (based on Howard and Reed 1998).

How to conduct a measurement project



**Think
Do More
Less Well
Cheaply
Anywhere
Yourself
!!!**

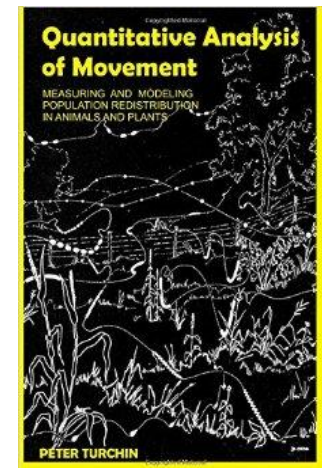
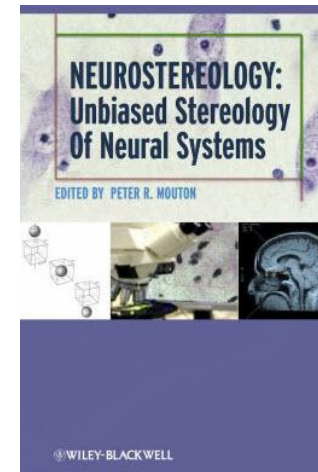
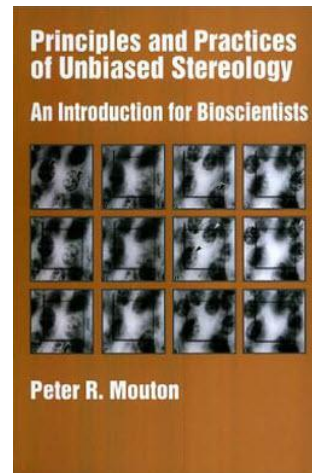
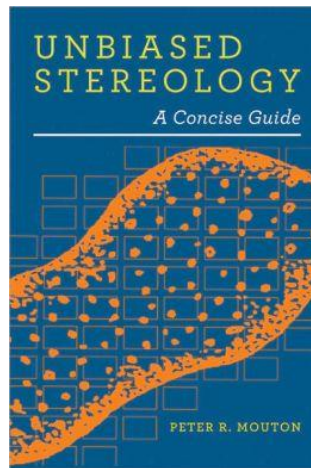
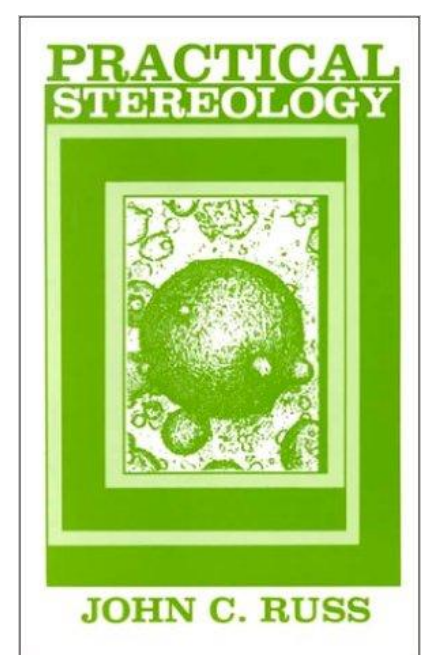
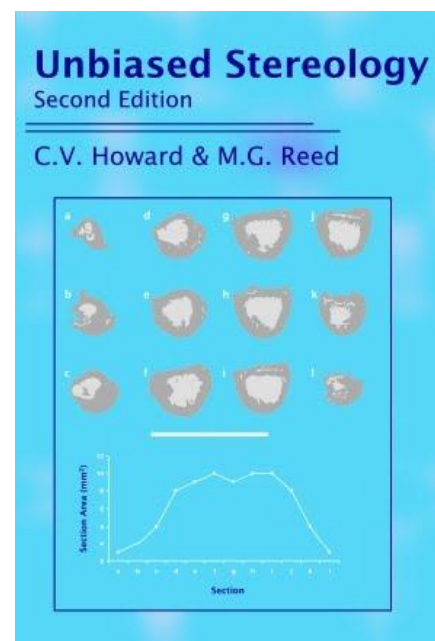
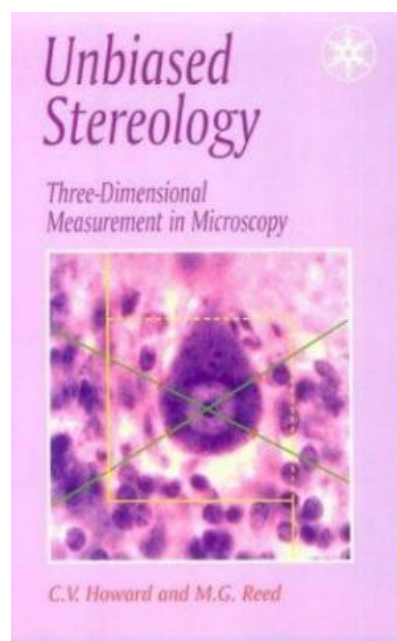
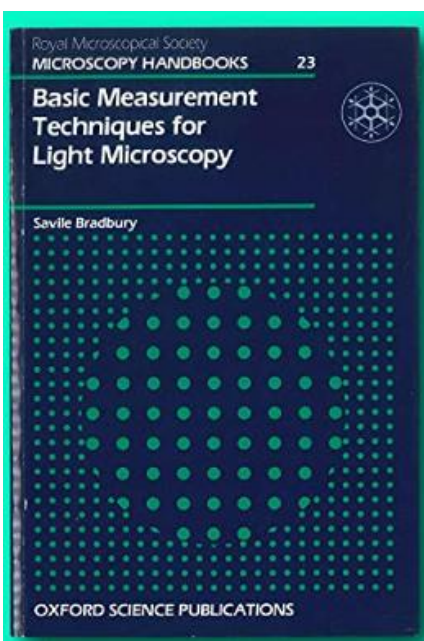
But 1st ...Guess !!

References

- **Aherne WA & Dunnill MS (1982) Morphometry, Arnold, London**
- **Bradbury S 1991 Basic Measurement Techniques for Light Microscopy RMS #23**
- **Howard & Reed 1998, 2004 Unbiased Stereology**
- [Russ, 2019 Robust Measurements](#)
- [Russ et al 2018 The Problem of Perimeter](#)
- www.stereology.info

- **Culley, S., Caballero, A. C., Burden, J. J., & Uhlmann, V. (2023). Made to measure: An introduction to quantifying microscopy data in the life sciences. Journal of Microscopy, 1–22. <https://doi.org/10.1111/jmi.1320>**

- **Russ JC (1990) Computer-Assisted Microscopy: The measurement and analysis of images. Plenum Press, New York.**
- **Steer MW (1981) Understanding Cell Structure. Cambridge University Press.**
- **Weibel ER (1963) Morphometry of the Human Lung. Springer-Verlag, Heidelberg**
- **Weibel ER (1979) Stereological methods. Vols 1 & 2. Academic Press, London Edinburgh.**
- **Weibel ER (1984) The pathway for oxygen. Structure and function in the mammalian respiratory system. Harvard University Press**
- **Williams MA (1977) Quantitative methods in biology (Practical methods in Electron Microscopy; 6 pt1 ed AM Glauert). North-Holland**
- **Gundersen HJG et al (1988) The new stereological tools: disector, fractionator and point-sampled intercepts and their use in pathological research. Acta Pathologica et Microbiologica Scandinavica, 96, 857-881.**
- **Howard CV, Reed MG (1998) Unbiased Stereology: Three-Dimensional Measurement in Microscopy. RMs Handbook 41. Bios Scientific**
- **Mayhew TM & Gundersen HJG (1996) 'If you assume, you can make an ass out of u and me': a decade of the disector for stereological counting of particles in 3D space. Journal of Anatomy, 188, 1-15.**
- **Diggle PJ (1983) Statistical Analysis of Spatial Point Patterns. Academic Press, London.**
- **Dormer KJ (1980) Fundamental Tissue Geometry for Biologists. Cambridge University Press, Cambridge.**
- **Ebdon D (1985) Statistics in Geography. 2nd ed. Blackwell, Oxford.**
- **Mahon M & Cumming WJK (1985) SPAM: Spatial pattern analysis of muscle in neuromuscular diseases. Neuropathology & Applied Neurobiology, 11, 74-75.**
- **Murray JD (1990) Mathematical Biology. Springer Verlag, Berlin.**
- **Unwin D (1981) Introductory Spatial Analysis. Methuen, London.**
- **MANDARIM-DE-LACERDA, C. A. & DEL SOL, M. Tips for studies with quantitative morphology (morphometry and stereology). Int. J. Morphol., 35(4):1482-1494, 2017.**
- **Hally, A. D. A counting method for measuring the volumes of tissue components in microscopical sections. J. Cell Sci., 105(S3): 503-17, 1964.**



<http://www.lab.anhb.uwa.edu.au/mb140/scope/stereology/stereology.htm>

http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0001-37652003000400006

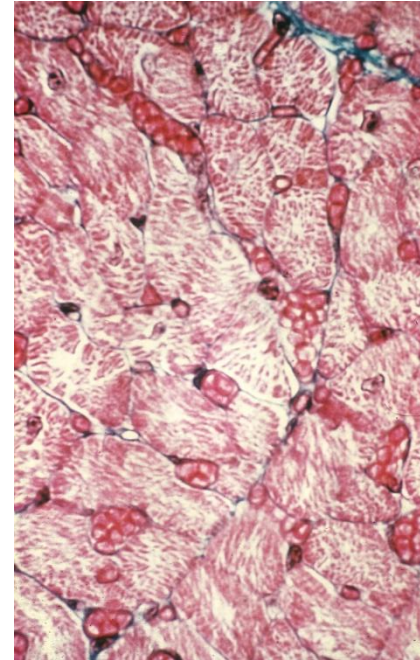
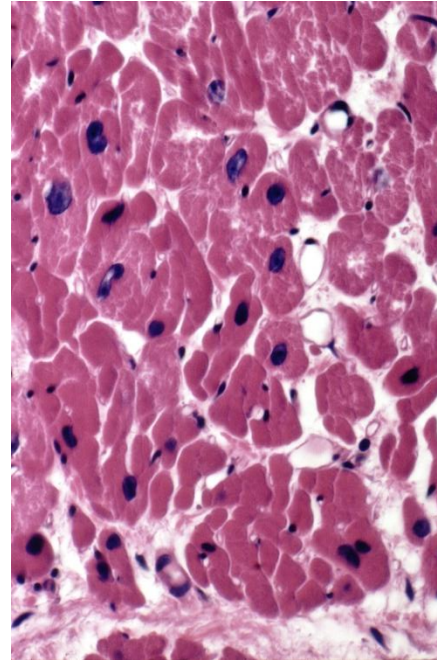
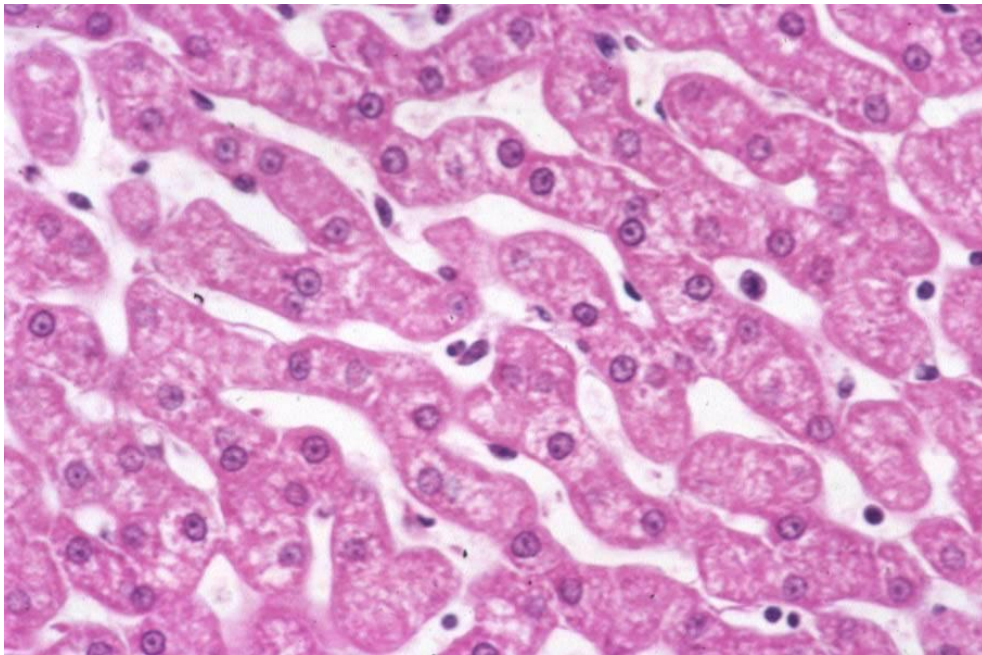
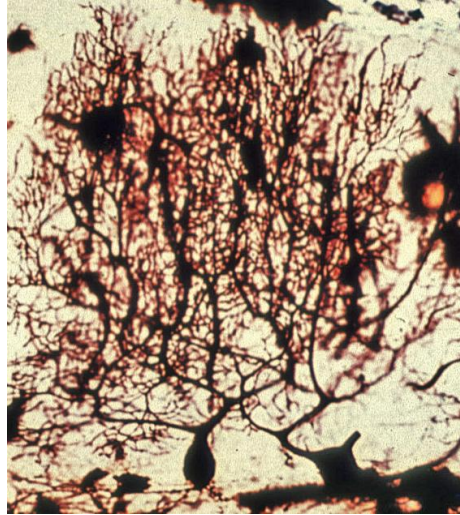
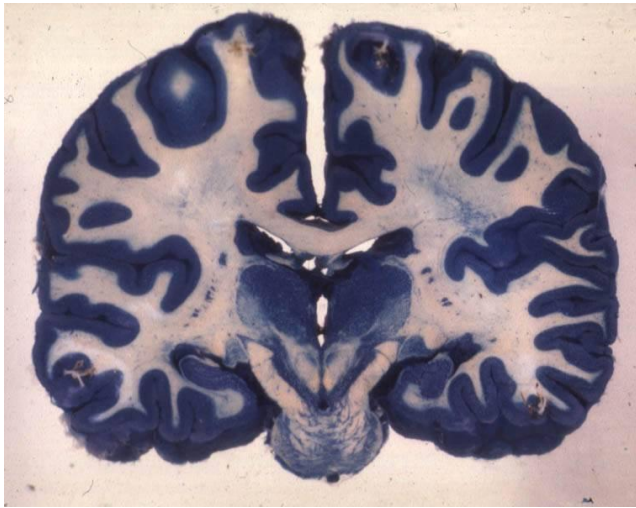
Question

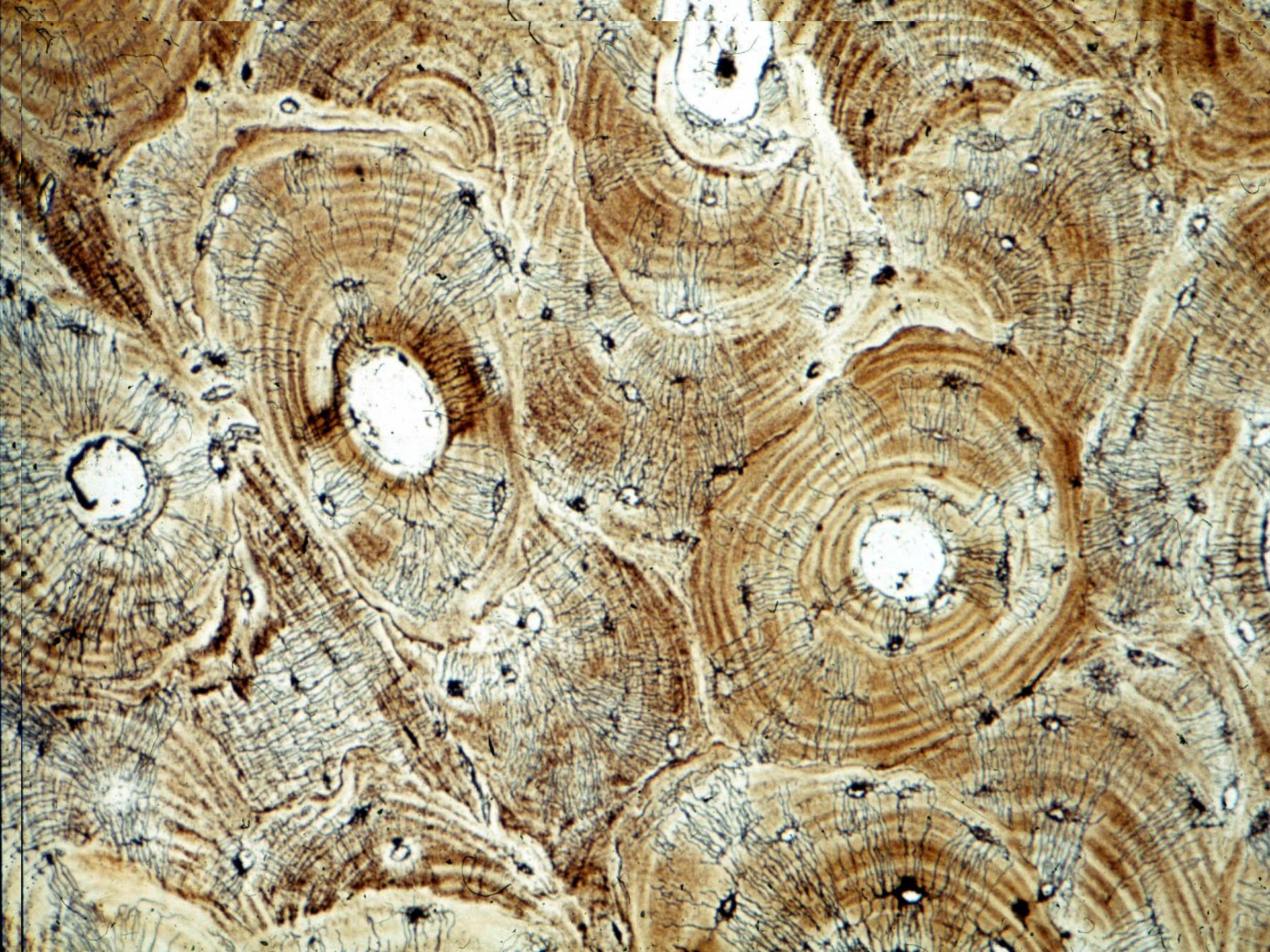
**What is the Answer to the Ultimate Question
of Life, the Universe & Everything ?**

= ??

Key Reference: Douglas Adams "Hitchhikers Guide to the Galaxy"

Volumes – Surface Areas – Lengths – Numbers - Branching ???





Practical

- 1. Magnification Calibration**
- 2. Morphometry**
3. Stereology
4. (Pattern/Shape analysis)